

# Fate of applied urea $^{15}\text{N}$ in a soil-maize system as affected by urease inhibitor and nitrification inhibitor

L. Zhang<sup>1,2</sup>, Z. Wu<sup>1</sup>, Y. Jiang<sup>1</sup>, L. Chen<sup>1</sup>, Y. Song<sup>1</sup>, L. Wang<sup>3</sup>, J. Xie<sup>3</sup>, X. Ma<sup>4</sup>

<sup>1</sup>*Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang, P.R. China*

<sup>2</sup>*Graduate School of the Chinese Academy of Sciences, Beijing, P.R. China*

<sup>3</sup>*Agricultural Environment and Resources Research Center, Jilin Academy of Agricultural Sciences, Changchun, P.R. China*

<sup>4</sup>*Institute of Heilongjiang Academy of Agricultural Sciences, Harbin, P.R. China*

## ABSTRACT

A micro-plot field experiment with reduced urea  $^{15}\text{N}$  application was conducted to study the effects of urease inhibitor NBPT (N-(*n*-butyl) thiophosphoric triamide) and nitrification inhibitor DMPP (3,4-dimethyl-1H-pyrazolium dihydrogen) on the fate of applied urea  $^{15}\text{N}$ ; it aimed to find an efficient way to reduce the urea N application rate while improving the agronomic and environmental benefits. Five treatments were installed, i.e., 180 kg N/ha ( $\text{N}_1$ , conventional application rate), 126 kg N/ha ( $\text{N}_2$ , reduced to 70% conventional application rate),  $\text{N}_2$  + NBPT,  $\text{N}_2$  + DMPP, and  $\text{N}_2$  + NBPT + DMPP. Compared with treatment  $\text{N}_1$ , all the other treatments had a significantly higher total  $^{15}\text{N}$  recovery by both soil and plant ( $P < 0.05$  48.20, 41.39, 37.69, 38.85 and 34.83% soil recovery for  $\text{N}_2$  + NBPT + DMPP,  $\text{N}_2$  + DMPP,  $\text{N}_2$  + NBPT,  $\text{N}_2$  and  $\text{N}_1$  treatment, respectively; and 42.68, 40.86, 40.25, 37.18 and 36.30% plant recovery for  $\text{N}_2$  + NBPT + DMPP,  $\text{N}_2$  + DMPP,  $\text{N}_2$  + NBPT,  $\text{N}_2$ , and  $\text{N}_1$  treatment, respectively). In the plant  $^{15}\text{N}$  recovery, the  $^{15}\text{N}$  absorbed in grain/stem was highest in treatment  $\text{N}_2$  + NBPT + DMPP. The maize biomass and the maize yield had a slight increase in treatment  $\text{N}_2$  + NBPT + DMPP, compared with those in treatment  $\text{N}_1$ . In sum, for the maize production in study area,  $\text{N}_2$  + NBPT + DMPP application method would be a feasible way to ensure the normal maize yield while improving yield quality, saving urea fertilizer, and protecting the environment.

**Keywords:** nitrogen; fertilizer; N losses; N use efficiency

Agroecosystem relies on the N inputs from chemical and organic N fertilizers to sustain its productivity. However, N is a highly mobile element, which makes its efficient use and management a challenging task, especially for the over-fertilized agricultural systems where significant N losses can occur via  $\text{NO}_3^-$  leaching,  $\text{NH}_4^+$  runoff, and N gaseous emissions (Zaman et al. 2009). These N losses bring threat to the economical implications and environmental quality worldwide (Boyer et al. 2002, Ledgard et al. 2008, Vitousek et al. 2009, Zhang et al. 2009).

Various management strategies were adopted to improve the fertilizer N use efficiency and mitigate its losses from intensively cultivated lands, among

which, moderately reducing fertilizer N application rate is a commonly adopted countermeasure (Chen et al. 2006, Zhao et al. 2006, Fan et al. 2007, Ju et al. 2007, 2009). Current researches usually combine less fertilizer application with multiple amendment or with better water management in order to avoid crop yield decrease (Fan et al. 2007, Ju et al. 2007, 2009). It must be indicated that these cooperative methods usually need extra labor. Amending with urease/nitrification inhibitors was also proved as an effective means to improve the fertilizer N use efficiency and mitigate its losses. (Chen et al. 1998, Xu et al. 2001, 2002, Giller et al. 2004, Yu et al. 2007, Ledgard et al. 2008, Yu et al. 2008, Zaman et al. 2008, 2009). Lots of studies

---

Supported by the National Basic Research Program of China, Project No. 2007CB109307, and National Science & Technology Pillar Program, Project No. 2006BAD10B01.

focused on the agronomical, economical, and environmental effects under inhibitors amendment (Chen et al. 1998, Xu et al. 2001, 2002, Yu et al. 2007). However, few studies have been done on fate of N in the plant-soil system under reduced N application plus inhibitors amendment. Of the inhibitors nowadays used, N-(*n*-butyl) thiophosphoric triamide (NBPT) is an available urease inhibitor. Applied into soil, it can be quickly converted into its oxygen analogue N-(*n*-butyl) phosphoric triamide (NPBTO), which forms a tridentate ligand with soil urease (Manunza et al. 1999), retarding urea N hydrolysis. 3,4-dimethyl-1H-pyrazolium dihydrogen phosphate (DMPP) is an effective nitrification inhibitor, which can inhibit the activity of nitrifying bacteria responsible for the oxidation of  $\text{NH}_4^+$  to  $\text{NO}_2^-$ , and thereby, reduce the  $\text{NO}_3^-$  leaching and  $\text{N}_2\text{O}$  emission (Abbasi and Adams 2000, Cameron et al. 2005, Di et al. 2007, Yu et al. 2008).

In China, the consumption of chemical fertilizer-N increased rapidly since the early 1970s from 2.865 Tg in 1970 to 24.8 Tg in 1998, which accounts for ca. 30% of the world total (China Agriculture Yearbook, 1980–1999). It was estimated that the apparent recovery for agricultural production in China might be 30–35% (Zhu and Chen 2002), which causes unnecessarily high economic input and severe environmental problem.

In northeast China, the farmers often apply up to 180 kg N/ha (according to a survey of 100 farmers) to keep a high maize yield on chernozem, one of the main agricultural soils (about 20.2% of agricultural land, Wang et al. 2009) in this region. Under such a high N application rate, the use efficiency of applied N is relatively low (approximately 35%). So, in this paper, a micro-plot field experiment was conducted in Gongzhuling City of Jilin Province, Northeast China to investigate the effects of reduced N application supplemented with NBPT and/or DMPP amendment on the fate of applied  $^{15}\text{N}$  in a chernozem-maize system, aimed to approach an efficient way to reduce the urea N application rate while improving the agronomic and environmental benefits in Northeast China.

## MATERIALS AND METHODS

**Field site.** The field site is located in Gongzhuling City, Jilin province ( $123^\circ 26' 17''\text{E}$ ,  $41^\circ 44' 51''\text{N}$ ). The annual temperature is  $4.5^\circ\text{C}$ , and the annual precipitation is 450–650 mm. The soil is chernozem (USDA), with its basic characteristics listed in Table 1.

**Field experiment and lab analysis.** Chernozem (USDA) was selected as test soil (Table 1), and maize cultivar Zhengdan 958 was chosen as test crop. Five treatments were installed, i.e., conventional N application rate (180 kg N/ha,  $\text{N}_1$ ), reduced N application rate (126 kg N/ha,  $\text{N}_2$ ), reduced N application rate plus NBPT ( $\text{N}_2 + \text{NBPT}$ ), reduced N application rate plus DMPP ( $\text{N}_2 + \text{DMPP}$ ), and reduced N application rate plus NBPT + DMPP ( $\text{N}_2 + \text{NBPT} + \text{DMPP}$ ). Each treatment had three replicates. The micro plots ( $1.26 \times 0.80$  m) were enclosed with iron panel, which was inserted into soil 60 cm deep and the upper edge was 5 cm above the soil surface. Before sowing (May 2, 2008), each plot was amended with 28 g  $\text{CaH}_2\text{PO}_4$  (24 kg  $\text{P}_2\text{O}_5$ /ha) and 20 g KCl (34 kg  $\text{K}_2\text{O}$ /ha) as basal dressing. The labeled urea- $^{15}\text{N}$  (10.12 atom % excess) supplied by the China Shanghai Chemical Institute was also applied as basal fertilizer. Both NBPT and DMPP supplied by the J & K CHEMICAL LIMITED were mixed with urea- $^{15}\text{N}$  at a rate of 1% (w/w). Urea was mixed with inhibitor and then mixed with P and K fertilizer. Surface soil (0–15 cm) was dug up, then 10 kg surface soil was mixed with fertilizer and spread into micro plot uniformly. Then all the surface soil left was put back to the plot. After watering, seeds were sown on May 2, 2008.

Maize in the micro-plots was sampled at physiological yield maturity (September 28, 2008). Three samples were taken from each plot and then partitioned into the organs (root, stem and grain) to determine plant dry matter yield and total N uptake. Plant material was ground to < 0.25-mm sieve before analyses for total N (Kjeldahl method) and  $^{15}\text{N}$  abundance (Mat-251 mass spectrometer, Finnigan, Germany).

Table 1. Main physical and chemical properties of test soil

Total N	Total P	SOC	Available N	Olsen-P	pH	CEC	Sand	Silt	Clay	Bulk density (g/cm <sup>3</sup> )
	(g/kg)		(mg/kg)			(cmol/kg)	(g/kg)			
1.35	0.49	14.87	25.87	14.8	6.2	22.43	110	400	580	1.13

SOC – soil organic carbon; available N – sum of soil  $\text{NH}_4^+$ -N; ( $\text{NO}_3^- + \text{NO}_2^-$ )-N; pH, soil:water = 1:2.5; CEC – cation exchange capacity

Soil (0–60 cm) samples were taken from the micro-plots after each harvest using a steel corer of 20-mm diameter. From each micro-plot, two 60 cm deep soil cores were obtained and partitioned into 20-cm segments, with segments of the same depth being composited. Three samples were taken from each plot.

Fresh soil samples were sieved with 5 mm mesh. An aliquot of 30 g soil was extracted with 100 ml 2.0M KCl on a rotary shaker (175 r.p.m.) at 25°C for 30 min, filtered, and kept in freezer (4°C) before analysis. Soil  $\text{NH}_4^+$ -N and  $(\text{NO}_3^- + \text{NO}_2^-)$ -N concentrations were determined by AAIH Continuous Flow Analyzer. For the measurement of  $^{15}\text{N}$  enrichment, the liberated  $\text{NH}_3$  was absorbed in 10mM  $\text{H}_2\text{SO}_4$ , the  $(\text{NH}_4)_2\text{SO}_4$  solution was concentrated up to 0.25 mg N/ml, and the  $^{15}\text{N}/^{14}\text{N}$  ratio was measured with JAS CO-150  $^{15}\text{N}$  Analyzer (Axmann 1990).

Soil total N concentration was determined by Kjeldahl digestion (Keeney and Nelson 1982). 1 g soil (sieved with 0.25 mm mesh) was digested with  $\text{K}_2\text{SO}_4 + \text{CuSO}_4 + \text{Se}$  (10:1:0.1) and  $\text{H}_2\text{SO}_4$  for about 8 h. An aliquot of the digest containing 1 mg N was distilled into 10mM  $\text{H}_2\text{SO}_4$ , and the  $^{15}\text{N}/^{14}\text{N}$  was measured as described before.

Plant samples were oven-dried at 70°C, and their N concentration was determined by Kjeldahl digestion (Keeney and Nelson 1982). 0.3 g plant (ground < 0.25 mm) was digested with  $\text{K}_2\text{SO}_4 + \text{CuSO}_4 + \text{Se}$  (10:1:0.1) and  $\text{H}_2\text{SO}_4$  for about 4 h. An aliquot of the digest containing 1 mg N was distilled into 10mM  $\text{H}_2\text{SO}_4$ , and the  $^{15}\text{N}/^{14}\text{N}$  was measured as described before.

The double-distillation procedure of Pruden et al. (1985) was used to minimize  $^{15}\text{N}$  memory effect, and each  $^{15}\text{N}$  analysis was tri-replicated.

Soil organic C and total N were determined by elemental analyzer (Elementar Analysensysteme Vario EL III, Germany). Soil available N was analyzed colorimetrically using a continuous flow autoanalyzer (AutoAnalyzer 3, BRAN + LUEBBE, Germany). Soil pH was measured in a 1:2.5 soil: water suspension. Soil Olsen-P was extracted with 0.5M  $\text{NaHCO}_3$  on a reciprocal shaker for 30 min, and the P concentration in the filtrate was determined colorimetrically (880 nm) by using the procedure of Olson and Sommers (1982). Soil total phosphorus was determined by the method of Keeney and Nelson (1982). Soil CEC was determined according to Sumner and Miller (1996). Soil bulk density was measured by using the volumetric ring method (Blake 1965). Soil particle size distribution was determined by pipette method.

**Calculation and statistical analysis.** All data were computed on oven-dried weight basis. The urea- $^{15}\text{N}$  recovery by soil and plant was calculated based on the natural  $^{15}\text{N}$  enrichment of relative samples from unfertilized N plot. The difference between soil total  $^{15}\text{N}$  and mineral  $^{15}\text{N}$  ( $\text{NH}_4^+$ - $^{15}\text{N}$  and  $\text{NO}_3^-$ - $^{15}\text{N}$ ) was considered as soil organic  $^{15}\text{N}$ . Significance test was analyzed by the Duncan's multiple range test using SPSS 11.5 software for Windows, with a confidence interval of 95%.

## RESULTS AND DISCUSSION

**Soil  $^{15}\text{N}$  recovery.** Compared to the  $\text{N}_1$  treatment, soil total  $^{15}\text{N}$  recovery in the other four treatments was significantly higher ( $P < 0.05$  48.20, 41.39, 37.69, 38.85 and 34.83% for  $\text{N}_2 + \text{NBPT} + \text{DMPP}$ ,  $\text{N}_2 + \text{DMPP}$ ,  $\text{N}_2 + \text{NBPT}$ ,  $\text{N}_2$  and  $\text{N}_1$  treatments, respectively). In addition, soil total  $^{15}\text{N}$  recovery in the  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment was greater than the other three N reduction treatments ( $P < 0.05$ ). The  $\text{N}_2 + \text{DMPP}$  and  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatments significantly enhanced  $\text{NH}_4^+$ - $^{15}\text{N}$  recovery ( $P < 0.05$ ), but  $\text{N}_2$  and  $\text{N}_2 + \text{NBPT}$  did not affect it. The  $(\text{NO}_3^- + \text{NO}_2^-)$ - $^{15}\text{N}$  recovery did not change under reduced N application and reduced N application with inhibitor(s) treatments. Organic  $^{15}\text{N}$  recovery was not affected by the  $\text{N}_2$  treatment. In contrast, organic  $^{15}\text{N}$  recovery was by 54.3, 208.5, and 316.5% higher under the  $\text{N}_2 + \text{NBPT}$ ,  $\text{N}_2 + \text{DMPP}$ , and  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatments, respectively (Figure 1).

**Plant  $^{15}\text{N}$  recovery.** No significant differences in root  $^{15}\text{N}$  recovery were observed among the treatments ( $P > 0.05$ ) (Figure 2a). In comparison with  $\text{N}_1$  treatment,  $^{15}\text{N}$  recovery in stem did not change under treatments of  $\text{N}_2$ ,  $\text{N}_2 + \text{NBPT}$ ,  $\text{N}_2 + \text{DMPP}$ , but significantly decreased by 21.26% under the  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment. The  $\text{N}_2$ ,  $\text{N}_2 + \text{NBPT}$  and  $\text{N}_2 + \text{DMPP}$  treatments did not affect  $^{15}\text{N}$  recovery in grain and whole plant while the  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment significantly enhanced them by 31.64 and 17.57%, respectively.  $^{15}\text{N}$  recovery in grain accounted for the majority of  $^{15}\text{N}$  recovery in plants (Figure 2b). Either the  $\text{N}_2$  or  $\text{N}_2 + \text{DMPP}$  affected the proportion of root  $^{15}\text{N}$  recovery in the whole plants. However, the  $\text{N}_2 + \text{NBPT}$  and  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatments caused significant reductions in the proportion of root  $^{15}\text{N}$  recovery (Figure 2b). Compared to the  $\text{N}_1$  treatment, all the other four N reduction treatments significantly reduced the proportion of stem  $^{15}\text{N}$  recovery. Compared with  $\text{N}_1$  treat-

ment, the proportion of grain  $^{15}\text{N}$  recovery in the other four treatments was significantly higher ( $P < 0.05$ ). In addition, the proportion of stem  $^{15}\text{N}$  recovery under the  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment was also greater than the other three N reduction treatments. Moreover, all the four N reduction treatments stimulated the  $^{15}\text{N}$  allocation ratio of grain/stem ( $P < 0.05$ ) (Figure 2b).

**Urea  $^{15}\text{N}$  use efficiency and maize yield.** In comparison with the  $\text{N}_1$  treatment, the urea  $^{15}\text{N}$  use efficiency increased significantly under the  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment only (17.57%). Maize biomass was reduced by 5.26% under the  $\text{N}_2$  treatment, and did not change under the  $\text{N}_2 + \text{NBPT}$ ,  $\text{N}_2 + \text{DMPP}$  or  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment.

The  $\text{N}_2$  treatment caused a significant decrease (6.08%) in grain yield. No changes in grain yield were observed under the other three N reduction treatments (Figure 3).

An overall scene about the fate of applied urea  $^{15}\text{N}$  in test chernozem-maize system (Figure 4) showed that 71–86% of applied  $^{15}\text{N}$  was absorbed by plant and remained in soil, and 13–28% was lost via different ways. In comparison with the  $\text{N}_1$  treatment, the proportions of  $^{15}\text{N}$  recovery in plants and soil as well as total recovery were significantly greater (26.90, 21.43 and 22.14% for plant, soil and total recovery, respectively) under the  $\text{N}_2 + \text{NBPT} + \text{DMPP}$  treatment only. In contrast, the loss of applied  $^{15}\text{N}$  was significantly

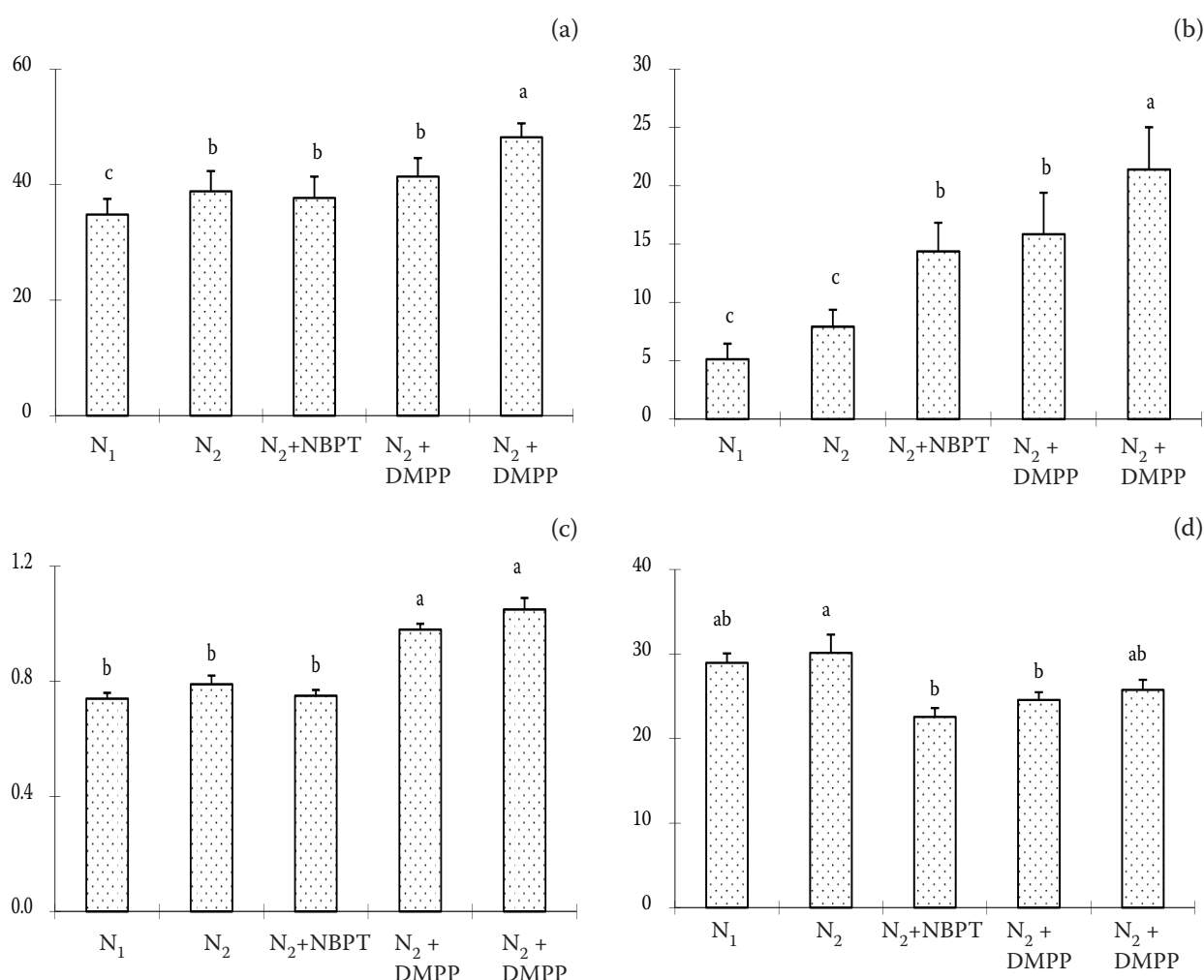


Figure 1. (a) Total  $^{15}\text{N}$  recovery (% of applied  $^{15}\text{N}$ ); (b) organic  $^{15}\text{N}$  recovery (% of applied  $^{15}\text{N}$ ); (c)  $\text{NH}_4^+$ - $^{15}\text{N}$  recovery (% of applied  $^{15}\text{N}$ ); (d)  $\text{NO}_3^- + \text{NO}_2^-$ - $^{15}\text{N}$  recovery (% of applied  $^{15}\text{N}$ ) Mean value  $\pm$  SD ( $n = 3$ ). Different letters among treatments indicate statistically different values ( $P < 0.05$ ). Organic  $^{15}\text{N}$ , the difference between soil total  $^{15}\text{N}$  and mineral  $^{15}\text{N}$  ( $\text{NH}_4^+$ - $^{15}\text{N}$  and  $\text{NO}_3^-$ - $^{15}\text{N}$  +  $\text{NO}_2^-$ - $^{15}\text{N}$ ).  $\text{N}_1$ , 180 kg N/ha;  $\text{N}_2$ , 126 kg N/ha;  $\text{N}_2 + \text{NBPT}$ , 126 kg N/ha + urease inhibitor (N-(n-butyl) thiophosphoric triamide);  $\text{N}_2 + \text{DMPP}$ , 126 kg N/ha + nitrification inhibition (3,4-dimethyl-1H-pyrazolium dihydrogen phosphate);  $\text{N}_2 + \text{NBPT} + \text{DMPP}$ , 126 kg N/ha + urease inhibitor (N-(n-butyl) thiophosphoric triamide) + nitrification inhibition (3,4-dimethyl-1H-pyrazolium dihydrogen phosphate). Vertical bars denote standard deviation of the mean (average of 3  $^{15}\text{N}$  field experiments)



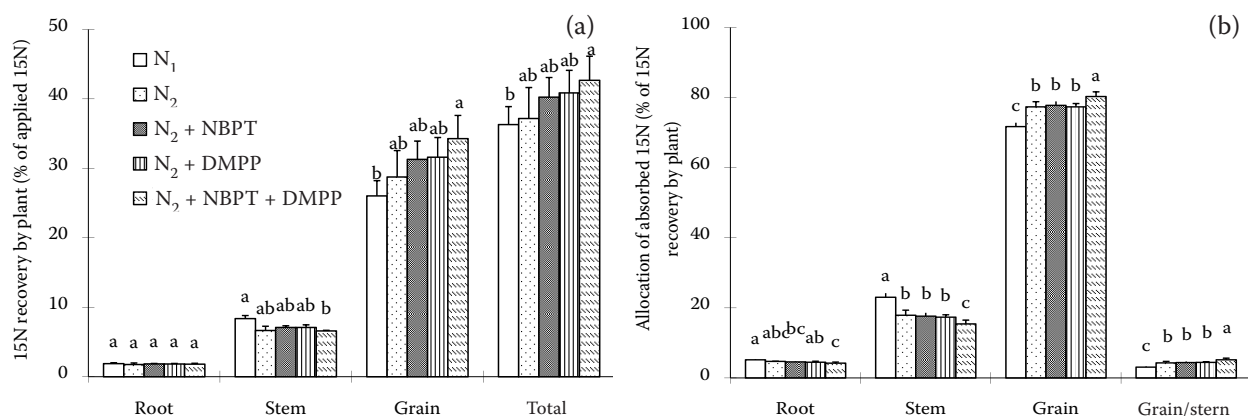


Figure 2. (a) <sup>15</sup>N recovery by plant (% of applied <sup>15</sup>N); (b) Allocation of absorbed <sup>15</sup>N (% of <sup>15</sup>N recovery by plant). Mean ± SD (*n* = 3). Different letters within treatments indicate significant differences (*P* < 0.05). For abbreviations see Figure 1

decreased (54.55%) by the N<sub>2</sub> + NBPT + DMPP treatment only.

In this study, the highest soil NH<sub>4</sub><sup>+</sup>-<sup>15</sup>N recovery observed in treatments with DMPP applied and the highest soil organic <sup>15</sup>N recovery in the treatment with combined application of DMPP and NBPT suggested the mitigation effect of DMPP on urea-<sup>15</sup>N losses via nitrification-denitrification and the synergistic effect of DMPP and NBPT on the immobilization of remained soil <sup>15</sup>N. Similar results were obtained in the related researches

with other soil types. Application of dicyandiamide (DCD) increased soil NH<sub>4</sub><sup>+</sup>-N (Vilsmeier 1991, Zaman et al. 2009). Hydroquinone (HQ) enhanced the N immobilization of fertilizer <sup>15</sup>N by 5–30% in an alkaline soil (Wang et al. 1991). The combination of HQ and DCD increased the organic N in a brown soil. Figure 1 showed lower soil NO<sub>3</sub><sup>-</sup>-<sup>15</sup>N recovery by soil in the treatments with single and combined inhibitors applied. DMPP slowed down nitrification and therefore exhibited comparatively lower amounts of NO<sub>3</sub><sup>-</sup>-<sup>15</sup>N than N<sub>1</sub>

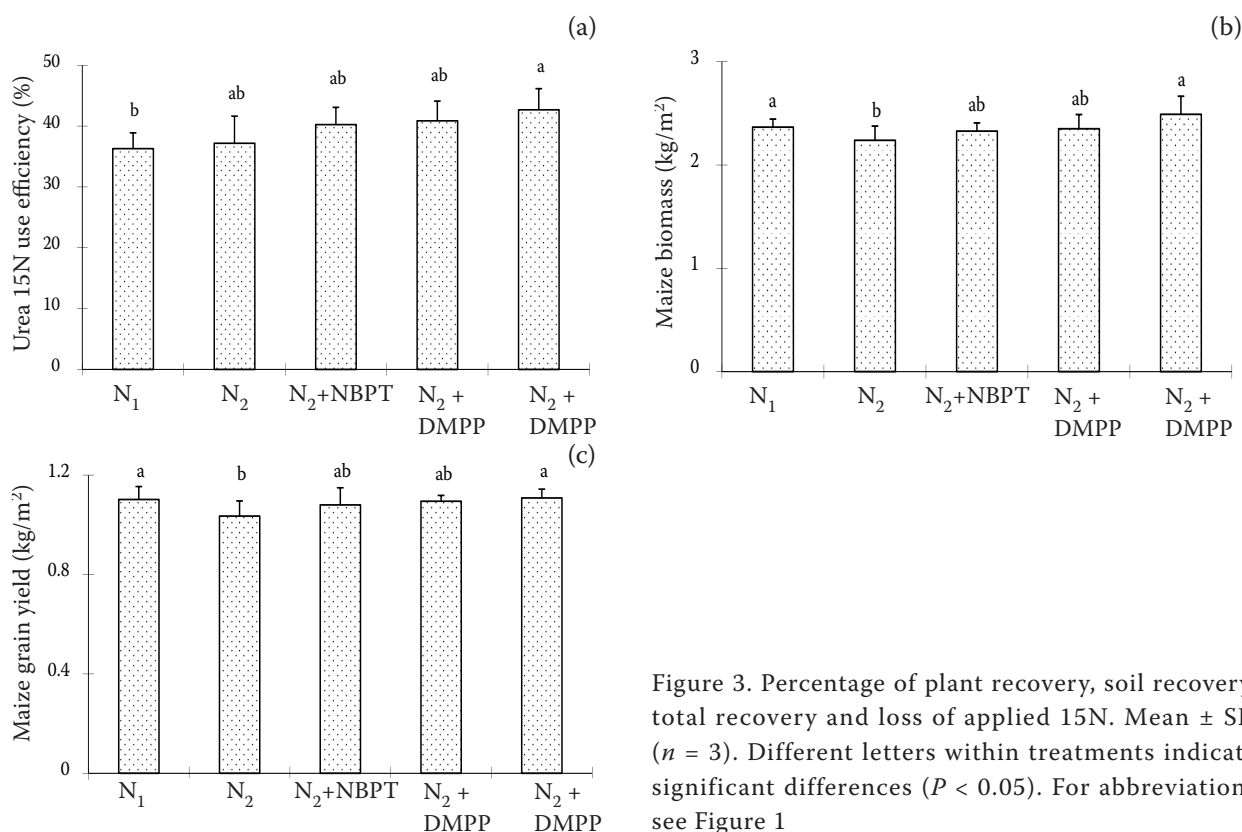


Figure 3. Percentage of plant recovery, soil recovery, total recovery and loss of applied <sup>15</sup>N. Mean ± SD (*n* = 3). Different letters within treatments indicate significant differences (*P* < 0.05). For abbreviations see Figure 1

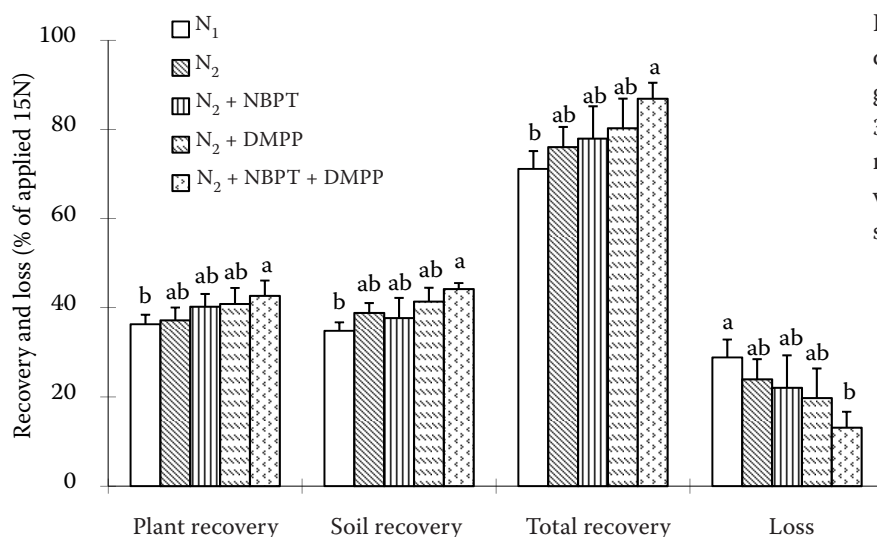


Figure 4. (a) Urea  $^{15}\text{N}$  use efficiency; (b) maize biomass; (c) maize grain yield. Mean value  $\pm$  SD ( $n = 3$ ). Different letters among treatments indicate statistically different values ( $P < 0.05$ ). For abbreviations see Figure 1

and  $\text{N}_2$ . Such reduction in nitrification is related to the partial inhibition of the activity of nitrifying bacteria by DMPP (López et al. 2003). Di et al. (2007) also observed slow nitrification after application of urine with DCD to pasture soil. Slow nitrification in the  $\text{N}_2 + \text{NBPT}$  treatment is attributed to the delayed urea hydrolysis by urease inhibitor. A similar pattern of  $\text{NO}_3^-$  production from urea fertilizer coated with NBPT was also observed by other research work (Zaman et al. 2008). The lower  $\text{NO}_3^-^{15}\text{N}$  in soil meant less risk of loss from leaching.

The NBPT, DMPP, and especially NBPT + DMPP increased the maize  $^{15}\text{N}$  recovery and promoted the translocation of absorbed  $^{15}\text{N}$  from stem to grain (Figure 2), suggesting their positive effects in improving grain quality. Recous et al. (1988a, b), Kiran and Patra (2003) and Giller et al. (2004) also reported the similar results in their studies with winter wheat, spring wheat and mint. They indicated that soil treated with inhibitors retained more available N for crop uptake by regulating the urea hydrolysis and nitrification-denitrification.

It was shown from our Figure 3 that reduced N application and its combination with amendment of NBPT or DMPP increased the urea- $^{15}\text{N}$  use efficiency but decreased the maize biomass and grain yield, which implied that the reduced N application, even amended with single NBPT or DMPP, could not satisfy the N requirement of maize growth and development. The fact that the reduced N application plus NBPT and DMPP amendment increased the maize biomass and obtained the similar grain yield as the control illustrated the synergistic effect of NBPT and DMPP on eliminating the impact of deficient fertilizer N supply. A lot of research works were consistent with our work that the ni-

trogen use efficiency and crop yield significantly increased when urease and nitrification inhibitors were applied with fertilizer (Xu et al. 2001, 2002, Kiran and Patra 2003, Zaman et al. 2009).

It could be concluded that in the maize production on chernozem in the study area, to decrease the conventional urea N application rate by 30% and amend with both NBPT and DMPP would be a feasible way to ensure the normal maize yield while improving yield quality and soil N fertility, saving urea fertilizer, and protecting the environment.

## Acknowledgments

The support by the National Basic Research Program of China (2007CB109307) and National Science & Technology Pillar Program (2006BAD10B01) is gratefully acknowledged. We thank Likai Zhou, professor of Institute of Applied Ecology for revising this manuscript. We thank Kuan Zhang and Xiufang Wang, Professor of Jilin Academy of Agricultural Sciences for providing the experimental field.

## REFERENCES

- Abbasi M.K., Adams W.A. (2000): Gaseous N emission during simultaneous nitrification-denitrification associated with mineral N fertilization to a grassland soil under field conditions. *Soil Biology and Biochemistry*, 32: 1251–1259.
- Axmann H. (1990): Method for  $^{15}\text{N}$  determination in use of nuclear techniques in studies of soil-plant relationship. IAEA Training Course, series No. 2, IAEA, Vienna.
- Blake G.R. (1965). Bulk density. In: Black C.A., Evans D.D., White J.L., Ensminger L.E., Clarck F.E. (eds.): *Methods of Soil Analysis – Physical and Mineralogical Properties Including Statistics*

- of Measurements and Sampling (Part 1). American Society of Agronomy, Madison, 374–390.
- Boyer E.W., Goodale C.L., Jaworski N.A., Howarth R.W. (2002): Anthropogenic nitrogen sources and relationships to riverine nitrogen export in the northeastern USA. *Biogeochemistry*, 57: 137–169.
- Cameron K.C., Di H.J., Moir J., Roberts A., Pellow R., Christie R. (2005): Treating grazed pasture soil with a nitrification inhibitor 'ECO-N' to decrease nitrate leaching. In: Currie L.D., Hanly J.A. (eds): *Developments in Fertilizer Application Technologies and Nutrient Management*. Fertilizer and Lime Research Centre Occasional Report No. 18. Massey University Palmerston North, New Zealand, 93–103.
- Chen L., Boeckx P., Zhou L., Cleemput O.V., Li R. (1998): Effect of hydroquinone, dicyandiamide and encapsulated calcium carbide on urea N uptake by spring wheat, soil mineral N content and N<sub>2</sub>O emission. *Soil Use Management*, 14: 230–233.
- Chen X., Zhang F., Römhild V., Horlacher D., Schulz R., Böning-Zikens M., Wang P., Claupein W. (2006): Synchronizing N supply from soil and fertilizer and N demand of winter wheat by an improved N min method. *Nutrient Cycling in Agroecosystems*, 74: 91–98.
- Di H.J., Cameron K.C., Sherlock R.R. (2007): Comparison of the effectiveness of a nitrification inhibitor, dicyandiamide, in reducing nitrous oxide emissions in four different soils under different climatic and management conditions. *Soil Use Management*, 23: 1–9.
- Fan M.S., Lu S.H., Jiang R.F., Liu X.J., Zeng X.Z., Keith W.T., Zhang F.S. (2007): Nitrogen input, <sup>15</sup>N balance and mineral N dynamics in a rice-wheat rotation in southwest China. *Nutrient Cycling Agroecosystem*, 79: 255–265.
- Giller K.E., Chaulk P., Dobermann A., Hammond L., Heffer P., Ladha J.K., Nyamudeza P., Maene L., Ssali H., Freney J. (2004): Emerging technologies to increase the efficiency of use of fertilizer nitrogen. In: Mosier A.R., Syers J.K., Freney J.R. (eds.): *Agriculture and the Nitrogen Cycle*. Island Press, USA, Washington D.C., 35–51.
- Ju X.T., Liu X.J., Pan J.R., Zhang F.S. (2007): Fate of <sup>15</sup>N-labeled urea under a winter wheat-summer maize rotation on the north china plain. *Pedosphere*, 17: 52–61.
- Ju X.T., Xing G.X., Chen X.P., Zhang S.L., Zhang L.J., Liu X.J., Cui Z.L., Yin B., Christie P., Zhu Z.L., Zhang F.S. (2009): Reducing environmental risk by improving N management in intensive Chinese agricultural systems. *Proceedings of the National Academy of Sciences of the United State of America*, 106: 3041–3043.
- Keeney D.R., Nelson R.H. (1982): *Methods of Soil Analysis*. In: Keeney D.R., Nelson R.H. (eds.): *Chemical and Microbiological Properties*. 2<sup>nd</sup> Edition. American Society of Agronomy, Madison, 643–687.
- Kiran U., Patra D.D. (2003): Medicinal and aromatic plant materials as nitrification inhibitors for augmenting yield and nitrogen uptake of Japanese mint (*Mentha arvensis* L. var. *piperascens*). *Bioresource Technology*, 86: 267–276.
- Ledgard S.F., Menneer J.C., Dexter M.M., Kear M.J., Lindsey S., Peters J.S., Pacheco D. (2008): A novel concept to reduce nitrogen losses from grazed pastures by administering soil nitrogen process inhibitors to ruminant animals: A study with sheep. *Agriculture Ecosystems and Environment*, 125: 148–158.
- López N.I., Austin A.T., Sala O.E., Méndez B.S. (2003): Controls on nitrification in a water-limited ecosystem: experimental inhibition of ammonia-oxidising bacteria in the Patagonian steppe. *Soil Biology and Biochemistry*, 35: 1609–1613.
- Manunza B., Deiana S., Pintore M., Gessa C. (1999): The binding mechanism of urea, hydroxamic acid and N-(*n*-butyl)-phosphoric triamide to the urease active site. A comparative molecular dynamics study. *Soil Biology and Biochemistry*, 31: 789–796.
- Olson S.R., Sommers L.E. (1982): Phosphorus. In: Page A.L., Miller R.H., Keeney D.R. (eds): *Methods of Soil Analysis*, part 2, Vol 9. 2<sup>nd</sup> Edition. American Society of Agronomy, Soil Science Society of America, Madison, Wisconsin USA, 403–430.
- Pruden G., Powlson D.S., Enkinson D.S. (1985): The measurement of <sup>15</sup>N in soil and plant material. *Fertility Research*, 6: 205–218.
- Recous S., Fresneau C., Faurie G., Mary B. (1988): The fate of labeled <sup>15</sup>N urea and ammonium nitrate applied to a winter wheat crop. II. Plant uptake and N efficiency. *Plant and Soil*, 112: 215–224.
- Recous S., Machet J.M. (1999): Short-term immobilization and crop uptake of fertiliser nitrogen applied to winter wheat: effect of date of application in spring. *Plant and Soil*, 206: 137–149.
- Sumner M.E., Miller W.P. (1996): Cation exchange capacity and exchange coefficients. In: Sparks D.L. (ed.): *Methods of Soil Analysis: Chemical Methods*. Soil Science Society of America, Inc. American Society of Agronomy, Inc., Madison, Wisconsin, USA, 1201–1205.
- Vilsmeier K. (1991): Fate of ammonium-N in pot studies as affected by DCD addition. *Fertility Research*, 29: 187–189.
- Vitousek P.M., Naylor R., Crews T., David M.B., Drinkwater L.E., Holland E., Johnes P.J., Katzenberger J., Martinelli L.A., Matson P.A., Nziguheba G., Ojima D., Palm C.A., Robertson G.P., Sanchez P.A., Townsend A.R., Zhang F.S. (2009): Nutrient imbalances in agricultural development. *Science*, 324: 1519–1520.
- Wang Z.P., Cleemput O.V., Li L., Baet L. (1991): Effect of organic matter and urease inhibitor on urea hydrolysis and immobilization of urea nitrogen in an alkaline soil. *Biology and Fertility of Soils*, 11: 101–104.
- Wang J.K., Wu Y., Su Q.R. (2009): General situation of northeast black soil. In: Han G.Q., Yang L.Z. (eds): *Present land use and development strategy of Northeast black soil resources*. China: Chinese Land Publishing House, 1–15.
- Xu X., Poxckx D., Wang Y., Huang Y., Zheng X., Hu F., Cleemput O.V. (2002): Nitrous oxide and methane emissions during rice growth and through rice plants: effect of dicyandiamide and hydroquinone. *Biology and Fertility of Soils*, 36: 53–58.
- Xu X.K., Huang Y., Zhou L.K., Huang G.H., Cleemput O.V. (2001): Effect of dicyandiamide and hydroquinone on the transforma-

- tion of urea-nitrogen-15 in soil cropped to wheat. *Biology and Fertility of Soils*, 34: 286–290.
- Yu Q.G., Chen Y.X., Ye X.Z., Tian G.M., Zhang Z.J. (2007): Influence of the DMPP (3,4-dimethyl pyrazole phosphate) on nitrogen transformation and leaching in multi-layer soil columns. *Chemosphere*, 69: 825–831.
- Yu Q.G., Ye X.Z., Chen Y.X., Zhang Z.J., Tian G.M. (2008): Influences of nitrification inhibitor 3,4-dimethyl pyrazole phosphate on nitrogen and soil salt-ion leaching. *Journal of Environmental Sciences*, 20: 304–308.
- Zaman M., Nguyen M.L., Blennerhassett J.D., Quin B.F. (2008): Reducing  $\text{NH}_3$ ,  $\text{N}_2\text{O}$  and  $\text{NO}_3^-$  losses from a pasture soil with urease or nitrification inhibitors and elemental S-amended nitrogenous fertilizers. *Biology and Fertility Soils*, 44: 693–705.
- Zaman M., Saggar S., Blennerhassett J.D., Singh J. (2009): Effect of urease and nitrification inhibitors on N transformation, gaseous emissions of ammonia and nitrous oxide, pasture yield and N uptake in grazed pasture system. *Soil Biology and Biochemistry*, 41: 1271–1280.
- Zhang J., Qin J., Yao W., Bi L., Lai T., Yu X. (2009): Effect of long-term application of manure and mineral fertilizers on nitrogen mineralization and microbial biomass in paddy soil during rice growth stages. *Plant, Soil and Environment*, 55: 101–109.
- Zhao R.F., Chen X.P., Zhang F.S., Zhang H.L., Schroder J., Römhild V. (2006): Fertilization and nitrogen balance in a wheat-maize rotation system in north China. *Agronomy Journal*, 98: 938–945.
- Zhu Z.L., Chen D.L. (2002): Nitrogen fertilizer use in China: Contributions to food production, impacts on the environment and best management strategies. *Nutrient Cycling in Agroecosystems*, 63: 117–127.

Received on June 29, 2009

---

*Corresponding author:*

Prof. Zhijie WU, Institute of Applied Ecology, Chinese Academy of Sciences, P.O. Box 417, Shenyang 110016, P.R. China  
 phone: + 86 24 839 703 57, fax: + 86 24 839 703 00, e-mail: wuzhijiesy@yahoo.com.cn

---