Biological effects of oomycetes elicitins

Martina Janků¹, Lucie Činčalová¹, Lenka Luhová¹, Jan Lochman², Marek Petřivalský¹*

¹Department of Biochemistry, Faculty of Science, Palacký University Olomouc, Olomouc, Czech Republic ²Department of Biochemistry, Faculty of Science, Masaryk University, Brno, Czech Republic *Corresponding author: marek.petrivalsky@upol.cz

Citation: Janků M., Činčalová L., Luhová L., Lochman J., Petřivalský M. (2020): Biological effects of oomycetes elicitins. Plant Protect. Sci., 56: 1–8.

Abstract: Successful plant defence responses to pathogen challenges are based on fast and specific pathogen recognition and plant reaction mechanisms. Elicitins, proteinaceous elicitors secreted by the *Phytophthora* and *Pythium* species, were first described in *Phytophthora* culture filtrates as proteins able to induce a hypersensitive response (HR) and resistance in tobacco at low concentrations. Later, they were classified as microbial-associated molecular patterns (MAMPs) able to induce defences in a variety of plant species. In this review, we present a comprehensive summary of the actual knowledge on the representative elicitins and their structure, perception and activation of plant signalling pathways. The current research of elicitins has been focused on a detailed understanding of the molecular mechanisms of the elicitin recognition by plant cells. Moreover, the possibility of elicitin involvement in the establishment and enhancement of plant host resistance to a broad spectrum of pathogens has been intensively studied.

Keywords: Phytophthora; cryptogein; infestin; plant immunity; resistance; pathogen

Extensive infections of agricultural crops by plant fungal and oomycete pathogens repeatedly have resulted in catastrophic harvest failures, thus, crop protection has constituted a major challenge of the past and present agricultural practise. Unlike animals, plants lack the adaptive immunity systems comprised of specialised mobile immune cells; however, they can efficiently activate a multilayer innate immune system eventually leading to plant resistance (JONES & DANGL 2006). Chemical compounds recognised by plants and able to elicit plant defence responses have been termed elicitors. A major part of plant defence responses against microbial pathogens depends on the effective detection of conserved microbial-associated molecular patterns (MAMPs) by membrane-localised pattern recognition receptors (PRRs) that induce a basal resistance response called MAMP-triggered immunity (MTI). Despite an increasing number of plant PRRs discovered over the past 20 years, for most of them, it has not been determined whether they can be recognized as true receptors thus far. This especially concerns receptor-like proteins, representing a parallel with the Toll-like receptors described in animal cells. In addition, an exposition to MAMPs can promote plants to a primed state of enhanced defences. This primed state is characterised by faster and stronger plant responses to a pathogenic stimulus compared to non-primed plants. A well-described model of plantpathogen interactions is represented by the interactions of Solanaceae plants with proteinaceous elicitors secreted by oomycetes including many significant crop pathogens like Phytophthora infestans (Mont.) de Bary, the causal agent of late blight. Oomycete pathogens, on the other hand, also include the mycoparasite Pythium oligandrum Drechsler, used as a biocontrol agent against pathogenic fungi. Thus, understanding

Supported by the Palacký University in Olomouc, Project No. IGA_PrF_2019_022.

the mechanisms involved in the oomycete infection process can lead to the development of new methods providing increased and durable plant resistance.

Elicitins – proteinaceous elicitors of oomycetes. Oomycetes, fungus-like eukaryotic phytopathogens, which can exhibit biotrophic, necrotrophic or hemibiotrophic lifestyles, secrete elicitor proteins called elicitins. Elicitins are small proteins of about 10 kDa secreted by almost all the studied Phytophthora and some Pythium species (Fefeu et al. 1997; PANABIERES et al. 1997). Elicitins are structurally highly conserved with no similarity to protein families in the plant kingdom (JIANG et al. 2006). The most studied elicitin-producing oomycetes include the hemibiotrophic Phytophthora spp. and some members of the necrotrophic Pythium spp.. Elicitins possess typical characteristic of MAMPs activating plant MTI and they have been suggested essential for an oomycetes life cycle as sterol transporters (PONCHET et al. 1999). On the other hand, their expression is down-regulated during the early biotic step of a *Phytophthora* infection (ATTARD et al. 2008; COLAS et al. 2001) and they could constitute a class of elicitors in the middle of the evolutionary zigzag model (DEREVNINA et al. 2016).

Elicitins were divided according to the phylogenetic analysis into 4 elicitin (ELI) and 13 elicitin-like (ELL) clades. All the members of the ELI clades share a typical highly conserved 98-amino acid elicitin domain with at least 66% sequence identity and six cysteine residues at conserved positions forming three disulphide bridges (Figure 1A) (BOISSY *et al.* 1996).

In the case of the ELL clades, the elicitin domain is more diverse at the sequence levels with various sequence spacing between the six cysteine residues (JIANG *et al.* 2006). Compared to elicitins from the ELI-1 clade composed uniquely by the elicitin domain, the elicitins from the other ELI clades possess an additional C-terminal domain of variable length and sequence, rich in threonine, serine and proline residues (Figure 1A) (JIANG *et al.* 2006). A potential *O*-glycosylation of this domain likely serves to anchor these elicitins to the cell wall, as proven for the glycoproteins POD-1 and POD-2 from *P. oligandrum* showing a typical elicitin signature and *O*-linked glycosylation sites (TAKENAKA *et al.* 2006).

During the last few decades, elicitins secreted during plant-oomycete interactions belonging to the ELI-1 clade have been extensively studied. The ELI-1 clade contains proteins with a conserved structure formed by five α -helixes, one β -antiparallel sheet and one ω -loop as in the case of β -CRY secreted by *Phytophthora cryptogea* Pethybr. & Laff. 1919 (PANABIERES *et al.* 1997; GOOLEY *et al.* 1998). Inside the protein core, a hydrophobic cavity exists able to accommodate fatty acids and sterols (MIKEŠ *et al.* 1997). This class is further divided into two subclasses, ELI-1A (α -elicitins) and ELI-1B (β -elicitins), based on the isoelectric points of the corresponding elicitins (Figure 1). In general, basic β -elicitins display much stronger necrotic features in comparison to acidic α -elicitins, and also provide higher plant protection against any subsequent pathogen attacks.

Elicitin perception. A major part of the plant defences against microbial pathogens depends on the effective detection of MAMPs by the plasma membrane-localised PRRs that induce basal resistance responses within the MTI. However, the role of the receptor-like proteins (RLPs) in the MAMPs recognition by the plant cells is still poorly understood. Recently, a cell surface elicitin response RLP was characterised in the wild potato Solanum microdontum Bitter (Du et al. 2015). In this plant, the perception of elicitins was reported to depend on the receptor-like kinase (RLK) SERK3/BAK1, a major modulator of PRR-mediated immunity (HEESE et al. 2007; CHAPARRO-GARCIA et al. 2011). It was recently shown that ELR constitutively associates with a proposed general interactor for RLPs, the RLK SUPPRESSOR OF BIR1-1 (SOBIR1), which is known to be required for the INF1-triggered cell death in Nicotiana benthamiana Domin and S. microdontum (DOMAZAKIS *et al.* 2018). Elicitins recognition by ELR is likely mediated by the conserved structure of the elicitin domain, as suggested by observation that elicitins from different ELI clades, which exhibit relatively low sequence identity (JIANG et al. 2006), induced a hypersensitive response (HR) in transgenic potato plants expressing ELR (Du et al. 2015). The specific ω-loop region contains a highly conserved Leu41 residue important for the elicitin perception in different plant species (DOKLÁDAL et al. 2012; STARÝ et al. 2018).

Signalling events. Early events following the elicitin recognition have been only partially disclosed and include a typical immune response characterised by changes in the ion fluxes (LECOURIEUX-OUAKED *et al.* 2000) followed by the subsequent membrane depolarisation (WENDEHENNE *et al.* 2002), acidification of the cytoplasm and alkalisation of the extracellular space (Figure 2) (PUGIN *et al.* 1997). Elicitins also trigger a burst of reactive oxygen species (ROS) and activation of the conserved pathogen-responsive MEK2-SIPK/



Figure 1. The classification and structures of the elicitin and elicitin-like proteins in oomycetes: (A) Structures of four ELI and ELL clades according to JIANG *et al.* (2006) showing the N-terminal signal peptide, the ELI domain and the variable C-terminal domain with a percentage representation of the Ser/Thr and Pro residues (JIANG *et al.* 2006), (B) the three-dimensional structures of the cinnamomin dimer, (C) the superimposition of β -CRY and the sylvaticin structures and (D) multiple sequence alignment of the 98 amino acid elicitin domain of ELI-1 from *Phytophthora* sp.

WIPK cascade mediated by the phosphorylation and up-regulation of several WRKY transcription factors (Xu et al. 2014; STARÝ et al. 2018). In tobacco, the plasma membrane NADPH oxidase, termed respiratory-burst oxidase homologue (RBOH), has a pivotal role in the first transient and the second sustained elicitin-induced ROS burst playing an important role in the regulation of the HR. The first rapid ROS burst is driven by the activation of the existing respiratory burst oxidase homologue protein D (RBOHD) proteins in the plasma membrane by the PRR-associated kinase BIK1 and calcium uptake; whereas the second massive ROS burst occurring hours after the elicitins perception is mediated by the prolonged activation of the MAPK activity, which is involved in the *de-novo* transcription of the RBOHD gene (Noirot et al. 2014).

Nitric oxide signalling participates in both the early and late phases of plant defence responses, including the HR and systemic acquired resistance (SAR). It has been recognised that nitric oxide (NO) involvement in the plant immune responses includes a cross-talk with the signalling pathways of ROS, involving S-nitrosation, a reversible posttranslational modification of the cysteine residues, which is considered to be a link between the redox changes occurring after the pathogen attack and the gene regulation of the defence proteins. The S-nitrosation of NADPH oxidase has been suggested as the crucial regulatory step for a pathogen-triggered oxidative burst (Yun et al. 2011). Nitrite reduction by the assimilatory nitrate reductase (NR, EC 1.7.1.1) is considered as the main NO source of elicitin-induced NO (YAMAMOTO-KATOU et al.



Figure 2. The signalling pathways activated by the oomycetes and elicitins with the highlighted early and late ROS bursts

cADPR – cyclic ADP ribose; E – effector; IP₃ – inositol trisphosphate; NO – nitric oxide; PK – protein kinase; RBOHA/B – respiratory burst oxidase homolog protein A/B; ROS – reactive oxygen species; SOD – superoxide dismutase; SIPK – salicylic acid-induced protein kinase; WIPK – wound-induced protein kinase; ELR – elicitin-response receptor; Fi. – fluidity increase; Oli. – order level increase; Dep. – depolarization

2006). NO production following β -CRY recognition is partly affected by the ROS, whereas NO regulates the intracellular levels of H₂O₂. Besides that, the rapid formation of peroxynitrite, the product of the reaction between NO and the superoxide radical, occurs in the elicited cells and is likely involved in the regulation of the plant responses to the elicitins (KULIK *et al.* 2015).

Elicitation also leads to changes in the intracellular trafficking. The β -CRY elicitin causes decreased levels of dynamins, proteins involved in cell trafficking, and increased levels of a specific 14-3-3 protein (STANI-SLAS *et al.* 2009). This study proposed that plant micro-domains, similar to the lipid rafts present in animal cells, are involved in the early signalling steps during the elicitation. Moreover, the effects of the elicitins on the cellular trafficking during the plant defence responses, including clathrin-mediated endocytosis, were also described (CHAPARRO-GARCIA *et al.* 2015). These processes ultimately lead to the up-regulation of the defence-related genes and to the establishment of the SAR against a wide spectrum of phytopathogens (KAMOUN *et al.* 1993).

Plant responses to elicitins. Nowadays, recognised elicitin-responsive plant species include tomato, po-

https://doi.org/10.17221/21/2019-PPS

tato and pepper (Solanaceae), grapevine (Vitaceae), citrus (Rutaceae) and some radish and turnip cultivars (Brassicaceae) (VLEESHOUWERS et al. 2006; KAMOUN et al. 1997; DALIO et al. 2017; AKINO et al. 2014). However, elicitins induce HR cell death exclusively in plants of the Nicotiana genus and some Solanum spp. but not in other Solanaceae (PONCHET et al. 1999; VLEESHOUWERS et al. 2006). It seems obvious that the SAR induced by elicitins is derived from their presence mediated by the systemic movement across the plants (Devergne *et al.* 1992; Keller *et al.* 1996; Uhlíková et al. 2016). Initially, activation of the SAmediated signalling pathway was considered the major signalling event in the elicitin-treated plants leading to induction of the SR, which can protect the host plant against the subsequent pathogen attacks (RICCI et al. 1989; KAMOUN et al. 1993; YU 1995; KELLER et al. 1996). However, the treatment of different tomato genotypes with the INF1 (*P. infestans*) and β -CRY (P. cryptogea) elicitins or with elicitin-like proteins of the non-pathogenic organism Pythium oligandrum OLI and POD-1/POD-2 activates the jasmonic acid (JA) - and ethylene (ET) - mediated expression of the pathogenesis-related proteins. The elicitin treatment also induced a plant resistance against bacterial wilt disease, powdery mildew and Phytophthora parasitica Dastur, but not P. infestans (PICARD et al. 2000; Benhamou et al. 2001; Kawamura et al. 2009; SATKOVÁ et al. 2017; STARÝ et al. 2018). In summary, these findings suggest largely different signalling pathways in response to the elicitins across the plant taxa and probably reveal that other receptors, besides the recently described ELR in the potato, are involved in the elicitin perception in other plant species.

Features determining the ability to induce plant *response*. Elicitins were previously reported as a class of sterol carrier proteins and as small proteins able to cross the cell wall and transport sterols and fatty acids. This sterol-transporting ability has been generally considered as their primary function as the *Phytophthora* and *Pythium* species are not able to synthesise the sterols necessary for their life cycle and are completely reliant on external sterol sources (HENDRIX 1970). After the sterol binds into the elicitin cavity, a shift in the ω -loop conformation of β -CRY is observed (BOISSY *et al.* 1999), which has been proposed to play a substantial role in the elicitin-inducted plant resistance (OSMAN et al. 2001). Nevertheless, construction of elicitin mutants impaired in the sterol binding showed that

this is not the crucial factor determining the defence elicitation capabilities of elicitins. Rather, the sterol binding enhances the fluidity of the plant plasma membrane, which seems to be another key point in the early signalling steps and induced ROS production (DOKLÁDAL *et al.* 2012; SANDOR *et al.* 2016).

In recent studies, the important role of the individual amino acid residues responsible for the global charge of elicitins on the ability to induce HR-cell death and the SAR has been suggested. A specific role in the HR-inducing capacity was repeatedly demonstrated for the residue in the position 13 of the elicitin domain (O'Donohue et al. 1995; Plešková et al. 2011). The neutral amino acid valine is found in this position in the acidic α -elicitins, which require micro-molar concentrations to induce the necrosis in tobacco leaves or tobacco cells. In comparison, basic β-elicitins with the amino acid lysine in this position already induce strong necrosis at nano-molar concentrations (Figure 1). Moreover, a mutation of lysine 13 for valine in β -CRY was found to change its behaviour as a basic elicitin to an acidic one (PLEŠKOVÁ et al. 2011). UHLÍKOVÁ et al. (2016) systematically replaced other lysine residues of β -CRY by threonine and also described the important role for residue lysine 39 involved in the induction of the necrosis and plant resistance (Figure 1). Principally, this study uncovered that the biological activity of elicitins, in terms of the SAR induction, results from a combination of several factors including the overall surface charge, the presence of specific lysine residues or the capacity to interact with other endogenous plant partners. Interestingly, plant lipid transfer proteins, sharing certain structural and functional similarities with elicitins, have been suggested as their potential interacting partners, the endogenous (BLEIN et al. 2002), when recently nsLTP1-elicitin (nonspecific lipid transfer proteins) complex formation curves were measured by QCM driven by a counter UZ 2400 (Grundig, Germany) (UHLÍKOVÁ et al. 2016).

Early studies also proposed that the formation of elicitin homodimers might be a crucial point of the biological activity of elicitins (PONCHET *et al.* 1999). Recently, UHLÍKOVÁ *et al.* (2016) brought new evidence supporting this hypothesis by the characterisation of the kinetic parameters of the elicitin homodimer formation and the demonstration of the key role of the specific lysine residues for the dimer formation and the SAR induction of SAR in distal plant leaves. Nevertheless, the exact role of the dimer formation in the biological activity of elicitins remains to be elucidated upon.

CONCLUSIONS

Elicitins and their effects on plant immunity have been extensively studied since their discovery in 1980s; however multiple components involved in their perception and signal transduction within the plant cells still have not been completely characterised. Studies of plant response to elicitins established an excellent model system of plant-pathogen interactions to uncover the signalling pathways, which may lead to the establishment of host resistance. Currently, the elicitin research is focused on the identification and characterisation of the cell surface receptors responsible for the elicitin recognition (CHAPARRO-GARCIA et al. 2011; DU et al. 2015; PENG et al. 2015) and on the characterization of the molecular mechanisms of the plant responses to the elicitins (JIANG et al. 2006). Recent studies have been focused on the detailed characterisation of the molecular mechanisms involved in the priming of the plant defences. Chromatin modifications were proposed to affect the priming of the defence genes through faster and stronger transcription, represented by the reported involvement of histone deacetylases as negative regulators of the elicitor-induced cell death in tobacco (BOURQUE et al. 2011). Besides this, the utilisation of elicitins to achieve an increased resistance in agricultural crops against a wide spectrum of microbial pathogens is being explored (Du et al. 2015; OUYANG et al. 2015).

References

- Akino S., Takemoto D., Hosaka K. (2014): *Phytophthora infestans*: A review of past and current studies on potato late blight. Journal of General Plant Pathology, 80: 24–37.
- Attard A., Gourgues M., Galiana E., Panabières F., Ponchet M., Keller H. (2008): Strategies of attack and defense in plant-oomycete interactions, accentuated for *Phytophthora parasitica* Dastur (syn. *P. nicotianae* Breda de Haan). Journal of Plant Physiology, 165: 83–94.
- Benhamou N., Bélanger R.R., Rey P., Tirilly Y. (2001): Oligandrin, the elicitin-like protein produced by the mycoparasite *Pythium oligandrum*, induces systemic resistance to *Fusarium* crown and root rot in tomato plants. Plant Physiology and Biochemistry, 39: 681–696.
- Blein J.P., Coutos-Thévenot P., Marion D., Ponchet M. (2002): From elicitins to lipid-transfer proteins: A new insight in cell signalling involved in plant defence mechanisms. Trends in Plant Science, 7: 293–296.
- Boissy G., De La Fortelle E., Kahn R., Huet J.C., Bricogne G., Pernollet J.C., Brunie S. (1996): Crystal structure of

a fungal elicitor secreted by *Phytophthora cryptogea*, a member of a novel class of plant necrotic proteins. Structure, 4: 1429–1439.

- Boissy G., O'Donohue M., Gaudemer O., Perez V., Pernollet J.C., Brunie S. (1999): The 2.1 A structure of an elicitinergosterol complex: a recent addition to the Sterol Carrier Protein family. Protein Science, 8: 1191–1199.
- Bourque S., Dutartre A., Hammoudi V., Blanc S., Dahan J., Jeandroz S., Pichereaux C., Rossignol M., Wendehenne D. (2011): Type-2 histone deacetylases as new regulators of elicitor-induced cell death in plants. New Phytologist, 192: 127–139.
- Chaparro-Garcia A., Wilkinson R.C., Gimenez-Ibanez S., Findlay K., Coffey M.D., Zipfel C., Rathjen J.P., Kamoun S., Schornack S. (2011): The receptor-like kinase SERK3/ BAK1 is required for basal resistance against the late blight pathogen *Phytophthora infestans* in *Nicotiana benthamiana*. PLoS ONE, 6(1): e16608. doi: 10.1371/ journal.pone.0016608
- Chaparro-Garcia A., Schwizer S., Sklenar J., Yoshida K., Petre B., Bos J.I.B., Schornack S., Jones A.M.E., Bozkurt T.O., Kamoun S. (2015): *Phytophthora infestans* RXLR-WY effector AVR3a associates with dynamin-related protein 2 required for endocytosis of the plant pattern recognition receptor FLS2. PLoS ONE: 10(9): e0137071. doi: 10.1371/journal.pone.0137071
- Colas V., Conrod S., Venard P., Keller H., Ricci P., Panabieres F. (2001): Elicitin genes expressed *in vitro* by certain tobacco isolates of *Phytophthora parasitica* are down regulated during compatible interactions. Molecular Plant-Microbe Interactions, 14: 326–335.
- Dalio R.J.D., Magãlhaes D.M., Rodrigues C.M., Arena G.D., Oliveira T.S., Souza-Neto R.R., Picchi S.C., Martins P.M.M., Santos P.J.C., Maximo H.J., Pacheco I., De Souza A., Machado M. (2017): PAMPs, PRRs, effectors and R-genes associated with citrus-pathogen interactions. Annals of Botany, 119: 749–774.
- Derevnina L., Dagdas Y.F., De la Concepcion J.C., Bialas A., Kellner R., Petre B., Domazakis E., Du J., Wu C.H., Lin X., Aguilera-Galvez C., Cruz-Mireles N., Vleeshouwers V.G.A.A, Kamoun S. (2016): Nine things to know about elicitins. New Phytologist, 212: 888–895.
- Devergne J.-C., Bonnet P., Panabières F., Blein J.-P., Ricci P. (1992): Migration of the fungal protein cryptogein within tobacco plants. Plant Physiology, 99: 843–847.
- Dokládal L., Obořil M., Stejskal K., Zdráhal Z., Ptáčková N., Chaloupková R., Damborský J., Kašparovský T., Jeandroz S., Žďárská M., Lochman J. (2012): Physiological and proteomic approaches to evaluate the role of sterol binding in elicitin-induced resistance. Journal of Experimental Botany, 63: 2203–2215.

- Domazakis E., Wouters D., Visser R.G.F., Kamoun S., Joosten M.H.A.J., Vleeshouwers V.G.A.A. (2018): The ELR-SO-BIR1 complex functions as a two-component receptor-like kinase to mount defense against *Phytophthora infestans*. Molecular Plant-Microbe Interactions, 31: 795–802.
- Du J., Verzaux E., Chaparro-Garcia A., Bijsterbosch G., Keizer L.C., Zhou J., Liebrand T.W., Xie C., Govers F., Robatzek S. (2015): Elicitin recognition confers enhanced resistance to *Phytophthora infestans* in potato. Nature Plants, 1: 15034. doi: 10.1038/nplants.2015.34
- Fefeu S., Bouaziz S., Huet J.C., Pernollet J.C., Guittet E. (1997): Three-dimensional solution structure of beta cryptogein, a beta elicitin secreted by a phytopathogenic fungus *Phytophthora cryptogea*. Protein Science: A Publication of the Protein Society, 6: 2279–2284.
- Gooley P.R., Keniry M.A., Dimitrov R.A., Marsh D.E., Keizer D.W., Gayler K.R., Grant B.R. (1998): The NMR solution structure and characterization of pH dependent chemical shifts of the beta-elicitin, cryptogein. Journal of Biomolecular NMR, 12: 523–534.
- Heese A., Hann D.R., Gimenez-Ibanez S., Jones A.M.E., He K., Li J., Schroeder J.I., Peck S.C., Rathjen J.P. (2007): The receptor-like kinase SERK3/BAK1 is a central regulator of innate immunity in plants. Proceedings of the National Academy of Sciences, 104: 12217–12222.
- Hendrix J.W. (1970): Sterols in growth and reproduction of fungi. Annual Review of Phytopathology, 8: 111–130.
- Jiang R.H.Y., Tyler B.M., Whisson S.C., Hardham A.R., Govers F. (2006): Ancient origin of elicitin gene clusters in *Phytophthora* genomes. Molecular Biology and Evolution, 23: 338–351.

Jones J.D.G., Dangl J.L. (2006): The plant immune system. Nature, 444: 323–329.

- Kamoun S., Young M., Glasscock C., Tyler B.M. (1993): Extracellular protein elicitors from *Phytophthora*: Host-specificity and induction of resistance to bacterial and fungal phytopathogens. Molecular Plant-Microbe Interactions, 6: 15–25.
- Kamoun S., van West P., de Jong A.J., de Groot K.E., Vleeshouwers V.G.A.A., Govers, F. (1997): A gene encoding a protein elicitor of *Phytophthora infestans* is downregulated during infection of potato. Molecular Plant-Microbe Interactions, 10: 13–20.
- Kawamura Y., Hase S., Takenaka S., Kanayama Y., Yoshioka H., Kamoun S., Takahashi H. (2009): INF1 elicitin activates jasmonic acid- and ethylene-mediated signalling pathways and induces resistance to bacterial wilt disease in tomato. Journal of Phytopathology, 157: 287–297.
- Keller H., Bonnet P., Galiana E., Pruvot L., Friedrich L., Ryals J., Ricci P. (1996): Salicylic acid mediates elicitin-induced systemic acquired resistance, but not necrosis in tobacco. Molecular Plant-Microbe Interactions, 9: 696–703.

- Kulik A., Noirot E., Grandperret V., Bourque S., Fromentin J., Salloignon P., Truntzer C., Dobrowolska G., Simon-Plas F., Wendehenne D. (2015): Interplays between nitric oxide and reactive oxygen species in cryptogein signalling. Plant, Cell and Environment, 38: 331–348.
- Lecourieux-Ouaked F., Pugin A., Lebrun-Garcia A. (2000): Phosphoproteins involved in the signal transduction of cryptogein, an elicitor of defense reactions in tobacco. Molecular Plant-Microbe Interactions, 13: 821–829.
- Mikeš V., Milat M-L., Ponchet M., Ricci P., Blein J-P. (1997) The fungal elicitor cryptogein is a sterol carrier protein. FEBS Letters, 416: 190–192
- Noirot E., Der C., Lherminier J., Robert F., Moricová P., Kiêu K., Leborgne-Castel N., Simon-Plas F., Bouhidel K. (2014): Dynamic changes in the subcellular distribution of the tobacco ROS-producing enzyme RBOHD in response to the oomycete elicitor cryptogein. Journal of Experimental Botany, 65: 5011–5022.
- O'Donohue M.J., Gousseau H., Huet J.C., Tepfer D., Pernollet J.C. (1995): Chemical synthesis, expression and mutagenesis of a gene encoding β-cryptogein, an elicitin produced by *Phytophthora cryptogea*. Plant Molecular Biology, 27: 577–586.
- Osman H., Vauthrin S., Mikeš V., Milat M.L., Panabières F., Marais A., Brunie S., Maume B., Ponchet M., Blein J.P. (2001): Mediation of elicitin activity on tobacco is assumed by elicitin-sterol complexes. Molecular Biology of the Cell, 12: 2825–2834.
- Ouyang Z., Li X., Huang L., Hong Y., Zhang Y., Zhang H., Li D., Song F. (2015): Elicitin-like proteins Oli-D1 and Oli-D2 from *Pythium oligandrum* trigger hypersensitive response in *Nicotiana benthamiana* and induce resistance against *Botrytis cinerea* in tomato. Molecular Plant Pathology, 16: 238–250.
- Panabières F., Birch P.R.J., Unkles S.E., Ponchet M., Lacourt I., Venard P., Keller H., Allasia V., Ricci P., Duncan J.M. (1997): Heterologous expression of a basic elicitin from *Phytophthora cryptogea* in *Phytophthora infestans* increases its ability to cause leaf necrosis in tobacco. Microbiology, 144: 3343–3349.
- Peng K., Wang C., Wu C., Huang C., Liou R.-F. (2015): Tomato SOBIR1/EVR homologs are involved in elicitin perception and plant defense against the oomycete pathogen *Phytophthora parasitica*. Molecular Plant Microbe Interactions, 28: 913–926.
- Picard K., Ponchet M., Blein J.-P.P., Rey P., Tirilly Y., Benhamou N. (2000). Oligandrin. A proteinaceous molecule produced by the mycoparasite *Pythium oligandrum* induces resistance to *Phytophthora parasitica* infection in tomato plants. Plant Physiology, 124: 379–395.
- Plešková V., Kašparovský T., Obořil M., Ptáčková N., Chaloupková R., Ladislav D., Damborský J., Lochman

J. (2011): Elicitin-membrane interaction is driven by a positive charge on the protein surface: Role of Lys13 residue in lipids loading and resistance induction. Plant Physiology and Biochemistry, 49: 321–328.

- Ponchet M., Panabières F., Milat M.L., Mikeš V., Montillet J.L., Suty L., Triantaphylides C., Tirilly Y., Blein J.P. (1999): Are elicitins cryptograms in plant-oomycete communications? Cellular and Molecular Life Sciences, 56: 1020–1047.
- Pugin A., Frachisse J.M., Tavernier E., Bligny R., Gout E., Douce R., Guern J. (1997): Early events induced by the elicitor cryptogein in tobacco cells: Involvement of a plasma membrane NADPH oxidase and activation of glycolysis and the pentose phosphate pathway. The Plant Cell, 9: 2077–2091.
- Ricci P., Bonnet P., Huet J.-C., Sallantin M., Beuvais-Cante F., Bruneteau M., Billard V., Michel G., Pernollet J.-C. (1989): Structure and activity of proteins from pathogenic fungi *Phytophthora* eliciting necrosis and acquired resistance in tobacco. European Journal of Biochemistry, 183: 555–563.
- Sandor R., Der C., Grosjean K., Anca I., Noirot E., Leborgne-Castel N., Lochman J., Simon-Plas F., Gerbeau-Pissot P. (2016): Plasma membrane order and fluidity are diversely triggered by elicitors of plant defence. Journal of Experimental Botany, 67: 5173–5185.
- Satková P., Starý T., Plešková V., Zapletalová M., Kašparovský T., Činčalová-Kubienová L., Luhová L., Mieslerová B., Mikulík J., Lochman, J., Petřivalský M. (2017): Diverse responses of wild and cultivated tomato to BABA, oligandrin and *Oidium neolycopersici* infection. Annals of Botany, 119: 829–840.
- Stanislas T., Bouyssie D., Rossignol M., Vesa S., Fromentin J., Morel J., Pichereaux C., Monsarrat B., Simon-Plas F. (2009): Quantitative proteomics reveals a dynamic association of proteins to detergent-resistant membranes upon elicitor signaling in tobacco. Molecular & Cellular Proteomics, 8: 2186–2198.
- Starý T., Satková P., Piterková J., Mieslerová B., Luhová L., Mikulík J., Kašparovský T., Petřivalský M., Lochman J. (2018): The elicitin β-cryptogein's activity in tomato is mediated by jasmonic acid and ethylene signalling pathways independently of elicitin–sterol interactions. Planta, 249: 739–749.
- Takenaka S., Nakamura Y., Kono T., Sekiguchi H., Masunaka A., Takahashi H. (2006): Novel elicitin-like proteins isolated from the cell wall of the biocontrol agent *Pythium oligandrum* induce defence-related genes in sugar beet. Molecular Plant Pathology, 7: 325–339.
- Uhlíková H., Obořil M., Klempová J., Šedo O., Zdráhal Z., Kašparovský T., Skládal P., Lochman J. (2016): Elicitininduced distal systemic resistance in plants is mediated

through the protein-protein interactions influenced by selected lysine residues. Frontiers in Plant Science, 7: 59.

- Vleeshouwers V.G.A.A., Driesprong J.D., Kamphuis L.G., Torto-Alalibo T., Van'T Slot K.A.E., Govers F., Visser R.G.F., Jacobsen E., Kamoun S. (2006): Agroinfectionbased high-throughput screening reveals specific recognition of INF elicitins in *Solanum*. Molecular Plant Pathology, 7: 499–510.
- Wendehenne D., Lamotte O., Frachisse J.M., Barbier-Brygoo H., Pugin A. (2002): Nitrate efflux is an essential component of the cryptogein signaling pathway leading to defense responses and hypersensitive cell death in tobacco. Plant Cell, 14: 1937–1951.
- Xu J., Yang K.Y., Yoo S.J., Liu Y., Ren D., Zhang S. (2014): Reactive oxygen species in signalling the transcriptional activation of WIPK expression in tobacco. Plant, Cell and Environment, 37: 1614–1625.

- Yamamoto-Katou A., Katou S., Yoshioka H., Doke N., Kawakita K. (2006): Nitrate reductase is responsible for elicitin-induced nitric oxide production in *Nicotiana benthamiana*. Plant and Cell Physiology, 47: 726–735.
- Yu L.M. (1995): Elicitins from Phytophthora and basic resistance in tobacco. Proceedings of the National Academy of Sciences USA, 92: 4088–4094.
- Yun B-W., Feechan A., Yin M., Saidi N.B.B., Le Bihan T., Yu M., Moore J.W., Kang J-G., Kwon E., Spoel S.H. (2011): S-nitrosylation of NADPH oxidase regulates cell death in plant immunity. Nature, 478: 264–268.

Received: January 30, 2019 Accepted: October 22, 2019 Published online: December 12, 2019