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Chinese Yellow Cattle *PPARA* Gene: Analyses of Expression, Polymorphism and Trait Association

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Supplementary Online Material (SOM)

Table S1. Information of primer sequences for the bovine *PPARA* gene

| Name | Primer sequence (5'-3') | Size (bp) | Tm (°C) | Used for |
|--------------------|---|-----------|---------|--------------------------------|
| <i>PPARA</i> -P1 | F: ATACTGCCTTGGACTTCGC R: TAGTCTGTAGGGCAATGGAAGTA | 443 | 57 | partial intron 1 and 2; exon 2 |
| <i>PPARA</i> -P2 | F: CAAGCGGAAGGAGAATGGC R: AAGGCAGAAAGACGCAACC | 494 | 60 | partial intron 2 and 3; exon 3 |
| <i>PPARA</i> -P3 | F: TGAACAAGAAGCGTGAATAAAGG R: CGTTCCAAGCCCACAAGG | 285 | 60 | partial intron 4 and 5; exon 5 |
| <i>PPARA</i> -P4 | F: TCTGACTTCCTGCTGATGTTTCG R: CTGGGAGCGTTGAGGTGG | 561 | 60 | partial intron 5 and 6; exon 6 |
| <i>PPARA</i> -P5 | F: GATGACGCTGACTATCTTTCTGA R: TACCGCTGCTGGGTTCTC | 732 | 60 | partial intron 6 and 7; exon 7 |
| <i>PPARA</i> -P6 | F: TGAAGACTGTTCCCTTGGCGT R: AAAACAGCAAACCTGAACCGAAAC | 947 | 60 | partial intron 7 and 3'utr |
| <i>PPARA</i> -P7 | F: TTACAGCAGACAATCACGGGT R: GCTCCTCCTTTGGGAACGA | 573 | 60.5 | partial promoter region |
| <i>PPARA</i> -P8 | F: GCTCGCCGCCACAAATAGA R: GGGAACGCTGGTTGGGAG | 469 | 63 | partial promoter region |
| <i>PPARA</i> -P9 | F: TCTTGCTGCTCACCATTTG R: CCACCTCCTGTCCCCTG | 692 | 57 | partial promoter region |
| <i>PPARA</i> -qRT | F: GCTCCGTTATTACAGACACCC R: AACCTTGCAGCCCTCAC | 180 | 54 | mRNA expression |
| <i>ACTB</i> -qRT | F: CTGGGCGTAATGGTGGGC R: CTGATGCCGTGCTCAATGG | 107 | 54 | mRNA expression |
| <i>TUBA1A</i> -qRT | F: GGAGGTTTCGCACTGGCAC R: CGCCTTGCCAATGGTGTAG | 112 | 54 | mRNA expression |

2 Table S2. Primers for identifying mutations by PCR-RFLP and ACRS-PCR within the *PPARA* gene

| No. | Locus* | rsID # | Primers (5'-3') | Size (bp) | Tm (°C) | Enzyme | Cleavage sites | Genotype (bp) |
|-----|-----------------|-------------|--|-----------|---------|-----------------|----------------------|---|
| 1 | g-117148558 A>T | rs471506343 | F: TTTCA [□] GTGGGATGTC [□] CCATA [□] R: TTACTTTCTTAGGCTCTCGTGT [□] TAC | 338 | 57.5 | <i>Hpa</i> II | AGGC [□] ^T | TT: 338 TA: 338, 318, 20 AA: 318 |
| 2 | g-117195033 A>G | rs134580633 | F: CAAGCGGAAGGAGAATGGC R: AAGGCAGAAAAGACGCAACC | 494 | 60 | <i>Hinf</i> I | G^ANTC | AA: 494 AG: 494, 388, 106 GG: 388, 106 |
| 3 | g-117195348 A>G | rs135735531 | F: CAAGCGGAAGGAGAATGGC R: AAGGCAGAAAAGACGCAACC | 330 | 60 | <i>Hind</i> III | A^AGCTT | AA: 330, 164 GA: 330, 164, 89, 75 GG: 330, 89, 75 |
| 4 | g-117204210 G>A | rs477982176 | F: AATCCACAGGGTTTCTTT [□] CG R: AATAGGCAGACGGAGGCAT | 201 | 55 | <i>Asu</i> II | TT^CGAA | GG: 201 GA: 201, 183, 18 AA: 183, 18 |
| 5 | g-117204336 T>C | rs137668765 | F: GTCTTCCCTTTT [□] TACCGCTT R: ACCAAAGTCTTCCAAAATAA [□] TAT [□] G | 168 | 53 | <i>Hha</i> I | C^CGG | TT: 168 TC: 168, 146 CC: 146 |
| 6 | g-117228031 T>C | rs110745628 | F: TCCGTGGAGACCG [□] CAC R: TAGGCTACCAACATCCCATCTTTAT | 164 | 57.7 | <i>Hgi</i> C I | G^GYRCC | TT: 164 TC: 164, 150, 15 CC: 150, 15 |
| 7 | g-117228160 T>C | rs446377435 | F: TGTGTCTTCTGTGATGAATAAAG [□] C R: AATGATAGCAGCCACAAAGAGG | 191 | 55 | <i>Hpa</i> II | C^CGG | TT: 191 TC: 191, 166, 25 CC: 166, 25 |
| 8 | g-117232845 T>C | novel | F: CACTACAGAGACAGGAGC [□] AC R: ACAGTCAAAAAGCGGTAAAAGGG | 333 | 60 | <i>Pst</i> I | CTGCA^G | TT: 309 TC: 333, 309 CC: 333 |
| 9 | g-117233248 A>G | rs432147085 | F: CTTCCTTCGCCCTTAT [□] TCAA R: TTGCACATGCTATATAGC [□] G | 147 | 55 | <i>Hae</i> II | GCG^C | AA: 147 AG: 147, 122 |

PCR-RFLP = PCR-restricted fragment length polymorphisms, ACRS-PCR = artificially created restriction site-PCR, Tm = annealing temperature
 # rsID in database of dbSNP in NCBI; * identification of polymorphisms in *PPARA* gene among Chinese cattle according the reference sequence (GenBank No. NC_007303.6)
 Primers with [□] for identifying mutations by ACRS-PCR, [□] represents mismatched bases. Primers without [□] for identifying mutations by PCR-RFLP

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Table S3. Genotype and allelic frequencies, value of χ^2 test, and diversity parameters of the *PPARA* gene analysed in the study

| SNPs | Breeds | Sample | Genotype frequencies | | | Minor allele frequency | HWE <i>P</i> -value* | PIC | H _e | N _e |
|-----------------|--------|--------|----------------------|-------|-------|------------------------|----------------------|-------|----------------|----------------|
| | | | AA | TA | TT | | | | | |
| g.117148558 A>T | JX | 141 | 0.000 | 0.496 | 0.504 | 0.248 | 13.970 | 0.304 | 0.373 | 1.596 |
| | LX | 114 | 0.026 | 0.518 | 0.456 | 0.285 | 7.273 | 0.325 | 0.408 | 1.688 |
| | NY | 139 | 0.129 | 0.403 | 0.468 | 0.331 | 1.132 | 0.345 | 0.443 | 1.795 |
| | QC | 30 | 0.000 | 0.400 | 0.600 | 0.200 | 0.810 | 0.269 | 0.320 | 1.471 |
| g.117195033 A>G | JX | 141 | 0.121 | 0.539 | 0.340 | 0.390 | 2.486 | 0.363 | 0.476 | 1.908 |
| | LX | 114 | 0.105 | 0.763 | 0.132 | 0.487 | 31.706 | 0.375 | 0.500 | 1.999 |
| | NY | 139 | 0.187 | 0.597 | 0.216 | 0.486 | 5.298 | 0.375 | 0.500 | 1.998 |
| | QC | 30 | 0.033 | 0.900 | 0.067 | 0.483 | 16.970 | 0.375 | 0.499 | 1.998 |
| g.117195348 A>G | JX | 141 | 0.326 | 0.489 | 0.184 | 0.429 | 0.0154 | 0.370 | 0.490 | 1.961 |
| | LX | 114 | 0.044 | 0.763 | 0.193 | 0.425 | 35.882 | 0.369 | 0.489 | 1.957 |
| | NY | 139 | 0.108 | 0.561 | 0.331 | 0.389 | 4.556 | 0.362 | 0.475 | 1.905 |
| | QC | 30 | 0.033 | 0.933 | 0.033 | 0.500 | 20.017 | 0.375 | 0.500 | 2.000 |
| g.117204210 G>A | JX | 141 | 0.639 | 0.308 | 0.053 | 0.207 | 0.483 | 0.274 | 0.328 | 1.488 |
| | LX | 114 | 0.708 | 0.236 | 0.057 | 0.175 | 3.491 | 0.247 | 0.288 | 1.405 |
| | NY | 139 | 0.682 | 0.296 | 0.023 | 0.171 | 0.067 | 0.243 | 0.283 | 1.394 |
| | QC | 30 | 0.793 | 0.138 | 0.069 | 0.138 | 2.504 | 0.210 | 0.238 | 1.312 |
| g.117204336 T>C | JX | 141 | 0.319 | 0.553 | 0.128 | 0.404 | 3.109 | 0.366 | 0.482 | 1.929 |
| | LX | 114 | 0.491 | 0.430 | 0.079 | 0.294 | 0.145 | 0.329 | 0.415 | 1.709 |
| | NY | 139 | 0.425 | 0.482 | 0.093 | 0.334 | 0.948 | 0.346 | 0.445 | 1.803 |
| | QC | 30 | 0.300 | 0.500 | 0.200 | 0.450 | 0.003 | 0.373 | 0.495 | 1.980 |
| g.117228031 T>C | JX | 141 | – | 0.066 | 0.934 | 0.033 | 0.845 | 0.062 | 0.064 | 1.068 |
| | LX | 114 | – | 0.055 | 0.946 | 0.027 | 2.158 | 0.052 | 0.053 | 1.056 |
| | NY | 139 | – | 0.061 | 0.939 | 0.030 | 1.193 | 0.057 | 0.059 | 1.062 |
| | QC | 30 | – | 0.069 | 0.931 | 0.035 | 6.389 | 0.064 | 0.067 | 1.071 |
| g.117228160 T>C | JX | 141 | 0.698 | 0.198 | 0.103 | 0.202 | 18.718 | 0.271 | 0.323 | 1.477 |
| | LX | 114 | 0.709 | 0.209 | 0.081 | 0.186 | 11.616 | 0.257 | 0.302 | 1.433 |
| | NY | 139 | 0.640 | 0.270 | 0.090 | 0.225 | 5.099 | 0.288 | 0.349 | 1.536 |
| | QC | 30 | 0.778 | 0.185 | 0.037 | 0.129 | 0.063 | 0.200 | 0.228 | 1.291 |
| g.117232845 T>C | JX | 141 | 0.858 | 0.121 | 0.021 | 0.082 | 3.244 | 0.139 | 0.150 | 1.176 |
| | LX | 114 | 0.684 | 0.281 | 0.035 | 0.175 | 0.007 | 0.248 | 0.289 | 1.407 |
| | NY | 139 | 0.734 | 0.216 | 0.050 | 0.158 | 5.017 | 0.231 | 0.266 | 1.363 |
| | QC | 30 | 0.833 | 0.167 | 0.000 | 0.083 | 0.413 | 0.141 | 0.153 | 1.180 |
| g.117233248 A>G | JX | 141 | 0.078 | 0.922 | – | 0.461 | 100.569 | 0.374 | 0.497 | 1.988 |
| | LX | 114 | 0.167 | 0.833 | – | 0.417 | 55.962 | 0.368 | 0.486 | 1.946 |
| | NY | 139 | 0.252 | 0.748 | – | 0.374 | 47.724 | 0.359 | 0.468 | 1.881 |
| | QC | 30 | 0.433 | 0.567 | – | 0.283 | 3.278 | 0.324 | 0.406 | 1.684 |

SNP = single nucleotide polymorphism, HWE = Hardy–Weinberg equilibrium, PIC = polymorphism information content, H_e = heterozygosity, N_e = effective allele numbers, JX = Jiaxian, LX = Luxi, NY = Nanyang, QC = Qinchuan

*values in bold indicate that the genotype distribution was not in agreement with the HWE ($P < 0.05$)