

Influence of Supercritical Fluid Extract of *Cinnamomum zeylanicum* Bark on Physical, Bioactive and Sensory Properties of Innovative Cinnamaldehyde-Enriched Chocolates

MUHAMMAD AHMAR JAHANGIR^{1*}, AAMIR SHEHZAD¹, MASOOD SADIQ BUTT¹
and MUHAMMAD SHAHID²

¹National Institute of Food Science and Technology and ²Department of Biochemistry,
University of Agriculture, Faisalabad, Pakistan

*Corresponding author: majahangir03@gmail.com

Abstract

Jahangir M.A., Shehzad A., Butt M.S., Shahid M. (2018): Influence of supercritical fluid extract of *Cinnamomum zeylanicum* bark on physical, bioactive and sensory properties of innovative cinnamaldehyde-enriched chocolates. Czech J. Food Sci., 36: 28–36.

The potential utility of cinnamaldehyde obtained from cinnamon (*Cinnamomum zeylanicum*) bark powder in enhancing the antioxidant capacity of dark chocolates was evaluated. To this end, the effects of conventional solvent extracts (CSE) and supercritical fluid extracts (SFE) on total phenolic contents, physical and sensory attributes of cinnamaldehyde-enriched chocolates were determined over the course of 60 days of storage. The addition of 0.5% nutraceutical_{SFE} and 1% nutraceutical_{CSE} extracts significantly increased the total phenolic contents of the chocolates to 15.62 ± 0.35 and 13.31 ± 0.26 mg GAE/g in T₂ and T₁, respectively. In terms of texture (hardness), 0.5% extract resulted in softer chocolates (64.87 ± 2.41 N) as compared to the control (70.91 ± 2.83 N). At storage termination, colorimetric results revealed a whitening of the chocolate surface that resulted in incremental increases in L^* and b^* , whereas a^* exhibited a declining trend. Moreover, the sensory results showed a better hedonic response for the enriched products. Taken together, cinnamaldehyde enrichment is beneficial for both the functional as well as oxidative stability of chocolates. We used cinnamaldehyde and maltitol (sugar replacer) to prepare functional chocolates with a special emphasis on diabetic patients, an approach that will open new horizons for innovative product development. We also optimised and characterised the extraction conditions for bioactive components using green extraction technology, i.e. a supercritical fluid extraction technique which is cost-effective and environmentally friendly.

Keywords: cinnamaldehyde; enrichment; maltitol; oxidative stability; sensorial

Nutrition and health are assuming ever-greater importance in today's world. Technological advancement, nutritional imbalances and sedentary lifestyle have led to numerous health issues that ultimately have revealed important linkages between nutrition and health and that have played a vital role in emphasising the importance of dietary interventions to cure these ailments. These connections between nutrition and health have directed consumer focus towards functional/nutraceutical food products (CHENG *et*

al. 2012). In urban settlements, confectionary such as functional chocolates have become important vehicles for the delivery of functional benefits to the consumer. The exceptional taste and numerous health benefits of chocolate are important factors for its success in the functional foods market (BELŠČAK-CVITANOVIĆ *et al.* 2012).

Chocolate is obtained from the fruit of the cocoa tree (*Theobroma cacao* L.), i.e., cocoa beans that grow in West Africa and South America (RUSCONI

<https://doi.org/10.17221/237/2016-CJFS>

& CONTI 2010). In chocolate manufacturing, the key ingredients include cocoa butter, cocoa solids, sugar and lecithin. Nevertheless, a wide range of products available on the market incorporate various ingredients like fruits, nuts and cereals. Cocoa solids, one of the major ingredients in chocolate products, are a rich source of polyphenols that provide protection to the vascular system and to health in general through their antioxidant activity (THAMKE *et al.* 2009).

Cinnamon (*Cinnamomum zeylanicum*) belongs to the family *Lauraceae* and has been utilised as a potential therapeutic agent in various cultures for centuries. Historically, cinnamon bark is amongst the oldest known spices used against gastrointestinal complaints, chronic bronchitis and inflammation of the eyes; it has been used in ayurvedic medicine for over 6000 years (SANGAL 2011). The German Commission E and the European Scientific Cooperative on Phytotherapy (ESCOP) have approved two medicinal herbs of the genus *Cinnamomum*: *C. zeylanicum*, and *C. cassia* (European Scientific Cooperative on Phytotherapy 2003).

Cinnamaldehyde (3-phenyl-2-propanal) represents the main constituent of cinnamon bark oil and constitutes 49.9–62.8% of the total (SIMI *et al.* 2004; EL-BAROTY *et al.* 2010). It provides protection against metabolic syndromes like cardiovascular complications and diabetes. It also improves the functionality of insulin receptors by virtue of activation of the enzyme (insulin receptor kinase) which is responsible for the binding of insulin to cells. Moreover, it is responsible for inhibiting the activity of the enzyme that impedes this process (insulin receptor phosphatase), ultimately leading to the maximum phosphorylation of insulin receptor, which is associated with improved insulin sensitivity. It also increases glucose tolerance by enhancing the activity of hexokinase and mobilising glycogen stores in the liver and skeletal muscle. Cinnamon polyphenols contribute to the regulation of various proteins like glucose transporter 4 (GLUT4), insulin receptor β and tristetraprolin involved in the insulin signal transduction pathway (MAHFOUZ *et al.* 2010). Furthermore, it exhibits good antioxidant and hypolipidaemic properties, which reduce the concentration of total cholesterol and triglyceride and curb intestinal α -glycosidase activity (AL-JAMAL 2009). To the best of our knowledge, insufficient research has been conducted with respect to polyphenol-enriched chocolates. The enrichment of cinnamaldehyde as an antioxidant from cinnamon bark at different concen-

trations enhances the bioactive profile of enriched chocolates and contributes to further improving the attractive flavour of the product. Consequently, the aim of this study was, for the first time, to evaluate the bioactive profile and hedonic response of chocolates enriched with cinnamaldehyde.

MATERIAL AND METHODS

Procurement of raw material. The dried cinnamon (*Cinnamomum zeylanicum*) bark was procured from a local market and was ground to obtain particle sizes in the range of 300–500 μm for further analysis. The cocoa liquor, cocoa butter, soy lecithin and maltitol were purchased from a local supplier. The reagents (analytical and HPLC grade) and standards were purchased from Merck KGaA (Germany) and Sigma-Aldrich (Japan).

Conventional solvent extraction (CSE). The extraction of cinnamaldehyde was performed using aqueous ethanol (50% v/v) as a solvent. Thirty grams of cinnamon (*Cinnamomum zeylanicum*) bark powder were poured into the solvent and extracted for 60 min under a constant temperature of 50°C following the guidelines of MARIOD *et al.* (2010). The resulting solvent extracts were filtered and recovered using a rotary evaporator (Eyela, Japan).

Supercritical fluid extraction (SFE). Cinnamon powder was subjected to a supercritical fluid extraction system (SFT-150) to obtain supercritical fluid extracts using 99.8% pure CO_2 . Accordingly, samples were placed in a 100-ml extraction vessel followed by optimisation of CO_2 at 5000 psi pressure while maintaining time and temperature conditions constant (MARONGIU *et al.* 2007).

Preparation of cinnamaldehyde-enriched chocolates. In the product development steps, three types of functional chocolates were prepared. T_1 contained nutraceutical_{CSE}, whilst T_2 was prepared using nutraceutical_{SFE} supplementation. T_0 acted as a control (Table 1). Briefly, to produce cinnamaldehyde-enriched chocolates, cocoa butter was pre-melted by heating at 60°C to achieve 25% total fat content and added to cocoa liquor, maltitol (sucrose alternative) and vanillin followed by mixing for 10 minutes. Afterwards, the chocolate mixtures were conched for 6 h with the addition of lecithin as an emulsifier at the end of the conching process. The resulting functional chocolates were stored at 18°C after tempering. During functional/nutraceutical chocolate

Table 1. Treatments used in product development

Treatment	Description
T ₀	control
T ₁	cinnamaldehyde enriched chocolate with nutraceutical _{CSE}
T ₂	cinnamaldehyde enriched chocolate with nutraceutical _{SFE}

CSE – conventional solvent extraction; SFE – supercritical fluid extraction

production, nutraceutical_{CSE} and nutraceutical_{SFE}, were added at 1 and 0.5% in T₁ and T₂, respectively. Moreover, for treatment T₀ (control) the same procedure was followed only without the addition of bioactive ingredient.

Storage. The cinnamaldehyde-enriched chocolates were examined for physical parameters, phenolic contents and sensory response at storage intervals of 0, 15, 30, 45, and 60 days.

Preparation of chocolate extract. Chocolate extracts were prepared using the protocols published by BELŠČAK-CVITANOVIĆ *et al.* (2015). Frozen chocolate (2 g) was subjected to extraction three times with 10 ml of *n*-hexane in order to obtain fat free samples; this was followed by 24 h of air drying to remove residual organic solvents. Afterwards, extraction of phenolic compounds was performed using 10 ml aqueous methanol (70% v/v) for 30 min and the mixture was centrifuged at 3000 g for 10 minutes. The supernatants were removed, filtered and then combined to make 20 ml of extract.

Colour analysis. Colour analysis of the prepared chocolates was performed using the CIELAB Color Meter (CIELAB SPACE; Color Tech-PCM, USA) following the guidelines of LARA *et al.* (2010). The surface of the products were exposed to light and the colour values L^* , a^* , and b^* were determined. The data thus obtained were used to calculate chroma (C^*) and hue angle (h):

$$C^* = [(a^*)^2 + (b^*)^2]^{1/2}$$

$$h = \tan^{-1} (b^*/a^*)$$

Texture analysis. The triple beam snap (three-point break) technique of a texture analyser (TA-HDi; Stable Microsystems, UK) was used to determine the texture of prepared chocolates. A penetration test was conducted by using a 2-mm cylinder probe with a load cell of 5 kg. The chocolate samples were penetrated with a crosshead speed of 0.5 mm/s at

room temperature. Force (N) required for penetration was noted and average values were calculated according to the prescribed methods (LARA *et al.* 2010; BELŠČAK-CVITANOVIĆ *et al.* 2012).

Determination of total phenolic contents (TPC). Total phenolic contents of all chocolates were estimated following the method of LACHMAN *et al.* (1998).

Sensory evaluation. The end products were evaluated with quantitative descriptive analysis by a panel of judges in accordance to ISO standards using a 9-point hedonic scale system as mentioned by SIM *et al.* (2016).

Statistical analysis. The resulting data were examined using completely randomised design (CRD) in Cohort v6.1 (Costat; 2003). Moreover, significance was determined using analysis of variance (ANOVA) following the principles outlined by STEEL *et al.* (1997).

RESULTS AND DISCUSSION

Colour. Colour is a critical discriminating feature of a given product and is used for rational selection by consumers. It also constitutes the basis of raw material assessment and quality product development. The colour of cinnamaldehyde-enriched chocolate was measured using a CIELAB colour system that includes L^* for lightness, a^* for redness, and b^* for yellowness. The mean L^* of chocolates are presented in Table 2. The L^* for control chocolates (T₀), cinnamaldehyde-enriched chocolates with nutraceutical_{CSE} (T₁), and cinnamaldehyde-enriched chocolates with nutraceutical_{SFE} (T₂) were recorded as 21.19 ± 0.57 , 17.36 ± 0.45 , and 19.82 ± 0.48 , respectively. Storage resulted in a significant increase in this trait, from 18.69 ± 0.58 (day 0) to 20.21 ± 0.43 (day 60). Similarly, a^* were also significantly affected by treatments. The highest value for this trait, 8.48 ± 0.23 , was observed in T₂ followed by 8.13 ± 0.25 in T₁. The lowest value, 7.78 ± 0.19 , was noted in T₀. Over the course of storage, values for a^* decreased non-significantly, from 8.42 ± 0.24 at the initiation of the study to 7.88 ± 0.20 at its end (Table 2).

Likewise, it is obvious from the mean values for b^* (Table 2) that there were significant variations in different chocolates. The highest value was measured for T₀ (10.01 ± 0.42) followed by T₂ (9.67 ± 0.39), whereas the smallest value was reported in T₁ (8.52 ± 0.36). A significant increase in this parameter was observed for this trait over the course of storage.

<https://doi.org/10.17221/237/2016-CJFS>

Table 2. Effect of treatments and storage on the L^* , a^* , b^* , chroma, and hue angle of chocolates

Days	Treatments			Means
	T ₀	T ₁	T ₂	
<i>L</i> [*]				
0	20.27 ± 0.86	16.65 ± 0.49	19.14 ± 0.57	18.69 ± 0.58 ^c
15	20.84 ± 0.65	16.93 ± 0.38	19.46 ± 0.43	19.08 ± 0.47 ^b
30	21.20 ± 0.53	17.42 ± 0.57	19.89 ± 0.64	19.50 ± 0.61 ^{ab}
45	21.49 ± 0.74	17.78 ± 0.61	20.11 ± 0.39	19.79 ± 0.55 ^{ab}
60	22.13 ± 0.48	18.01 ± 0.42	20.49 ± 0.51	20.21 ± 0.43 ^a
Means	21.19 ± 0.57 ^a	17.36 ± 0.45 ^b	19.82 ± 0.48 ^{ab}	
<i>a</i> [*]				
0	8.12 ± 0.16	8.39 ± 0.23	8.75 ± 0.34	8.42 ± 0.24
15	7.89 ± 0.22	8.30 ± 0.19	8.63 ± 0.26	8.28 ± 0.27
30	7.75 ± 0.18	8.14 ± 0.27	8.47 ± 0.25	8.12 ± 0.19
45	7.61 ± 0.13	7.96 ± 0.16	8.32 ± 0.12	7.97 ± 0.15
60	7.53 ± 0.24	7.87 ± 0.21	8.24 ± 0.17	7.88 ± 0.20
Means	7.78 ± 0.19 ^b	8.13 ± 0.25 ^{ab}	8.48 ± 0.23 ^a	
<i>b</i> [*]				
0	9.45 ± 0.38	7.99 ± 0.25	9.23 ± 0.44	8.89 ± 0.32 ^b
15	9.72 ± 0.41	8.28 ± 0.32	9.36 ± 0.35	9.12 ± 0.37 ^b
30	9.97 ± 0.33	8.53 ± 0.27	9.71 ± 0.42	9.40 ± 0.29 ^{ab}
45	10.31 ± 0.57	8.77 ± 0.49	9.89 ± 0.36	9.66 ± 0.45 ^{ab}
60	10.59 ± 0.45	9.04 ± 0.48	10.15 ± 0.51	9.93 ± 0.44 ^a
Means	10.01 ± 0.42 ^a	8.52 ± 0.36 ^b	9.67 ± 0.39 ^{ab}	
Chroma				
0	12.46 ± 0.53	11.59 ± 0.38	12.72 ± 0.60	12.26 ± 0.42
15	12.52 ± 0.45	11.72 ± 0.36	12.73 ± 0.57	12.33 ± 0.53
30	12.63 ± 0.49	11.79 ± 0.41	12.89 ± 0.44	12.44 ± 0.45
45	12.82 ± 0.61	11.85 ± 0.54	12.93 ± 0.56	12.53 ± 0.57
60	12.99 ± 0.47	11.99 ± 0.45	13.08 ± 0.63	12.69 ± 0.48
Means	12.68 ± 0.51 ^a	11.79 ± 0.43 ^b	12.87 ± 0.49 ^a	
Hue angle				
0	0.86 ± 0.027	0.76 ± 0.019	0.81 ± 0.021	0.81 ± 0.024
15	0.89 ± 0.015	0.79 ± 0.018	0.83 ± 0.010	0.83 ± 0.017
30	0.91 ± 0.023	0.81 ± 0.015	0.86 ± 0.024	0.86 ± 0.020
45	0.94 ± 0.018	0.84 ± 0.011	0.87 ± 0.013	0.88 ± 0.015
60	0.95 ± 0.029	0.86 ± 0.017	0.89 ± 0.016	0.90 ± 0.021
Means	0.91 ± 0.022 ^a	0.81 ± 0.013 ^b	0.85 ± 0.017 ^{ab}	

T₀ – control chocolate (without active ingredient); T₁ – cinnamaldehyde enriched chocolate with nutraceutical_{CSE}; T₂ – cinnamaldehyde enriched chocolate with nutraceutical_{SFE}

The recorded values for b^* were 8.89 ± 0.32, 9.40 ± 0.29, and 9.93 ± 0.44, respectively at days 0, 30, and 60. Regarding chroma, substantial increases up to 11.99 ± 0.45, 12.99 ± 0.47, and 13.08 ± 0.63 were detected in T₁, T₀, and T₂, respectively (Table 2).

In contrast, storage resulted in non-significant increase in chroma values at day 60 compared to day 0, i.e., 12.69 ± 0.48 vs. 12.26 ± 0.42. The hue angle underwent a significant increase in response to treatments. The highest value for this trait was noted in

T_0 (0.91 ± 0.022) followed by T_2 (0.85 ± 0.017), and T_1 (0.81 ± 0.013). Furthermore, at initiation and termination of the study, hue angle presented values of 0.81 ± 0.024 and 0.90 ± 0.021 , respectively, that differed non-significantly from each other (Table 2). The findings are in agreement with the results of ROSSINI *et al.* (2011) who investigated the impact of synthetic and natural casein peptide antioxidants on non-enzymatic browning and lipid oxidation of milk chocolate stored at 20 and 28°C for 10 months. They observed a rise in the red colour intensity of milk chocolates as evidenced by increasing a^* . They also reported the negative effects of antioxidants on yellow colour development: b^* in antioxidant-enriched chocolates increased to a lesser degree compared to samples without antioxidants.

The results regarding the functional/nutraceutical chocolates are also supported by the work of BRIONES and AGUILERA (2005), who examined the effect of temperature fluctuations between 16 and 28°C on the colour of milk chocolate tablets over the course of 52 days of storage. They witnessed an increasing trend in the whiteness index (WI) and in L^* after 36 days of storage. This change in colour might be due to the migration of fat to the chocolate surface resulting in a non-uniform colour pattern. Similarly, AFOAKWA *et al.* (2008) observed a higher L^* of 80.60 ± 1.26 in under-tempered dark chocolate compared with 43.43 ± 1.02 in over-tempered and 44.79 ± 1.16 in optimally tempered samples using the same 25- μm particle size. They were of the view that the tempering process results in the occurrence of blooming which affects the colour of dark chocolate samples during storage.

The colorimetric results of NIGHTINGALE *et al.* (2009) revealed the variations in colour, i.e., L^* , a^* , and b^* in dark chocolate stored at various temperatures and degrees of relative humidity. They concluded that chocolates stored at high temperatures with fluctuations were substantially lighter in colour with a 70% increase in the whiteness index over the course of eight weeks of storage. MEXIS *et al.* (2010) reported that chocolate packaged under a vacuum illustrated the smallest increase in the L^* , from 32.89 to 49.00, and that the highest increase, from 32.89 to 56.12, occurred in commercially packaged chocolates. They concluded that lightness in chocolates is related to the incidence of white spots on the surface during storage.

Texture. Means of the hardness of cinnamaldehyde-enriched chocolates in response to treatments and

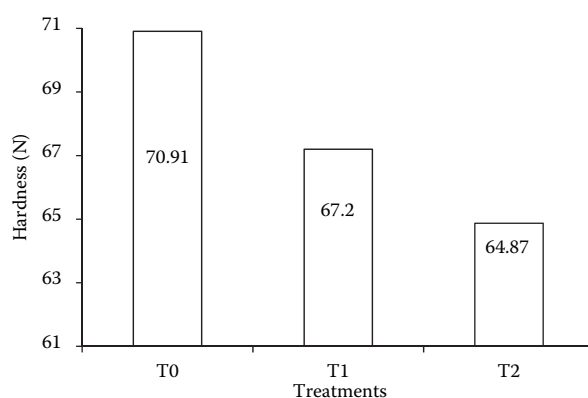


Figure 1. Effect of treatments on hardness (N) of chocolate T_0 – control chocolate (without active ingredient); T_1 – cinnamaldehyde enriched chocolate with nutraceutical_{CSE}; T_2 – cinnamaldehyde enriched chocolate with nutraceutical_{SFE}

storage are shown in Figures 1 and 2, respectively. Amongst treatments, T_0 (control chocolate) exhibited the highest hardness of 70.91 ± 2.83 N followed by 67.20 ± 2.94 and 64.87 ± 2.41 N in cinnamaldehyde-enriched chocolate with nutraceutical_{CSE} (T_1) and cinnamaldehyde-enriched chocolate with nutraceutical_{SFE} (T_2), respectively. Nevertheless, during storage, there was a non-significant reduction in hardness which fell from 67.96 ± 2.65 N at the beginning to 67.62 ± 2.49 and 67.37 ± 2.36 N on days 30 and 60, respectively. Recently, BELŠČAK-CVITANOVIĆ *et al.* (2015) investigated the influence of natural sweeteners on the texture and particle size distribution of chocolate. They showed that a combination of fructose, maltitol and stevia leaves leads to increased hardness (86.60 ± 0.06 N) as compared to control (56.82 ± 8.63 N) chocolate, which was attributed to an increased particle size median. Moreover, these

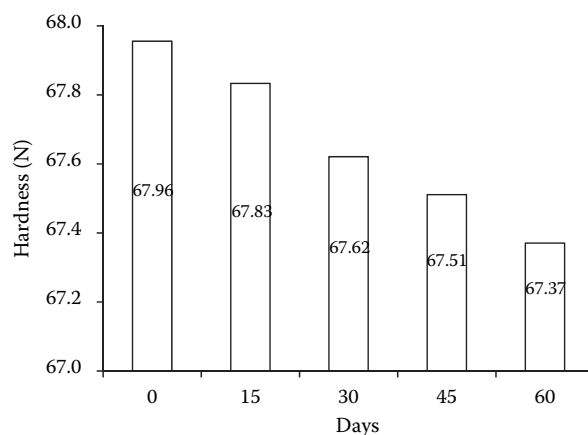


Figure 2. Effect of storage on the hardness (N) of chocolates

<https://doi.org/10.17221/237/2016-CJFS>

authors suggested that the particle size of the mixtures significantly affected the hardness of chocolates.

Previously, ALI *et al.* (2001) reported that the increased migration of the palm mid fraction (PMF) into the chocolate layer stored at higher temperatures causes polymorphic transformation and blooming. Chocolates stored at 18°C exhibited very slow PMF and DCN migration, which resulted in minimal changes in glossiness, hardness, and polymorphic stability. The current findings are also in agreement with the work of DO *et al.* (2007) who estimated the effect of particle size distribution on the textural properties of chocolate. They reported optimising PSD and fat content resulted in a reduction in hardness from 55 to 42 N. Mechanistically, a decline in hardness is due to inadequate contact between particles in the suspension. In a more recent study, LEE *et al.* (2009) replaced cocoa butter with β -glucan-rich hydrocolloid and evaluated its impact on the rheological and textural properties of chocolate. The results revealed that the addition of 15% C-trim30 (β -glucan-rich hydrocolloid) resulted in hardness dropping to 27 N compared to 40 N in control. Replacement of cocoa butter with β -glucan hydrocolloid influences the tempering process which results in soft-textured chocolates.

The current results relating to the cinnamaldehyde enrichment of dark chocolates are in harmony with the findings of BELŠČAK-CVITANOVIĆ *et al.* (2012) who evaluated the enrichment of 1 and 3% concentrated and 1% freeze-dried red raspberry leaf extract. They found the lowest hardness of 13.06 ± 0.31 N

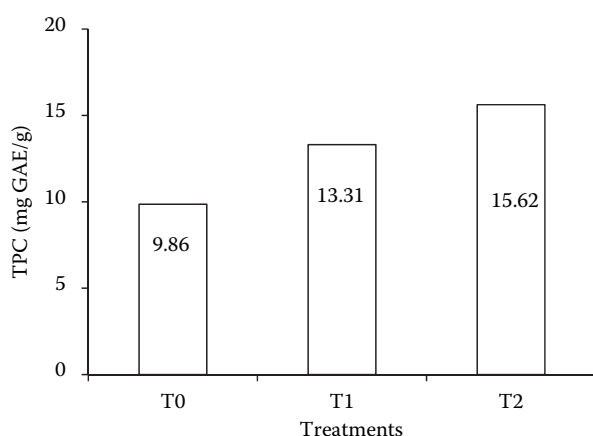


Figure 3. Effect of treatments on TPC (mg GAE/g) of chocolate

T₀ – control chocolate (without active ingredient); T₁ – cinnamaldehyde enriched chocolate with nutraceutical_{CSE}; T₂ – cinnamaldehyde enriched chocolate with nutraceutical_{SFE}

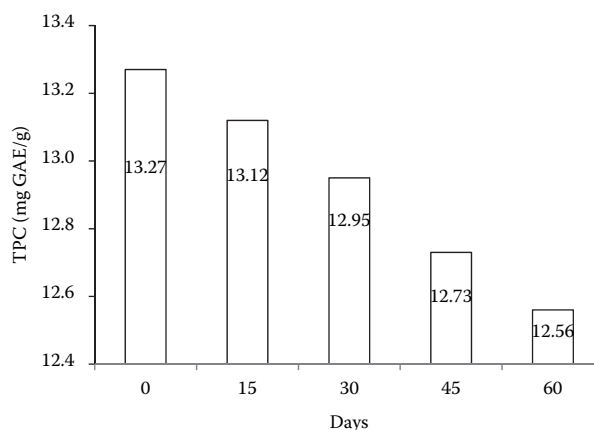


Figure 4. Effect of storage on TPC (mg GAE/g) of chocolates

for chocolates enriched with freeze-dried extract, whereas, amongst dark chocolates, addition of 3% concentrated extract resulted in an increased hardness of 37.08 ± 0.41 N. Furthermore, they concluded that the addition of red raspberry leaf extract significantly reduced the hardness of chocolates.

Total phenolic contents (TPC). It is evident from Figure 3 that the means for total phenolic contents (TPC) of cinnamaldehyde-enriched chocolates showed the highest value of 15.62 ± 0.35 mg GAE/g in T₂ (cinnamaldehyde-enriched chocolate with nutraceutical_{SFE}) followed by 13.31 ± 0.26 mg GAE/g in T₁ (cinnamaldehyde-enriched chocolate with nutraceutical_{CSE}), while the lowest TPC value of 9.86 ± 0.29 mg GAE/g was measured in control chocolate (T₀). Likewise, during storage, the values for TPC varied from 13.27 ± 0.33 mg GAE/g (day 0) to 12.56 ± 0.22 mg GAE/g (day 60) representing a significant decrease (Figure 4). The increases in TPC values caused by cinnamaldehyde enrichment are in agreement with the results of SIM *et al.* (2016) who assessed the rise in total phenolic contents in response to the addition of mangosteen pericarp powder in dark (DC) and compound chocolates (CC). The research findings showed 13 and 50% increases in DC_{3%} and CC_{3%}, respectively, for this trait when compared with their control counterparts.

KOMES *et al.* (2013) determined the polyphenolic contents and antioxidant capacity of milk and bitter chocolates enriched with various dried fruits. Their findings are in agreement with ours and they discovered bitter chocolate enriched with the acetone extract of dried prunes evinced higher total phenolic contents (6.08 mg GAE/g) than plain chocolate (4.60 mg GAE/g). BELŠČAK-CVITANOVIĆ *et al.* (2012) described the impact of *Rubus idaeus* L. polyphenol

enrichment on semisweet chocolate, milk chocolate and dark chocolate. They found that enrichment with 3% concentrated red raspberry extract increased the total phenolic contents (TPC) from 0.64 mg GAE/g in milk chocolate to 1.79 mg GAE/g in semisweet chocolates.

CERVELLATI *et al.* (2008) described total phenolic contents of 16 ± 1 in artisan-made rosemary flavoured chocolate and 13.2 ± 0.8 mg GAE/g multinational-made chocolates. This was due to the presence of rich antioxidant compounds in rosemary, i.e., carnosic acid, rosmarinic acid and carnosol. MILLER *et al.* (2006) reported a polyphenol level of 12.97 mg GAE/g in commercially available dark chocolate in the United States. In another investigation, reduced sugar chocolates were produced by incorporating sugar alcohols, syrups, dietary fibres and natural sweeteners. The results indicated that amongst reduced sugar chocolates, the highest TPC (11.27 mg GAE/g) was mainly attributable to the presence of peppermint and stevia leaves (BELŠČAK-CVITANOVIĆ *et al.* 2015).

Sensory evaluation. Hedonic response is critical for interpreting consumer response to a product and for determining acceptance and marketability. With this in mind, the functional/nutraceutical chocolates were evaluated for various sensory attributes including appearance, texture, aroma, mouthfeel, aftertaste, and overall acceptability.

Figure 5 presents the sensory evaluation of cinnamaldehyde-enriched chocolates as a function of treatments and storage. It is obvious from Figure 5A that treatments resulted in significant variations in appearance, aroma, mouthfeel, and aftertaste, whilst texture and overall acceptability differed non-significantly. Cinnamaldehyde-enriched chocolate containing 0.5% nutraceutical_{SFE} (T_2) scored higher

for appearance, aroma and mouthfeel followed by T_1 (cinnamaldehyde-enriched chocolate with 1% nutraceutical_{CSE}) and the T_0 control (without active ingredient). However, the addition of 0.5% and 1% extract negatively altered the aftertaste of chocolate with maximum scores attained by T_1 followed by T_2 , and T_0 which got lowest scores for this trait. During storage, all the sensory attributes changed non-significantly except for the mouthfeel of enriched chocolates, which exhibited a significant decline in sensory acceptance for this trait when comparing initiation and termination of storage (Figure 5B). A descriptive study on chocolates conducted by DÜRRSCHMID *et al.* (2006) concluded that appearance, texture, mouthfeel and taste are the most important sensory categories for product description and consumer perception of novel products.

The current findings relating to the hedonic response of cinnamaldehyde-enriched chocolate are in agreement with the results of ALBAK and TEKIN (2015) who conducted a sensory trial of cinnamon chocolate to evaluate its taste, smell and textural profile. They found significant variations for bitterness, coarseness, thickness and hardness, whilst the overall acceptability of cinnamon chocolate differed non-significantly revealing a high acceptance of the expert panellists of the new product. In a recent investigation, SIM *et al.* (2016) determined the effect of mangosteen pericarp powder enrichment on sensory and nutritional properties of chocolates. They reported non-significant differences in sensory attributes including sweetness, bitterness, flavour, and aftertaste between plain and enriched chocolates.

Similar findings were documented by BELŠČAK-CVITANOVIĆ *et al.* (2012), which support the sen-

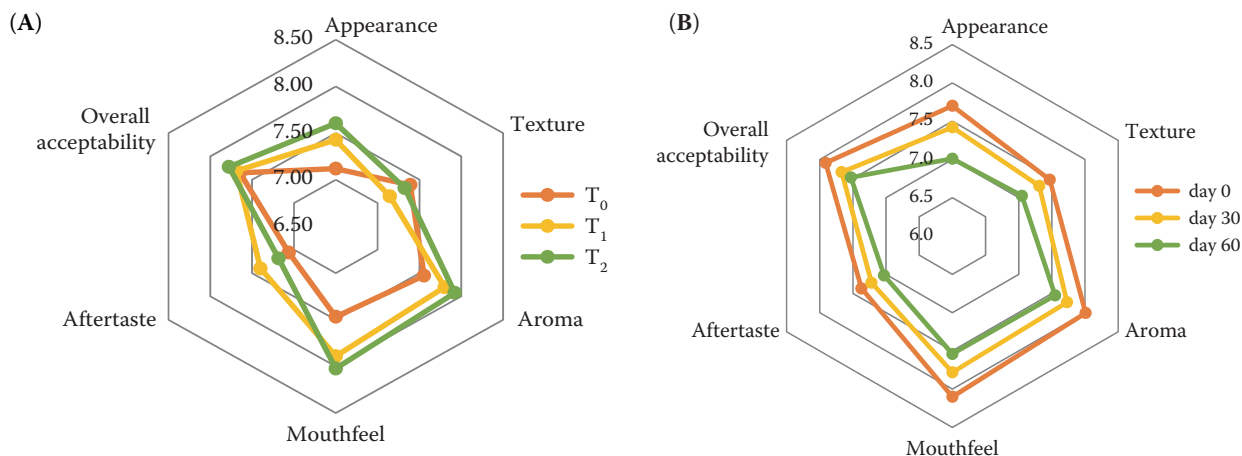


Figure 5. Effect of (A) treatments and (B) days of storage on the sensory attributes of chocolates

<https://doi.org/10.17221/237/2016-CJFS>

sory profiling results described here. These authors prepared dark chocolates enriched with freeze-dried (1%) and concentrated (1 and 3%) raspberry leaf extract and evaluated their hedonic response. The dark chocolates enriched with freeze-dried and 1% concentrated extract obtained the highest scores for appearance and texture, whilst chocolates containing 3% extract exhibited the lowest sensory scores for these properties. Likewise, mouthfeel and aftertaste also differed significantly in response to the treatments; the highest scores were observed for chocolates enriched with freeze-dried extract as the low temperature and pressure involved enable a preservation of aroma and taste.

CONCLUSIONS

In the present study, enrichment with a bioactive ingredient, i.e., cinnamaldehyde, imparted significant changes to various sensory attributes (appearance, aroma, mouthfeel, and aftertaste) of developed chocolates. These effects might be due to its intense aroma and strong antioxidant, colouring and flavouring properties. Furthermore, cinnamaldehyde enrichment proved to be efficient during storage and did not exert any detrimental effects on the resulting functional/nutraceutical chocolates. It is worth mentioning that during sensory profiling, all the chocolates attained acceptable scores and achieved superior hedonic responses.

References

- Afoakwa E.O., Paterson A., Fowler M., Vieira J. (2008): Effects of tempering and fat crystallisation behaviour on microstructure, mechanical properties and appearance in dark chocolate systems. *Journal of Food Engineering*, 89: 128–136.
- Albak F., Tekin A.R. (2015): Effect of cinnamon powder addition during conching on the flavor of dark chocolate mass. *Journal Food Science and Technology*, 52: 1960–1970.
- Ali A., Selamat J., Man Y.B.C., Suria A.M. (2001): Effect of storage temperature on texture, polymorphic structure, bloom formation and sensory attributes of filled dark chocolate. *Food Chemistry*, 72: 491–497.
- Al-Jamal A.R. (2009): Effects of cinnamon on blood glucose and lipids levels in diabetic patients (Type1). *African Journal of Biochemistry Research*, 3: 81–184.
- Belščak-Cvitanović A., Komes D., Benković M., Karlović S., Hečimović I., Ježek D., Bauman I. (2012): Innovative formulations of chocolates enriched with plant polyphenols from *Rubus idaeus* L. leaves and characterization of their physical, bioactive and sensory properties. *Food Research International*, 48: 820–830.
- Belščak-Cvitanović A., Komes D., Dujmović M., Karlović S., Biškić M., Brncić M., Jezek D. (2015): Physical, bioactive and sensory quality parameters of reduced sugar chocolates formulated with natural sweeteners as sucrose alternatives. *Food Chemistry*, 167: 61–70.
- Briones V., Aguilera J.M. (2005): Image analysis of changes in surface color of chocolate. *Food Research International*, 38: 87–94.
- Cervellati R., Greco E., Costa S., Guerra M.C., Speroni E. (2008): A comparison of antioxidant properties between artisan-made and factory-produced chocolate. *International Journal of Food Science and Technology*, 43: 1866–1870.
- Cheng D.M., Kuhn P., Pouley A., Rojo L.E., Lila M.A., Raskin I. (2012): *In vivo* and *in vitro* antidiabetic effects of aqueous cinnamon extract and cinnamon polyphenol-enhanced food matrix. *Food Chemistry*, 135: 2994–3002.
- Do T.A.L., Hargreaves J.M., Wolf B., Hort J., Mitchell J.R. (2007): Impact of particle size distribution on rheological and textural properties of chocolate models with reduced fat content. *Journal of Food Science*, 72: 9.
- Dürschmid K., Albrecht U., Schleining G., Kneifel W. (2006): Sensory evaluation of milk chocolates as an instrument of new product development. In: 13th World Congress of Food Science & Technology-Food is Life, Sept 17–21, 2006, Nantes: IUFOST. France: 1331–1332.
- El-Baroty G.S., EL-Baky H.H.A., Farag R.S., Saleh M.A. (2010): Characterization of antioxidant and antimicrobial compounds of cinnamon and ginger essential oils. *African Journal of Biochemistry Research*, 4: 167–174.
- Komes D., Belščak-Cvitanović A., Skrabal S., Vojvodić A., Busic A. (2013): The influence of dried fruits enrichment on sensory properties of bitter and milk chocolates and bioactive content of their extracts affected by different solvents. *LWT-Food Science Technology*, 53: 360–396.
- Lachman J., Hosnedl V., Pivec V., Orsák M. (1998): Polyphenols in cereals and their positive and negative role in human and animal nutrition. In: *Proceedings of Conference Cereals for Human Health and Preventive Nutrition*, July 7, 1998, Brno, Czech Republic: 118–125.
- Lara E., Cortés P., Briones V., Perez M. (2010): Structural and physical modifications of corn biscuits during baking process. *LWT-Food Science and Technology*, 44: 622–630.
- Lee S., Biresaw G., Kinney M.P., Inglett G.E. (2009): Effect of cocoa butter replacement with a β -glucan-rich hydro-

- colloid (C-trim30) on the rheological and tribological properties of chocolates. *Journal of Science of Food and Agriculture*, 89: 163–167.
- Mahfouz M.H., Ghanem M.H., Mohamed M.A. (2010): Modulation of insulin receptor substrate-1 and some inflammatory variables in hyperinsulinemic rats treated with cinnamon extract. *American journal of Biochemistry Biotechnology*, 6: 11–18.
- Mariod A.A., AL-Naqeb G., Ismail M. (2010): *Monechma ciliatum* methanolic extract regulates low density lipoprotein receptor and 3-hydroxy-3-methylglutaryl co-enzyme A reductase genes expression in HepG2 cells. *African Journal of Biotechnology*, 9: 5813–5819.
- Marongiu B., Piras A., Porcedda S., Tveri E., Sanjust E., Meli M., Sollai F., Zucca P., Rescigno A. (2007): Supercritical CO₂ extract of *Cinnamomum zeylanicum*: chemical characterization and antityrosinase activity. *Journal of Agriculture and Food Chemistry*, 55: 10022–10027.
- Mexis S.F., Badeka A.V., Riganakos K.A., Kontominas M.G. (2010): Effect of active and modified atmosphere packaging on quality retention of dark chocolate with hazelnuts. *Innovative Food Science and Emerging Technology*, 11: 177–186.
- Miller K.B., Stuart D.A., Smith N.L., Lee C.Y., Mchale N.L., Flanagan J.A. (2006): Antioxidant activity and polyphenol and procyanidin contents of selected commercially available cocoa-containing and chocolate products in the United States. *Journal of Agriculture and Food Chemistry*, 54: 4062–4068.
- Nightingale L.M.A., Lee S., Engeseth N.J. (2009): Textural changes in chocolate characterized by instrumental and sensory techniques. *Journal of Texture Studies*, 40: 427–444.
- Rossini K., Caciano P.Z., Noreña, Brandelli A. (2011): Changes in the color of white chocolate during storage: potential roles of lipid oxidation and non-enzymatic browning reactions. *Journal of Food Science and Technology*, 48: 305–311.
- Rusconi M., Conti A. (2010): *Theobroma cacao* L. the food of the gods: A scientific approach beyond myths and claims. *Pharmacology Research*, 61: 5–13.
- Sangal A. (2011): Role of cinnamon as beneficial antidiabetic food adjunct: a review. *Advances in Applied Sciences Research*, 2: 440–450.
- Sim S.Y.J., Ng J.W., Ng W.K., Forde C.G., Henry C.J. (2016): Plant polyphenols to enhance the nutritional and sensory properties of chocolates. *Food Chemistry*, 200: 46–54.
- Simi A., Sokovi M.D., Risti M., Gruji-Jovanovi S., Vukojevi J., Marin P.D. (2004): The chemical composition of some *Lauraceae* essential oils and their antifungal activities. *Phytotherapy Research*, 18: 713–717.
- Steel R.G.D., Torrie J.H., Dickey D. (1997): *Principles and Procedures of Statistics: a Biometrical Approach*. 3rd Ed. New York, McGraw Hill Book: 139–145.
- Thamke I., Durrschmid K., Rohm H. (2009): Sensory description of dark chocolates by consumers. *LWT-Food Science Technology*, 42: 534–539.

Received: 2016–06–22

Accepted after corrections: 2017–07–19

Published online: 2018–01–05