

Effect of microbial oil and fish oil on rumen fermentation and metabolism of fatty acids in artificial rumen

D. JALČ¹, M. ČERTÍK², K. KUNDRÍKOVÁ², P. KUBELKOVÁ³

¹Institute of Animal Physiology, Slovak Academy of Sciences, Košice, Slovak Republic

²Institute of Biotechnology and Food Sciences, Faculty of Chemical and Food Technology, Slovak University of Technology, Bratislava, Slovak Republic

³Research Institute of Animal Science, Prague-Uhřetěves, Czech Republic

ABSTRACT: The objective of this study was to examine the effect of microbial oil (MO, n-6 fatty acids) and fish oil (FO, n-3 fatty acids) used in their blends as supplements (5% wt/wt) to the diet containing 80% of hay and 20% of barley on rumen fermentation and lipid metabolism in artificial rumen. Overall, three different ratios of n-6 and n-3 fatty acids (1:1, 3:1, and 5:1) as the blends of MO and FO were used. Two similar consecutive experiments were carried out within 2 months. Each experiment lasted for 12 days with 6 days of stabilization period. The addition of all three oil blends did not affect the parameters of fermentation such as degradation of dry matter (DM), detergent fibre, total gas production, but increased the degradation of cellulose and hemicellulose in the diets. The supplementation of oil blends to the diet insignificantly (NS) decreased the methane production (mostly the n-6/n-3 ratio 1:1, about 23.5%), increased ($P < 0.01$) mol% of propionate (mostly the n-6/n-3 ratio 1:1, about 24.1%) and decreased ($P < 0.05$) mol% of acetate (mostly the n-6/n-3 ratio, 1:1, about 7.7%). The lipid metabolism in artificial rumen was also affected, when the oil blends increased ($P < 0.001$) the concentration of total fatty acids (FA) and long-chain FA (LCFA) in effluent. The concentration (mg/g rumen fluid DM) of *trans* (*trans* 11 C_{18:1}, TVA-vaccenic acid), *cis* C_{18:1} isomers and CLA-conjugated linoleic acid (*cis* 9, *trans* 11 C_{18:2}) was also increased ($P < 0.001$) by the oil blends. Finally, the oil blends caused the incomplete FA biohydrogenation by an increase in TVA concentration and TVA/C_{18:0} ratio in effluent in artificial rumen.

Keywords: microbial oil; fish oil; rumen fermentation; lipid metabolism

Fat supplements are included in the diet of ruminants to increase energy density, improve nutrient utilization, enhance milk and meat yields and affect fatty acid composition (Bauman et al., 2003). The type of diet fed to ruminants influences rumen

fermentation and fatty acid profile formed during biohydrogenation (BH). Ruminant diets are usually composed of plants that are rich in polyunsaturated fatty acids (PUFA), e.g. linoleic acid (LA, C_{18:2}) and α -linolenic acid (LNA, C_{18:3}). Fish oil supplements

Supported by the Grant Agency of Ministry of Education of the Slovak Republic and Slovak Academy of Sciences of the Slovak Republic (Grants Nos. 2/6174/06, 1/0747/08), Slovak Research and Development Agency of the Slovak Republic (No. APVV-0043-07) and Ministry of Agriculture of the Czech Republic (Grant MZE No. 000 270 1404).

to the ration as an additional source of energy introduce the long-chain fatty acids, EPA (eicosapentaenoic acid, $C_{20:5}$) and DHA (docosahexaenoic acid, $C_{22:6}$) as the predominant FA. Microbial oil isolated from oleaginous microorganisms is another source of various polyunsaturated fatty acids including gamma-linolenic acid (GLA, $C_{18:3}$), dihomogamma-linolenic acid (DGLA, $C_{20:3}$), arachidonic acid (AA, $C_{20:4}$) as well as EPA and DHA (Čertík and Shimizu, 1999; Papanikolaou et al., 2004). Microbial oil together with monensin and fumarate was used also in our previous experiments (Jalč and Čertík, 2005). PUFA can be divided into two categories according to the occurrence of double bonds in the fatty acyl chain: n-3 (omega-3) and n-6 (omega-6) fatty acids. Omega-3 FA's include LNA, EPA, and DHA. Omega-6 FA's include LA, GLA, DGLA and AA. It is known that dietary lipids undergo two important transformations in the rumen of ruminants. Rumen metabolism of dietary FA is initiated by microbial lipolysis and subsequent biohydrogenation of free PUFA. During this process, the concentrations of C_{18} PUFA, such as LNA and LA, decrease as they are biohydrogenated completely to stearic acid ($C_{18:0}$) with the formation of intermediates like CLA (*cis* 9, *trans* 11 $C_{18:2}$) and TVA (*trans* 11 $C_{18:1}$) as the most important known ones (Hartfoot and Hazlewood, 1997). Amounts of biohydrogenation intermediates produced in the rumen influence their concentrations in tissues or milk (Lor et al., 2003). TVA and CLA in meat and milk are examples of hydrogenation intermediates that may have beneficial implications in human health. The n-6 and n-3 ratio of fatty acids is highly influenced by the fatty acid composition of the diet fed to the ruminants and affects the concentration of CLA and TVA in the rumen, milk and meat. It is widely accepted that the ideal intake of omega-6 FA's should not exceed the intake of omega-3 FA's more than 4–5 times (Raes et al., 2004). A continuous culture fermenter (artificial rumen) was used to characterize the effect of microbial oil and fish oil blends as the supplements to the diet containing 80% of hay and 20% of barley: (a) on rumen fermentation (degradation of dry matter and detergent fibre, methane production, volatile fatty acid production), (b) on fatty acid metabolism (outputs of *cis* and *trans* isomers of $C_{18:1}$ and $C_{18:2}$ – CLA, TVA during incubation) in this study. Different ratios of n-6 and n-3 FA's (5:1, 3:1, and 1:1) prepared as the mixtures of selected microbial oil and fish oil were used.

MATERIAL AND METHODS

Oil supplements

Microbial oil was isolated from the lower fungus *Thamnidium elegans* CCF 1465 (Culture Collection of Fungi, Department of Botany, Charles University, Prague, Czech Republic). The fungus was maintained on modified Czapek-Dox agar slants with yeast extract (2.5 g/l) at 4°C. The pre-cultivation flasks (100 ml) containing 30 ml cultivation medium (glucose, 30 g/l, corn steep, 10 g/l) were inoculated with the fungal spore suspension in an isotonic solution at a final concentration of $1-2 \times 10^6$ spores per ml. After 2 days of pre-incubation on a rotary shaker (reciprocal speed of 130 rpm) at 25°C, 60 ml of pre-incubated culture was transferred to 2 000 ml Erlenmeyer flasks (equipped with baffles to improve aeration) containing 1 000 ml of cultivation medium. The culture was incubated on a rotary shaker (120 rpm) for 5 days at 25°C. After fermentation the mycelium was harvested by filtration, washed with water and gently dried at 65°C for 10 h. Dry fungal biomass was crushed mechanically and total microbial lipid was extracted with hexane with Soxhlet apparatus for 2 h (Čertík and Horenitzký, 1999). Hexane was finally evaporated under vacuum and microbial oil was used for further studies. Fish oil as cod liver oil was obtained from commercial sources. The fatty acid composition (expressed as % of fatty acid methyl esters – FAME) of feed ingredients, meadow hay, barley, MO and FO, is presented in Table 1. The oils were used as blends of MO and FO and the total 5% supplementation of the oil blends was applied in this experiment. Overall, three different ratios of n-6 and n-3 FA (5:1, 3:1, and 1:1) as blends of MO and FO were used.

In vitro fermentation system

The study was carried out using an artificial rumen as described by Czerkawski and Breckenridge (1977). The complete unit composed of four vessels (V_1 , V_2 , V_3 and V_4), each 850 ml in volume. The general incubation period was described by Czerkawski and Breckenridge (1977). The vessel inoculum was obtained from three ruminally cannulated Slovak Merino sheep (mean body weight 42 ± 2.1 kg) fed 960 g of dry matter (DM) of meadow hay and 240 g DM of crushed barley

Table 1. The fatty acid composition (%) of feed ingredients

(%)	Meadow hay	Barley	Microbial oil	Fish oil
C _{10:0}	0.20	–	–	–
C _{12:0}	0.60	–	–	–
C _{14:0}	1.20	0.30	0.60	4.70
C _{15:0}	0.40	0.10	0.10	0.50
C _{16:0}	21.0	18.0	16.10	12.40
C _{16:1}	2.40	0.20	0.80	6.40
C _{17:0}	0.40	0.10	–	0.30
C _{18:0}	2.50	1.70	7.20	2.50
C _{18:1'} <i>cis</i> 9	8.40	16.50	50.20	16.30
C _{18:1'} <i>cis</i> 7	0.50	0.70	–	–
C _{18:1'} <i>cis</i> 11	–	–	0.40	3.10
C _{18:1'} <i>trans</i> 9	–	–	0.10	–
C _{18:1'} <i>trans</i> 11	–	–	–	0.90
C _{18:2}	20.30	54.50	12.60	5.30
C _{18:3} (LNA)	27.70	5.00	–	1.60
C _{18:3} (GLA)	–	–	7.80	–
C _{20:0}	1.20	0.20	0.30	–
C _{20:1}	–	–	–	8.20
C _{20:5}	–	–	–	6.80
C _{22:5}	–	–	–	2.90
C _{22:6}	–	–	–	9.60
C _{22:0}	3.90	0.40	0.50	–
C _{23:0}	0.60	0.10	0.10	–
C _{24:0}	1.70	0.20	1.40	–
Others	7.00	2.00	1.80	18.30

(%) FAME, fatty acid methyl esters; LNA, α linolenic acid; GLA, γ linolenic acid

in two equal meals. The chemical composition of meadow hay and barley was as follows: DM, 93.44 (89.69); nitrogen, 1.17 (2.18); ash, 9.47 (3.69); neutral detergent fibre (NDF), 58.27 (26.32); acid detergent fibre (ADF), 37.19 (6.79); hemicellulose, 21.07 (19.49); cellulose, 29.50 (5.41); lignin, 7.68 (7.72) as % of initial DM. Fermentation inocula (solid and liquid ones) were collected through the rumen cannula immediately before the morning feeding and transferred to the artificial rumen. The solid digesta (80–100 g of wet weight) were

placed into nylon bags (100 μ m pore size) in each of the four fermentation vessels. The vessels were filled to overflowing with strained rumen fluid and artificial saliva (1:1) (McDougall, 1948). Including the first day of the experiment, the vessels were supplied with 12.8 g (11.96 g DM) of meadow hay and 3.2 g (2.87 g DM) of barley at the daily intervals. Vessel V₂ received also 5% (wt/wt) of the oil blends (MO + FO) with the n-6 to n-3 FA ratio 1:1; V₃ received 5% (wt/wt) of the oil blends (MO + FO) with the n-6 to n-3 FA ratio 3:1 and V₄

received 5% (wt/wt) of the oil blends (MO + FO) with the n-6 to n-3 FA ratio 5:1. Vessel V₁ was the control (without oil addition). To ensure that all vessels contained 12% crude protein (CP), 223 mg of urea were added in 1 000 ml McDougall buffer. A continual infusion of artificial saliva (pH 8.4) at the rate 665–728 ml was maintained through each vessel during the experiment.

Measurements and chemical analyses

Two similar consecutive experiments were carried out in artificial rumen within 2 months. Each experiment in artificial rumen lasted 12 days. To ensure a steady state within the vessels a 6-day adaptation period was followed by a 6-day collection period. On days 6–12 the following samples were collected: produced gas was collected into special bags and volumes of gas were measured with a gas-meter and methane concentrations were analysed in a gas chromatograph (Perkin-Elmer, Clarus 500) as reported by Czerkawski and Clapperton (1968). Liquid effluent was collected into flasks placed in ice bath and samples were taken for volatile fatty acids (VFA), ammonia nitrogen (NH₃-N) and fatty acid (FA) analyses. The daily productions of VFA were analysed by the gas chromatography procedure (Cottyn and Boucque, 1968). Ammonia nitrogen concentrations were measured by a microdiffusion method (Conway, 1962). The fatty acid content in effluent was determined in lyophilized samples. Lipids were extracted from 500 mg of freeze-dried effluent, meadow hay and barley using chloroform and methanol (2:1, vol/vol) followed by 6N HCL as described by Fellner et al. (1995). Heptadecenoate C_{17:0} (Supelco, USA) was used as the internal standard. Fatty acids of total lipids were analysed as their methyl esters (Christoperson and Glass, 1969) by gas chromatography according to Čertík et al. (2005). Dry matter, ash and nitrogen were analysed according to the methods of the Association of Official Analytical Chemists (AOAC, 1980). Neutral detergent fibre, acid detergent fibre, cellulose and hemicellulose in feed and residual feed samples were analysed by the method of Goering and Van Soest (1970). The other fermentation variables, energetic efficiency of VFA (Ørskov et al., 1968) and OMF, organic matter fermented (Demeyer and Van Nevel, 1979), were calculated from the stoichiometry of rumen fermentation.

Statistical analysis

Means of results from treatments were compared by one-way analysis of variance (Graphpad InStat, Graphpad Software Inc., San Diego, CA USA). Treatment means were statistically compared by the Turkey-Kramer multiple comparison test. The tables give the group means and the standard error of the mean (\pm SEM). Probability values of $P < 0.05$ were considered as significant.

RESULTS AND DISCUSSION

Effect of modifying n-6/n-3 ratio of dietary oil supplements on rumen fermentation *in vitro*

Rumen metabolism can be characterized by a fermentation pattern, consisting of the following parameters: the amount of molar proportions of volatile fatty acids produced; the amount of methane formed and OMF (Demeyer and Van Nevel, 1986). All these parameters of rumen fermentation were determined in this study. The n-6/n-3 ratio is highly influenced by the fatty acid composition of the diet fed to the animals. Finishing ruminants on pasture can decrease the n-6/n-3 ratio to a value of 2 or less, while concentrate fed ruminants had the ratios around 6–10 (Raes et al., 2004). In our study, the effect of modifying the n-6/n-3 ratios of dietary oil supplements to the diet consisting of hay and barley (80:20%) on the parameters of rumen fermentation in artificial rumen was studied. Fish oil contained higher amounts of EPA (6.8% FAME) and DHA (9.56% FAME) and lower amounts of LNA (1.63% FAME) as the sources of n-3 FA (Table 1). Microbial oil contained higher amounts of LA (12.63% FAME) and lower amounts of GLA (7.83% FAME) as the sources of n-6 FA (Table 1). Microbial oil is an alternative source of animal and plant oils (Čertík and Shimizu, 1999; Ratledge, 2003). In our study, the addition of oil blends up to 5% in DM at different ratios of n-6/n-3 FA to hay-barley diet showed a slight effect on some parameters of rumen fermentation *in vitro* (Table 2). The degradation of DM, NDF, ADF was not affected by the supplementation of oil blends. We could only determine the higher ($P < 0.05$) values of hemicellulose in fermentation vessels V₂, V₃, V₄ and cellulose in V₂ and V₄. Some papers reported no negative effects of supplemental saturated or unsaturated fat (Schroeder

Table 2. The effect of microbial and fish oil blends as supplements in the diet containing hay and barley (80:20%) on rumen fermentation in artificial rumen (n-12)

Fermentation vessel	Control	MO + FO blend	MO + FO blend	MO + FO blend
	V ₁	V ₂	V ₃	V ₄
n-6:n-3 ratio	–	1:1	3:1	5:1
DMD (%)	49.30 ± 1.0	46.90 ± 0.9	48.90 ± 1.7	48.80 ± 0.7
NDF (%)	18.70 ± 1.7	19.70 ± 1.3	23.80 ± 2.6	25.10 ± 1.0*
ADF (%)	19.90 ± 1.7	18.70 ± 1.4	25.30 ± 2.5	19.50 ± 1.2
Hemicelluloses (%)	16.30 ± 1.7	20.80 ± 1.3*	21.40 ± 2.7*	33.20 ± 0.9***
Cellulose (%)	29.30 ± 1.4	38.70 ± 1.1*	34.90 ± 2.2	41.20 ± 0.8**
VFA (mmol/day)	36.20 ± 0.9	35.50 ± 1.0	37.90 ± 1.0	36.60 ± 0.9
Acetate (mol%)	59.70 ± 0.2	55.10 ± 0.2*	58.0 ± 0.3*	59.0 ± 0.2*
Propionate (mol%)	19.50 ± 0.2	24.20 ± 0.3**	21.20 ± 0.3*	21.20 ± 0.3*
n-butyrate (mol%)	14.90 ± 0.3	14.60 ± 0.3	15.10 ± 0.4	14.80 ± 0.4
Total gas (l/day)	3.30 ± 0.1	3.20 ± 0.1	3.20 ± 0.1	3.40 ± 0.1
Methane (mol/day)	6.80 ± 0.4	5.20 ± 0.5	6.40 ± 0.3	6.80 ± 0.3
NH ₃ -N (mg/100ml)	12.40 ± 0.6	14.40 ± 1.0	18.30 ± 1.0**	17.10 ± 1.2*
E (%)	74.90 ± 0.1	76.90 ± 0.1**	75.70 ± 0.1*	75.50 ± 0.1*
OMF (g/day)	6.70 ± 0.1	6.30 ± 0.1	6.60 ± 0.1	6.60 ± 0.1

MO = microbial oil; FO = fish oil; DMD = dry matter digestibility; E = energetic efficiency of VFA's; OMF = organic matter fermented; ±SEM = standard error of the mean; * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$ differences from control

et al., 2002) or linseed, soybean and cottonseed oil supplements (4% wt/wt) with the dietary n-6/n-3 fatty acid ratio of 2.3:1, 8.8:1, 12.8:1 and 15.6:1, respectively, on apparent digestibility of DM, NDF or ADF (Kim et al., 2007). On the contrary, Choi et al. (1998) reported a non-significant trend towards on increase in fibre digestion in the rumen with the addition of fish oil. Our experiments showed that total gas production (3.2–3.3 l/day) was similar in all fermentation vessels. The methane production (mmol/day) was numerically (NS) reduced in V₂ (about 23.5%) and in V₃ (about 5.9%). There were not any different values in the methane production in V₄ compared to the control (Table 2). Our previous *in vitro* study with MO, borage oil (BO) and evening primrose oil (EPO) supplementation (5% wt/wt) of the diet consisting of hay and barley (60:40%) showed numerically decreased methane production by about 11.3% (MO), 2.04% (BO) and 11.4% (EPO, Jalč et al., 2005). Methane suppression with oil blend supplementation was accompanied by a shift of the fermentation pattern towards pro-

pionate without any effect on total VFA production (Table 2). Molar proportions of propionate were increased ($P < 0.01$) in V₂ (about 4.7 units), in V₃ and V₄ ($P < 0.05$; about 1.7 units). Molar proportions of acetate were decreased ($P < 0.05$) in V₂ (about 4.6 units), in V₃ and V₄ ($P < 0.05$; about 0.7 to 1.7 units), while molar proportions of n-butyrate were not affected by dietary treatment with the oil blend (Table 2). Kim et al. (2005) found that the oil supplementation containing the n-6/n-3 ratios of 2:1, 10:1, 16:1, and 20:1 by mixing linseed oil, cottonseed oil and soybean oil did not affect the ruminal concentrations of acetate and propionate in lambs. The NH₃-N pool produced by degradation of urea in artificial saliva (McDougall buffer) and feed nitrogen are the main sources of nitrogen used by bacteria for protein synthesis. Ammonia nitrogen concentrations depend on ammonia use and release by the microbial population (Mansfield et al., 1995). The results showed that the supplementation of oil blends with the n-6/n-3 ratio 3:1 and 5:1 in V₃ ($P < 0.01$) and V₄ ($P < 0.05$)

Table 3. The effect of microbial and fish oil blends as supplements in the diet containing hay and barley (80:20%) on lipid metabolism in artificial rumen (n-12)

Fatty acid, (mg/g) rumen fluid DM	Control	MO + FO	MO + FO	MO + FO	Pooled SEM
Vessel	V ₁	V ₂	V ₃	V ₄	
n-6/n-3		1:1	3:1	5:1	
C _{14:0}	0.11	0.35***	0.38***	0.26**	0.02
C _{15:0}	0.24	0.45***	0.36	0.25	0.02
C _{16:0}	1.03	2.36***	2.32***	2.14***	0.05
C _{16:1} , <i>cis</i> 9	0.22	0.55***	0.58***	0.54***	0.02
C _{17:0}	5.21	5.06	5.25	5.22	0.13
C _{18:0}	0.82	0.62*	0.83	0.94	0.06
TVA	0.15	0.69***	0.78***	0.82***	0.01
C _{18:1} , <i>cis</i> 9	0.22	0.86***	0.88***	0.67***	0.02
C _{18:1} , <i>cis</i> 11	0.13	0.67***	0.69***	0.82***	0.03
C _{18:2} , <i>cis</i> 9, 12	0.21	0.22	0.18	0.17	0.01
CLA	0.13	0.24***	0.26***	0.32***	0.01
Total FA	8.22	11.20***	11.60***	11.50***	0.22
MCFA (%)	83.20	75.70*	73.80**	71.90***	2.12
LCFA (%)	16.80	24.30***	26.10***	28.0***	1.40
SFA (%)	90.50	76.0***	75.80***	75.20***	2.10
UFA (%)	9.40	30.70***	29.20***	32.10***	0.60
SFA/UFA	9.60	2.50***	2.60***	2.30***	0.50

MCFA = medium-chain fatty acids (C_{14:0}–C_{17:0}); LCFA = long-chain fatty acids > C_{18:0}; SFA = saturated fatty acids; UFA = unsaturated fatty acids; TVA, *trans* 11 C_{18:1}; CLA, *cis* 9, *trans* 11 C_{18:2}; **P* < 0.05; ***P* < 0.01; ****P* < 0.001 differences from control

increased the NH₃-N concentration in effluent, while its concentration in V₂ was not affected by the supplementation of oil blends with the n-6/n-3 ratio 1:1 (Table 2). In the other experiment, the supplementation of oil blends (5% wt/wt) – linseed oil (LO), rapeseed oil (RO) and FO as LO + RO, LO + FO, and LO + RO + FO to the diet consisting of 60% fresh lucerne and 40% maize caused an increase in the ammonia nitrogen concentration in effluent (Jalč et al., 2006). The energetic efficiency of VFA (E) was increased (*P* < 0.01–0.05) by the oil blends, mainly by the oil blends of the n-6/n-3 ratio 1:1. This was evoked by an increase in mol% of propionate in the diets after the oil blend treatment. The amount of OMF was unchanged by

the oil blend supplementation to the mixed diet in artificial rumen.

Effect of modifying n-6/n-3 ratio of dietary oil supplements on lipid metabolism *in vitro*

The lipid composition of meadow hay mostly consists of glycolipids and phospholipids, and major unsaturated fatty acids are LNA and LA (Table 1). The lipid composition of barley consists of triglycerides and major FA's are oleic acid (C_{18:1}) and LA (Table 1). The studied microbial oil was rich in C_{18:1} and GLA, and fish oil contained higher levels of EPA and DHA (Table 1). Therefore, the elevated

ratio of n-6/n-3 FA's in the oil mixtures (1:1 > 3:1 > 5:1) was a consequence of increased amounts of MO to FO used for the oil mixtures. The rumen microbial lipid metabolism is characterized by lipolysis of dietary glycolipids, phospholipids and triglycerides and leads to a release of free fatty acids. The formed free FA's are hydrogenated by rumen microbes to saturated end products such as stearic acid with the formation of intermediates like CLA and TVA (Hartfoot and Hazlewood, 1997). Overall, the addition of oil blends (n-6/n-3 1:1, 3:1, 5:1) to the feed resulted in the increased total fatty acid concentration in rumen fluid effluent (mg/g rumen fluid DM) by 36%, 41%, and 40%, respectively (Table 3). An increase in the total fatty acid concentration in rumen fluid is a typical response to the lipid supplementation of ruminant diets (Beaulieu et al., 2002). The supplementation of oil blends to the diet significantly decreased the percent proportion (%) of MCFA ($C_{14:0}$ – $C_{17:0}$, about 7.4–11.2%) and increased ($P < 0.001$) the amount of LCFA ($> C_{18:0}$, about 7.5–11.6%) in effluent. The degree of BH of individual fatty acids was not estimated in this study; however many authors have reported different rates of BH of $C_{18:1}$, $C_{18:2}$ and $C_{18:3}$ FA. Wachira et al. (2000) reported that BH of $C_{18:2}$ and $C_{18:3}$ ranged between 80 and 93% when lambs were fed fish oil. Biohydrogenation of EPA and DHA increased from 49 and 74% to 79 and 86%, respectively, when fish oil in the concentrate increased from 1 to 4 g per 100 g (Lee et al., 2005). It is known that BH of PUFA is characterized by: (a) changes in the percent proportions of unsaturated (UFA) and saturated (SFA) fatty acids in the rumen, (b) accumulation of *trans* fatty acids in the rumen, (c) higher concentration of stearic acid (AbuGhazaleh et al., 2002). We can state that the extent of BH of unsaturated FA in our experiment was affected by the oil blends and was characterized by: (a) accumulation of *trans* fatty acids, especially TVA in effluent, (b) changes in % proportion of SFA and UFA in effluent. Indeed, UFA concentration (%) increased ($P < 0.001$; about 20–23%, mainly the n-6/n-3 ratio 3:1) and SFA concentration (%) decreased ($P < 0.001$; about 14–15%, mainly the n-6/n-3 ratio 3:1) when oil blends were supplied in comparison with the control. Thus, the diet enriched with oil mixtures resulted in a decrease ($P < 0.001$) in the ratio of SFA/UFA (about 4 times) compared to the control. However, the concentration of stearic acid was slightly ($P < 0.05$) or not significantly decreased in the diets supplement-

ed with oil mixtures (n-6/n-3 ratio, 1:1, 3:1) or slightly (NS) increased with oil blends (n-6/n-3, ratio 5:1). AbuGhazaleh et al. (2002) also found a lower proportion of $C_{18:0}$ in ruminal digesta when 2% menhaden oil or 1% menhaden oil plus 1% extruded soybeans were added to TMR (total mixed ration) for cows. The accumulation of *trans* intermediates is probably due to an excess of free fatty acids, which inhibits the final hydrogenation of $C_{18:1}$ *trans* isomers to stearic acid (Gulati et al., 2000). In our experiment, the concentration of the main *trans* $C_{18:1}$ isomer – TVA in effluent increased ($P < 0.001$; 4.6–5.2 times) with all three oil blends. This increase in TVA may be caused by the inhibition of reductase activity of ruminal microbes with DHA and EPA present in fish oil. AbuGhazaleh and Jenkins (2004) observed that the addition of DHA, soybean oil or their mix to ruminal cultures *in vitro* increased *trans* $C_{18:1}$ isomers by 141, 100 and 266%, respectively, compared with the control. The concentration of *cis* $C_{18:1}$ isomers (*cis* 9 and *cis* 11) increased ($P < 0.001$) in diets supplemented with oil blends 3.9 or 5.1; 4 or 5.3; and 3 or 6.3 times (n-6/n-3 ratio, 1:1, 3:1, 5:1). Mosley et al. (2002) found that *cis* 9 $C_{18:1}$ isomer could serve as a precursor for several *trans* FA isomers. The *cis* 9 $C_{18:1}$ might also interfere with BH of other PUFA in the diet, resulting in the accumulation of *trans* $C_{18:1}$ isomers. The increase in TVA concentration and in TVA/ $C_{18:0}$ ratio from 0.18 (control) to 1.11, 0.94, 0.87 (n-6/n-3 ratio, 1:1, 3:1, 5:1), respectively, is an indication of the incomplete biohydrogenation of unsaturated FA with fat supplements (Table 3). The concentration of the other main isomer (CLA, *cis* 9, *trans* 11 $C_{18:2}$) increased ($P < 0.001$) as the n-6/n-3 ratio increased (Table 3). Similar results were reported by Váradyová et al. (2007), who studied the effect of sunflower oil and rapeseed oil (5% wt/wt) supplement to meadow hay-barley grain (80:20%) diet in sheep on the profile of fatty acids and their isomers (CLA, TVA) in rumen fluid. On the contrary, Kim et al. (2005) found that the CLA concentration decreased as the n-6/n-3 ratio increased. These authors investigated the effect of modifying the n-6/n-3 ratio of dietary oil supplement treatments of 2:1, 10:1, 16:1 and 20:1 by mixing linseed oil, cottonseed oil and soybean oil in lambs.

It can be stated that the supplementation of oil blends with the n-6/n-3 ratio 1:1, 3:1, 5:1 to a mixed diet (hay-barley, 80:20%): (a) increased the degradation of hemicellulose and cellulose in the diets; (b) numerically (NS) decreased the methane pro-

duction (mostly the n-6/n-3 ratio 1:1, about 23.5%), increased ($P < 0.01$) mol% of propionate (mostly the n-6/n-3 ratio 1:1, about 4.7 units) and decreased ($P < 0.05$) mol% of acetate (mostly the n-6/n-3 ratio 1:1, about 4.6 units); (c) increased ($P < 0.001$) the concentration of total FA and % proportion of LCFA in effluent; (d) increased ($P < 0.001$) the production of *trans* (*trans* 11 C_{18:1}, TVA) and *cis* (*cis* 9, *cis* 11) C_{18:1} isomers; (e) increased ($P < 0.001$) the production of *cis* 9, *trans* 11 C_{18:2} (CLA). Finally, the oil mixture supplementation caused the incomplete biohydrogenation of fatty acids characterized by an increase in TVA concentration and TVA/C_{18:0} ratio in effluent.

REFERENCES

- AbuGhazaleh A.A., Jenkins T.C. (2004): Disappearance of docohexaenoic and eicosapentaenoic acids from cultures of mixed ruminal microorganisms. *Journal of Dairy Science*, 87, 645–651.
- AbuGhazaleh A.A., Shingoethe D.J., Hippen A.R., Kalscheur K.F., Whitlock I.A. (2002): Fatty acid profiles of milk and rumen digesta from cows fed with fish oil, extruded soybeans or their blend. *Journal of Dairy Science*, 85, 2266–2276.
- AOAC (1980): Official Methods of Analysis of the Association of Official Agricultural Chemists, 13th ed., AOAC, Washington, USA.
- Bauman D.E., Corl B.A., Peterson D.G. (2003): The biology of conjugated linoleic acid in ruminants. In: Sebedio J.L., Christie W.W., Adlof R. (eds.): *Advances in Conjugated Linoleic Acid Research*, AOCS Press, Champaign, IL, USA, 146–173.
- Beaulieu A.D., Drackley J.K., Merchen N.R. (2002): Concentrations of conjugated linoleic acid (*cis* 9, *trans* 11-octadecadienoic acid) are not increased in tissue lipids of cattle fed a high concentrate diet supplemented with soybean oil. *Journal of Animal Science*, 80, 847–856.
- Čertík M., Horenitzký R. (1999): Supercritical CO₂ extraction of fungal oil containing γ -linolenic acid. *Biotechnology Techniques*, 13, 11–15.
- Čertík M., Shimizu S. (1999): Production and application of single cell oils. *Agro Food Industry Hi-Technology*, 10, 26–32.
- Čertík M., Breierová E., Jursíková P. (2005): Effect of cadmium on lipid composition of *Aureobasidium pullulans* grown under addition of extracellular polysaccharides. *International Biodeterioration Biodegradation*, 55, 195–202.
- Christoperson S.W., Glass R.L. (1969): Preparation of milk fat methyl esters by alcoholysis in an essentially nonalcoholic solution. *Journal of Dairy Science*, 52, 1289–1290.
- Conway E.J. (1962): *Microdiffusion Analysis and Volumetric Error*. 5th ed., Crosby Lockwood, London, UK.
- Cottyn B.G., Boucque C.V. (1968): Rapid method for the gas chromatographic determination of volatile fatty acids in rumen fluid. *Journal of Agricultural and Food Chemistry*, 16, 105–107.
- Czerkawski J.W., Clapperton J.L. (1968): Analysis of gas produced metabolism of microorganisms. *Laboratory Practice*, 17, 994–996.
- Czerkawski J.W., Breckenridge G. (1977): Design and development of a long term rumen simulation technique (Rusitec). *British Journal of Nutrition*, 38, 371–384.
- Choi N.M.A., Enser M., Wood J.D., Scollan N.D. (1998): Incorporation of soya oil hydrolysate in the diet of defaunated or refaunated sheep: effect on rumen fermentation *in vitro*. *Archives of Animal Nutrition*, 40, 320–337.
- Demeyer D.I., Van Nevel C.J. (1979): Effect of defaunation on the metabolism of rumen microorganisms. *British Journal of Nutrition*, 42, 515–524.
- Demeyer D.I., Van Nevel C.J. (1986): Influence of substrate and microbial interaction on efficiency of rumen microbial growth. *Reproduction Nutrition Development*, 26, 161–179.
- Fellner V., Sauer F.D., Kramer J.K.G. (1995): Steady state rates of linoleic acid biohydrogenation by ruminal bacteria in continuous culture. *Journal of Dairy Science*, 78, 1815–1823.
- Goering H.K., Van Soest P.J. (1970): *Forage Fiber Analyses*. Agriculture Handbook No. 379, USDA, Washington, USA.
- Gulati S.K., Kitesa S.M., Ashes J.R., Fleck E., Byers E.B., Byers Y.B., Scott T.W. (2000): Protection of conjugated linoleic acids from ruminal biohydrogenation and their incorporation into milk fat. *Animal Feed Science and Technology*, 86, 139–148.
- Hartfoot C.G., Hazlewood G.P. (1997): Lipid metabolism in the rumen. In: Hobson P.N., Stewart C.S. (eds.): *The Rumen Microbial Ecosystem*, Chapman and Hall, London, UK.
- Jalč D., Čertík M. (2005): Effect of microbial oil, monensin and fumarate on rumen fermentation in artificial rumen. *Czech Journal of Animal Science*, 50, 467–472.
- Jalč D., Potkanski A., Szumacher-Strabel M., Cieslak A., Čertík M. (2005): Effect of microbial oil, evening primrose oil and borage oil on rumen fermentation *in vitro*. *Veterinarni Medicina*, 50, 480–486.

- Jalč D., Potkanski A., Szumacher-Strabel M., Kowalczyk J., Cieslak A. (2006): The effect of a high forage diet and different oil blends on rumen fermentation *in vitro*. *Journal of Animal and Feed Sciences*, 15, 141–144.
- Kim S.C., Adesogan A.T., Staples C.R., Badinga L. (2005): The effect of dietary n-6/n-3 fatty acid ratio on feed intake, digestibility, and fatty acid profiles in muscle of growing lambs. *Journal of Animal Science*, 83, 192.
- Kim S.C., Adesogan A.T., Badinga L., Staples C.R. (2007): Effects of dietary n-6:n-3 fatty acid ratio on feed intake, digestibility, and fatty acid profiles of the ruminal contents, liver, and muscle of growing lambs. *Journal of Animal Science*, 85, 706–716.
- Lee M.R.F., Tweed J.K.S., Moloney A.P., Scollan N.D. (2005): The effects of fish oil supplementation on rumen metabolism and biohydrogenation of unsaturated fatty acids in beef steers given diets containing sunflower oil. *Animal Science*, 80, 361–367.
- Loor J.J., Hoover V.H., Miller-Webster T.K., Herbein J. H., Polan C.E. (2003): Biohydrogenation of unsaturated fatty acids in continuous culture fermenters during digestion of orchard grass or red clover with three levels of ground corn supplementation. *Journal of Animal Science*, 81, 1611–1627.
- Mansfield H.R., Endres M.I., Stern M.D. (1995): Comparison of microbial fermentation in the rumen dairy cows and dual flow continuous culture. *Animal Feed Science and Technology*, 55, 47–66.
- McDougall E.I. (1948): Studies on rumen saliva. 1. The composition and output of sheep's saliva. *Biochemical Journal*, 43, 99–109.
- Mosley E.E., Powell G., Riley M., Jenkins T.C. (2002): Microbial biohydrogenation of oleic acid to *trans* isomers *in vitro*. *Journal of Lipid Research*, 43, 290–296.
- Ørskov E.R., Flatt W.P., Moe P.W. (1968): Fermentation balance approach to estimate extent of fermentation and efficiency of VFA fermentations in ruminants. *Journal of Dairy Science*, 51, 1429–1435.
- Papanikolaou S., Komaitis M., Aggelis G. (2004): Single cell oil (SCO) production by *Mortierella isabellina* grown on high-sugar content media. *Bioresearch Technology*, 95, 287–291.
- Raes K., De Smet S., Demeyer D. (2004): Effect of dietary fatty acids on incorporation of long chain polyunsaturated fatty acids and conjugated linoleic acid in lamb, beef and pork meat. *Animal Feed Science and Technology*, 113, 199–221.
- Ratledge C. (2003): Single Cell Oils in the 21st Century. In: 94th AOCS Annual Meeting and Expo, May 4–7, Kansas City, Missouri, USA.
- Schroeder G.F., Gagliostro G.A., Becu-Villalobos D., Lacau-Mengido I. (2002): Supplementation with partially hydrogenated oil in grazing dairy cows in early lactation. *Journal of Dairy Science*, 85, 580–594.
- Váradyová Z., Kišidayová S., Siroka P., Jalč D. (2007): Fatty acid profiles of rumen fluid from sheep fed diets supplemented with various oils and effect on the rumen ciliate population. *Czech Journal of Animal Science*, 52, 399–406.
- Wachira A.M., Sinclair L.A., Wilkinson R.G., Hallett K., Enser M., Wood J.D. (2000): Rumen biohydrogenation of n-3 polyunsaturated fatty acids and their effects on microbial efficiency and nutrient digestibility in sheep. *Journal of Agricultural Science*, 135, 419–428.

Received: 2008–06–26

Accepted after corrections: 2008–11–07

Corresponding Author

MVDr. Dušan Jalč, CSc., Institute of Animal Physiology Slovak Academy of Sciences, Šoltésovej 4-6, 040 01 Košice, Slovak Republic
Tel. +421 55 792 2963, fax +421 55 728 7842, e-mail: jalcd@saske.sk

INSTITUTE OF AGRICULTURAL AND FOOD INFORMATION

Slezská 7, 120 56 Prague 2, Czech Republic

Tel.: + 420 227 010 111, Fax: + 420 227 010 116, E-mail: redakce@uzpi.cz

In this institute scientific journals dealing with the problems of agriculture and related sciences are published on behalf of the Czech Academy of Agricultural Sciences. The periodicals are published in English with abstracts in Czech.

Journal	Number of issues per year	Yearly subscription in USD
Plant, Soil and Environment	12	285
Czech Journal of Animal Science	12	285
Agricultural Economics (Zemědělská ekonomika)	12	285
Journal of Forest Science	12	285
Veterinární medicína (Veterinary Medicine – Czech)	12	285
Czech Journal of Food Sciences	6	150
Plant Protection Science	4	85
Czech Journal of Genetics and Plant Breeding	4	85
Horticultural Science	4	85
Research in Agricultural Engineering	4	85
Soil and Water Research	4	85

Subscription to these journals be sent to the above-mentioned address.

INSTRUCTIONS TO AUTHORS

The journal publishes original scientific papers and critical reviews of articles in English. Manuscripts should have abstracts (including keywords). The author is fully responsible for the originality of the paper, its subject, and its formal correctness. The author's declaration that the paper has not been published anywhere else should be enclosed. The Board of Editors decides on the publication of papers, taking into account peer reviews, scientific importance, and manuscript quality. Good laboratory practices and ethical rules must be followed. The SI international system of measurement units should be used. Manuscripts must be grammatically and linguistically correct, and authors whose native language is not English are advised to seek the help of a native English-speaker. Manuscripts containing language errors are disfavored in the reviewing process and may be returned to the author for rewriting before peer review and/or before acceptance.

Only manuscripts assessed by leading experts in the field will be published. If such reviews are not available within four months after registration of the manuscript, the peer review process is terminated, and the authors are notified. They can resubmit the manuscript, after its thorough revision and/or update, either to the Czech Journal of Animal Science or another journal for a new assessment. This should eliminate a long waiting period and probable rejection of the manuscript.

If a revision of the manuscript following the recommendation of the reviewers is requested, the modified manuscript must be re-submitted within three weeks. The authors may, however, request an extension of the re-submission deadline if necessary. All parts of the manuscript, including tables and figures (even unchanged) must be re-submitted. A detailed reply by the authors to every point of the reviewer's recommendations must be attached to the revision manuscript. It is not necessary to accept all the requests of the reviewers, but a clear explanation of why the reviewers' comments were not accepted must be provided. If the deadline for re-submission is missed, the paper will be removed from the reviewing process.

The proof reading must be returned within two days. Only errors originating during preparation of the document for printing can be corrected. Standard proof marks will be used. No changes in the manuscript after acceptance for publication are permitted.

Manuscripts should be sent by e-mail as attachments. Alternatively, they can be submitted in duplicate in hard copy, and a properly labelled Compact Disk (CD) with identical contents, including figures, should be enclosed.

Copyright. The journal is protected by copyright held by the publisher after the manuscript has been accepted for publication. As regards the transfer of rights, the corresponding author assumes responsibility for all the authors. No part of this publication may be reproduced, stored, or transmitted in any form or by any means without the written permission of the publisher.

Manuscript layout. The Microsoft (MS) Word for Windows word-processing software should be used for creating the text in non-formatted style strictly following the journal layout. If any abbreviations are used in the paper, they must be appropriately explained when they are used in the text for the first time. It is not advisable to use any abbreviation in the title of the paper or in the abstract. Tables, graphs and other Word documents are to be submitted on separate pages appended to the article. The document must not be formatted in columns, heading styles, etc. This unique MS Word file must be saved under the first author's surname only. In the printed version lines should be numbered. Graphs should be provided in MS Excel, and they should be stored with the original data. Photographs and autotypes should be submitted in high resolution (min. 300 dpi) TIFF or JPG format. All tables, graphs and photos should be numbered in the order in which they are included in the text, using Arabic numerals.

The Title of the Paper should be short and informative, and no subtitles or numbering of "serial" articles (Part I, Part II, etc.) should be used.

The Abstract should not have more than 500 words. It should contain important information on methods used to solve the problem, a clear description of results and their statistical significance, and brief and unambiguous conclusions drawn from the results. References and discussion of the results should not be included in the abstract.

Keywords should not repeat nouns used in the title and should describe the studied problem as best as possible.

The Introduction section should provide information on the present state of research in the field concerned and on the objective of the study. It should also include references to literary sources used by the authors to document their present findings, but not all literary sources that have been published to date. References in the text should agree with those in the list of references. It is recommended to include references to papers from peer periodicals only. Citations from non-available sources (reports, national journals, proceedings, thesis, etc.) should be omitted. Papers published by one or two authors are to be cited by their names, those published by three or more authors by the name of the first one, et al. If more than one paper by the same author/two authors/first author, et al., published in the same year is cited, they should be differentiated by YEARA,b,c both in the text and the list of References. Names and year of publication are to be cited by including them in the text directly, e.g. "...as published by Brown (1995)" or indirectly – citing authors and year of publication in parentheses (Green and Grey, 1996), (Jakl et al., 2002). Several papers cited together should be arranged according to the year of publication starting with the oldest one.

Material and Methods. All preliminary material, conducted experiments, their extent, conditions and course (experimental design) should be described in detail in this section. All original procedures that were used for the processing of experimental material and all analytical methods used for evaluation should also be detailed. Data verifying the quality of the acquired data should be indicated for the methods used. The entire methodology is to be described only if it is an original one; in other cases it is sufficient to cite the author of the method and to mention any particular differences. Methods of statistical processing including the software used should also be listed in this section.

Results and Discussion. The results obtained from the experiments including their statistical evaluation and any commentary should be presented graphically or in tables in this section. The author should confront partial results with data published by other authors, whose names and year of publication are to be cited by including them in the text directly or indirectly.

References should be arranged in alphabetical order according to the surname and initials of authors. The year of publication cited in parentheses, the full title of the paper in English with the language of publication in parentheses, e.g. (in Czech) should follow. The title of the periodical should be preferably typed in full. Use of official ISI Journal Citation Reports or Current Contents abbreviations is an alternative but should be used only in exceptional cases.

In the case of books or proceedings, the title should be followed by the name of the publisher, its location (Paris, New York, etc.) and the total number of pages.

Only papers cited in the text should be included in the list of references. All names of the authors must be printed in English transcription without non-English letters. Authors are responsible for the accuracy of their references.

Examples of references in the list:

Brown J. (1995): Estradiol determination in post-partum sows. *Journal of Endocrinology*, 198, 155–169.

Gabler M.T., Heinrichs A.J. (2003): Dietary protein to metabolizable energy ratios on feed efficiency and structural growth of prepubertal Holstein heifers. *Journal of Dairy Science*, 86, 268–274.

Papers published in monographs or proceedings should be cited as follows:

Kalab J. (1995): Changes in milk production during the sexual cycle. In: Hekel K. (ed.): *Lactation in Cattle*. Academic Press, London, 876–888.

Janson L., Ahlin K.A. (1992): Postpartum reproductive performance in cattle selected for high and low fat content. In: Proc. 43rd Annu. Mtg., European Association for Animal Production (EAAP), Madrid, Spain, 93–95.

The Corresponding Author should include his or her full name including all academic, scientific and pedagogic titles and the detailed address of the institution with postal code, telephone and fax numbers, and e-mail address. The author who is responsible for any correspondence with the journal should be clearly indicated.

The Declaration of the Authors must be carefully completed and signed by the first author.

Offprints: Free reprint in Portable Document Format (PDF) sent via e-mail as an attachment.

Compliance with these instructions is obligatory for all authors. If a manuscript does not comply exactly with the above requirements, the editorial office will not accept it for consideration and will return it to the authors without reviewing it.