

# Nanopesticides: Current status and scope for their application in agriculture

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Table S1. Efficacy of nanoemulsion and nanosuspension/nanodispersion formulations against insect-pests and pathogens

Nanoformulation/carrier	Active ingredient	Attributes	Effective against	References
<b>Nanoemulsions</b>				
Lecithin and Tween 80	oregano and thyme essential oil	increased efficacy	<i>Sitophilus oryzae</i>	Hossain et al. 2019
Span 80, Tween 80 and lecithin	cinnamon essential oil and thiabendazole	reduced fruit decay	<i>Rhizopus stolonifera</i> and <i>Botrytis cinerea</i>	Yousef et al. 2019
Tween 80 and lecithin	pyrethrum	enhanced activity	<i>Myzus persicae</i>	Pascual-Villalobos et al. 2019
Alkyd resin	$\lambda$ -cyhalothrin	increased water stability	insects	Qin et al. 2017
Tween 80 and Span 20	clove essential oil	improved antifungal property	<i>Phytophthora palmivora</i>	Rattanapet and Aht-Ong 2019
Palm kernel oil ester	parthenium hysterophorus crude extract	herbicidal activity	<i>Diodia ocimifolia</i>	Zainuddin et al. 2019
Pluronic F-127, clove oil, MW 12600 and HLB 22	sulphonamides	$\beta$ -carbonic anhydrase inhibition	<i>Leishmania infantum</i> and <i>Leishmania amazonensis</i>	Cardoso et al. 2018
MCT oil and Tween 80	cinnamon oil	increased stability	<i>Microbial pathogens</i>	Chuesiang et al. 2018
Tween 80	<i>Pimpinella anisum</i> essential oil	enhanced stability and efficiency	<i>Tribolium castaneum</i>	Hashem et al. 2018
Span 85, Brij 97 and ethylene glycol	citral	increased stability and extended activity	bacteria	Lu et al. 2018
Tween 80	<i>Satureja hortensis</i> essential oil	enhanced bioherbicidal activity	<i>Chenopodium album</i> and <i>Amaranthus retroflexus</i>	Hazrati et al. 2017
Gelatin-chitosan, Tween 20 and Span 60	canola oil, $\alpha$ -tocopherol, cinnamaldehyde and garlic oil	high antioxidant activity	<i>Pseudomonas aeruginosa</i>	Pérez-Córdoba et al. 2018
Glycerol, Tween 80 and Agnique BL1754	tebuconazole	increased stability	–	Díaz-Blancas et al. 2016
Tween 20	<i>Simmondsia chinensis</i> oil	high mortality	<i>Sitophilus oryzae</i>	Abouelkassem et al. 2016
Lemon essential oil, surfactant, Tween 20 and glycerol	pyrethrin	dose reduction	<i>Aphis gossypii</i>	Kalaitzaki et al. 2015
Octyldodecyl myristate and polysorbate 80	<i>Manilkara subsericea</i> crude extract	non-target safety	<i>Dysdercus peruvianus</i>	Fernandes et al. 2014
Aqueous filtrate of <i>Pongamia glabra</i> and <i>Jatropha curcas</i>	<i>Eucalyptus globules</i> oil	enhanced toxicity	<i>Tribolium castaneum</i>	Pant et al. 2014
<i>N</i> -butyl acetate, ammonium glycyrrhizinate and sec-butyl alcohol	permethrin	greater efficacy and reduced effects on non-target organism	<i>Aedes aegypti</i>	Kumar et al. 2013
Tween 20	neem oil	enhanced toxicity	<i>Culex quinquefasciatus</i>	Anjali et al. 2012
Fatty acid methyl esters, SAPG, LAPG and organosilicone	glyphosate isopropylamine	enhanced leaf wettability, penetration and toxicity	<i>Eleusine indica</i>	Jiang et al. 2012
Methyl decanoate and polyoxyethylene lauryl ether	$\beta$ -cypermethrin	increased stability	–	Wang et al. 2007
<b>Nanosuspension/nanodispersion</b>				
Polycarboxylate and MRES and sucrose	abamectin	enhanced bioavailability	<i>Plutella xylostella</i>	Cui et al. 2019

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Table S1 to be continued

Nanoformulation/carrier	Active ingredient	Attributes	Effective against	References
Emulsifier 700	lambda-cyhalothrin	reduced amount of surfactants	<i>Lipaphis erysami</i>	Wang et al. 2019a
Ethyl acetate, emulsifier 1601, emulsifier 600 and lactose	lambda-cyhalothrin	enhanced stability	<i>Myzus persicae</i>	Wang et al. 2019b
Sodium alginate and Tween 80	pyridalyl	enhanced toxicity	<i>Helicoverpa armigera</i>	Saini et al. 2014
<i>N</i> -butyl acetate, ammonium glycyrrhizinate, sec-butyl alcohol and sucrose	permethrin	reduced dose	<i>Culex quinquefasciatus</i>	Anjali et al. 2010
Isobutyl acetate, isopropanol and surfactants	novaluron	nanosize and high efficiency	<i>Spodoptera littoralis</i>	Elek et al. 2010
Cyclohexanone and <i>N,N</i> -dimethylformamide	$\beta$ -cypermethrin	high drug loading capacity and controlled release	–	Zeng et al. 2008

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Table S2. Efficacy of polymer-based pesticide nanoformulations against insect-pests, pathogens and weeds

Nanoformulation/carrier	Active ingredient(s)	Attributes	Target organism(s)	References
<b>Polymer nanoencapsulations</b>				
Methoxy poly(ethylene glycol)- <i>b</i> -poly(lactide- <i>co</i> -glycolide)	metolachlor	water solubility, sustained release	<i>Oryza sativa</i> and <i>Digitaria sanguinalis</i>	Tong et al. 2017
Poly( $\epsilon$ -caprolactone)	carbendazim and tebuconazole	sustained release	–	Campos et al. 2015
Silica–epichlorohydrin– <i>co</i> -carboxymethylcellulose	emamectin benzoate	enzyme responsive with reduced genotoxicity	<i>Myzus persicae</i>	Guo et al. 2015
Sodium alginate and chitosan	acetamipirid	controlled release	–	Kumar et al. 2015
Poly(lactic- <i>co</i> -glycolic acid)	imidacloprid	reduced dose with enhanced effectivity	<i>Diaphorina citri</i>	Meyer et al. 2015
Poly(ethylene glycol) 6 000	temephos and imidacloprid	enhanced toxicity and reduced dose	<i>Culex quinquefasciatus</i>	Bhan et al. 2014
Poly( $\epsilon$ -caprolactone), Tween 80 and Span 60	ametryn and atrazine	reduced dose	weeds	Clemente et al. 2014
Poly( $\epsilon$ -caprolactone) and chitosan	atrazine	improved adhesion and slow release	–	Grillo et al. 2014
Polydopamine	avermectin	sustained and controlled release	–	Jia et al. 2014
Sodium alginate	imidacloprid	better efficacy	<i>Amrasca</i> spp.	Kumar et al. 2014
Poly(citric acid) and poly(ethylene glycol)	imidacloprid	increased release rate and better performance	<i>Glyphodes pyloalis</i>	Memarizadeh 2014; Naeini et al. 2010
Poly( $\epsilon$ -caprolactone)	atrazine	target specific with increased toxicity	<i>Brassica</i> spp.	Pereira et al. 2014
Carbon nanotubes with hyperbranched polycitric acid	mancozeb and zineb	increased contact area	<i>Alternaria alternate</i>	Sarlak et al. 2014
Carboxymethyl chitosan	methomyl	reduced UV-degradation	<i>Mythimna separata</i>	Sun et al. 2014
Poly(ethylene glycol) 400	acephate	increased controlled and targeted delivery	<i>Oligonychus coffeae</i> , <i>Spodoptera litura</i> , <i>Lipaphis erysimi</i> and <i>Bemisia tabaci</i>	Pradhan et al. 2013
Poly(ethylene glycol) 400	acephate	water solubility and reduced toxicity	–	Choudhury et al. 2012
Polyethylene glycol and polybutylene adipate	<i>Onopordon leptolepis</i> extract	controlled release	bacteria and fungi	Esmaili and Saremnia 2012
Poly( $\epsilon$ -caprolactone) and chitosan	ametryn, atrazine, and simazine	increased stability and controlled release	<i>Allium cepa</i>	Grillo et al. 2012
Methyl methacrylate-styrene	lansiumamide B	better efficacy	<i>Bursaphelenchus xylophilus</i> and <i>Meloidogyne incognita</i>	Yin et al. 2012
Poly(ethylene glycol) 6 000	<i>Allium sativum</i> essential oil	sustained release	<i>Tribolium castaneum</i>	Yang et al. 2009
Sodium alginate and chitosan	imidacloprid	increased toxicity	<i>Martianus dermestoides</i>	Guan et al. 2008
<b>Nanospheres</b>				
Attapulgate, NH <sub>4</sub> HCO <sub>3</sub> , amino silicon oil, poly(vinyl alcohol) and glyphosate	glyphosate	temperature responsive controlled release	weeds	Chi et al. 2017
Polyvinyl alcohol and polyoxyethylene castor oil (EL-40)	emamectin benzoate	slow release and reduced photo degradation	insects	Wang et al. 2017
Poly( $\epsilon$ -caprolactone)	azadirachtin	reduced UV-degradation	<i>Zabrotes subfasciatus</i>	da Costa et al. 2014

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Table S2 to be continued

Nanoformulation/carrier	Active ingredient(s)	Attributes	Target organism(s)	References
Poly(ethylene glycol) 600, 1 000, 1 500 and 2 000	thiram	controlled release	fungi	Kaushik et al. 2013
Poly(ethylene glycol) 600 and 900	carbofuran	controlled release	<i>Meloidogyne incognita</i>	Pankaj et al. 2012
Poly(ethylene glycol) 1 000, 2 000 and 4 000	thiamethoxam	controlled release	insects	Sarkar et al. 2012
Gelatin and methyl methacrylate	tebuconazole	reduced leaching losses	<i>Gloeophyllum trabeum</i>	Salma et al. 2010
Poly(acrylic acid)- <i>b</i> -poly(butyl acrylate) and poly(styrene)- <i>b</i> -poly(ethyleneoxide)	bifenthrin	increased stability	insects	Liu et al. 2008
<b>Nanogels</b>				
Poly(ethylene glycol)-poly(lactic acid)-dimethacrylate	small interfering RNA (siRNA)	sustained release	–	Wang et al. 2018
Carboxymethyl cellulose, citric acid and bentonite	thiamethoxam	higher release rate	insects	Sarkar and Singh 2017
$\beta$ -cyclodextrin	permethrin	reduced biodegradation	<i>Tineola bisselliella</i> and <i>Anthrenocerus australis</i>	Kettel et al. 2014
Myristic acid-chitosan	<i>Carum copticum</i> essential oil	increased toxicity and persistence	<i>Sitophilus garnarius</i> and <i>Tribolium confusum</i>	Ziaee et al. 2014
Low molecular mass gelators	methyl eugenol	reduced volatilization losses and extended release	<i>Bactrocera dorsalis</i>	Bhagat et al. 2013
Chitosan	copper (ii)	high loading and synergistic effect	<i>Fusarium graminearum</i>	Brunel et al. 2013
Poly( $\epsilon$ -caprolactone), Span 60 and Tween 80	<i>Azadirachta indica</i> extract	enhanced toxicity	<i>Plutella xylostella</i>	Forim et al. 2013
Chitosan and cashew gum	<i>Lippia sidoides</i> oil	enhanced loading capacity and sustained release	<i>Stegomyia aegypti</i>	Abreu et al. 2012
Polyacrylamide methylcellulose	paraquat	extended release	weeds	Aouada et al. 2011
Chitosan and cashew gum	<i>Lippia sidoides</i> oil	increased efficiency	<i>Aedes aegypti</i>	Paula et al. 2011
Sodium alginate	isoproturon, imidacloprid, and cyromazine	extended release	insects and weeds	Garrido-Herrera et al. 2006
<b>Micelles</b>				
Carboxymethyl chitosan and 2-nitrobenzyl	diuron	photo controlled sustained release	weeds	Ye et al. 2015
Chitin with polylactide and 1,2-dipalmitoyl- <i>sn</i> -glycero-3-phosphoethanolamine	chlorpyrifos	increased loading capacity and sustained release	insects	Zhang et al. 2013
Poly(ethylene glycol) 300, 600, 1 000 and dicarboxylic acids	imidacloprid	controlled and sustained release	insects and nematodes	Adak et al. 2012
Ricinoleic acid-grafted CMCS	azadirachtin	enhanced toxicity	insects	Feng and Peng 2012
Poly(ethylene glycol) 1 500 and 2 000	$\beta$ -cyfluthrin	slow release and sustained control	<i>Callosobruchus maculatus</i>	Loha et al. 2012
Chitosan with polylactide	imidacloprid	increased loading capacity and sustained release	insects	Li et al. 2011
Poly-[poly-(oxyethylene 600/1 000/1 500/2 000)-oxyisophthaloyl]	$\beta$ -cyfluthrin	controlled release	insects	Loha et al. 2011

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Table S2 to be continued

Nanoformulation/carrier	Active ingredient(s)	Attributes	Target organism(s)	References
Poly(ethylene glycol) 600, 900, 1 500 and 1 800	azadirachtin A	controlled release	insects	Kumar et al. 2010
<i>N</i> -(octadecanol-1-glycidyl ether)- <i>O</i> -sulfate chitosan	rotenone	increased solubility	insects	Lao et al. 2010
Poly[poly(oxyethylene 18 000)-oxy-5-decanyloxyiso-phthaloyl]	carbofuran	slow and sustained release	insects and nematodes	Shakil et al. 2010
<b>Nanofibers</b>				
Poly(lactic acid)/cellulose	thiamethoxam	controlled release	<i>Trialeurodes vaporariorum</i>	Xiang et al. 2013
Polyamide 6 and cellulose acetate	( <i>Z</i> )-9-dodecenyl acetate	extended release	lepidopterous moths	Hellmann et al. 2011
<b>Chitosan nanoparticles based formulations</b>				
Chitosan and laminarian	–	downy mildew control	<i>Plasmopara viticola</i>	Garde-Cerdán et al. 2017
Chitosan	imazapic and imazapyr	reduced toxicity	weeds	Maruyama et al. 2016
Chitosan and tripolyphosphate	Cu(II) ion	decreased disease severity	<i>Alternaria solani</i> and <i>Fusarium oxysporum</i>	Saharan et al. 2013
Chitosan	pyrifluquinazon	high efficiency and no non-target toxicity	<i>Myzus persicae</i>	Kang et al. 2012
Chitosan	saponin	spore germination inhibition	<i>Alternaria alternata</i>	Lanzotti et al. 2012a,b
Chitosan	capsaicin	high loading efficiency and controlled release	–	Feng and Zhang 2011
Chitosan	etofenprox	controlled release	<i>Spodoptera litura</i>	Hwang et al. 2011

Table S3. Efficacy of lipid-based pesticide nanoformulations against insect-pests and pathogens

Nanoformulation/carrier	Active ingredient	Attributes	Effective against	References
Glyceryl tripalmitate	carbendazim and tebuconazole	physical stability and controlled release	–	Campos et al. 2015
Glycerol tripalmitate	atrazine and simazine	controlled release profile	<i>Raphanus raphanistrum</i>	de Oliveira et al. 2015
Nanoliposomes	<i>Eucalyptus citriodora</i> oil	stimuli responsive release	<i>Staphylococcus aureus</i>	Lin et al. 2015
Poly(butyl meth-acrylate-diacetone acrylamide)	acetochlor	slow release	–	Guo et al. 2014
Corn oil and beeswax	deltamethrin	decreased photodegradation	–	Nguyen et al. 2012
Liposomes	etofenprox	controlled release	–	Bang et al. 2009
Compritol 888 ATO and Poloxamer 188 or Miranol Ultra C32	<i>Artemisia arborescens</i> essential oil	physical stability and controlled release	<i>Bemisia tabaci</i> , <i>Aphis gossipy</i> and <i>Lymantria dispar</i>	Lai et al. 2006

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Table S4. Efficacy of clay-based pesticide nanoformulations against insect-pests, pathogens and weeds

Nanoformulation/carrier	Active ingredient	Attributes	Effective against	References
Modified clay nanoparticles	alachlor, acetochlor, metolachlor and metolachlor	enhanced adsorption	weeds	Rodrigues et al. 2013
Zinc-aluminum-layered double hydroxide	3,4-D	slow and extended release	weeds	Ghazali et al. 2013
Modified montmorillonite clay	ethofumesate	enhanced biodegradation and slow release	weeds	Chevillard et al. 2012
Nanoclay	thymol	extended release	insects and bacteria	Lim et al. 2010
Layered double hydroxide	cinnamate	slow and extended release	<i>Phytophthora capsici</i>	Park et al. 2010
Bentonite, kaolinite and Fuller's earth	carbofuran	extended release	–	Choudhary et al. 2006
Arizona montmorillonite clay	2,4-D	slow release	–	Hermosin et al. 2006
Layered double hydroxide	2,4-D	controlled release	weeds	Bin Hussein et al. 2005
Wyoming and arizona montmorillonite clays	hexazinone	reduced leaching losses	–	Celis et al. 2002

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Table S5. Efficacy of silica-based pesticide nanoformulations against insect-pests pathogens and weeds

Nanoformulation/carrier	Active ingredient	Attributes	Effective against	References
Mesoporous silica NPs	2,4-D sodium salt	enhanced pesticide loading	–	Cao et al. 2018
Silica microcapsules	kasugamycin	improved photo-thermal stability and enhanced loading efficiency	–	Fan et al. 2017
Mesoporous silica NPs	chlorantraniliprole	enhanced pesticide loading	<i>Plutella xylostella</i>	Kaziem et al. 2017
Silica cross-linked microcapsules	pendimethalin	extended release	<i>Amaranthus retroflexus</i> and <i>Echinochloa crusgalli</i>	Liang et al. 2017
Silica nanostructures	neem oil	enhanced stability and high antioxidant capacity	<i>Acromyrmex crassispinus</i>	Mattos et al. 2017
Modified silica NPs	chitinase (Chi9602)	enhanced nematocidal activity	<i>Caenorhabditis elegans</i>	Qin et al. 2016
Amino-modified silica	kasugamycin	reduced photodegradation	<i>E. coli</i>	Ding et al. 2014
Silica NPs	–	higher disease resistance in maize	<i>Fusarium oxysporum</i> and <i>Aspergillus niger</i>	Suriyaprabha et al. 2013
Silica NPs	linalool	enhanced antifeedant potential	<i>Spodoptera litura</i> and <i>Achaea janata</i>	Usha Rani et al. 2014
Silica nanocapsules	fipronil	controlled release	termites	Wibowo et al. 2014
Silica-based microcapsules	prochloraz	reduced UV degradation and slow release	fungal diseases	Zhang et al. 2014
Nanonized diatomaceous earth	–	malformation in mouthparts and mid-gut morphology of the insects	<i>Heteracris littoralis</i>	Ebeid et al. 2013
Nanosilica	–	antifeedant	<i>Spodoptera littoralis</i>	El-Bendary and El-Helaly 2013
Mesoporous silica NPs	metalaxyl	controlled release in soil	–	Wanyika 2013
Mesoporous silica NPs	imidacloprid	controlled release	termites	Popat et al. 2012
Silica NPs	chlorfenapyr	increased biological efficacy	<i>Plutella xylostella</i>	Song et al. 2012
Amorphous hydrophilic, hydrophobic and lipophilic silica NPs	–	increased effectiveness	<i>Sitophilus oryzae</i>	Debnath et al. 2011; Rahman et al. 2009; Barik et al. 2008
Carboxyl-modified mesoporous silica	2,4-D and picloram	controlled release	crop weeds	Prado et al. 2011
Amorphous nanosilica	–	enhanced cuticular absorption	<i>Sitophilus oryzae</i> , <i>Tribolium castaneum</i> , <i>Spodoptera litura</i> and <i>Lipaphis pseudobrassicae</i>	Goswami et al. 2010
Surface charged modified hydrophobic silica	–	decrease fungal growth and enhanced germination of cereal seeds	many agricultural pests	Robinson and Salejova-Zadrazilova 2010; Ulrichs et al. 2006
Porous hollow silica NPs	avermectin	protection from UV degradation and slow release	-	Li et al. 2007
Porous hollow silica NPs	validamycin	sustained release profile	bacteria and fungi	Liu et al. 2006
Lipophilic silica-based	–	enhanced physiosorption through cuticle	agricultural insect pests	Ulrichs et al. 2005; Mewis and Ulrichs 2001

NPs – nanoparticles



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Table S6. Efficacy of metal nanoparticles (NPs) as active ingredients and their nanoformulations against insect-pests and pathogens

Nanoformulation/carrier	Active ingredient(s)	Attributes	Target organism(s)	References
<b>Silver nanoparticles (AgNPs)</b>				
AgNPs	–	fungal germination and growth inhibition	<i>Sclerotinia sclerotiorum</i>	Guilger et al. 2017
AgNPs	cyhalothrin	increased potency and delivery efficiency	<i>Culex pipiens</i>	Abouelkassem et al. 2016
Ag-dsDNA-GO	–	antibacterial activity	<i>Xanthomonas euvesicatoria</i> , <i>X. gardneri</i> , <i>X. perforans</i> , and <i>X. vesicatoria</i>	Strayer et al. 2016
AgNPs	–	antiviral agent	Bean Yellow Mosaic Virus	Elbeshehy et al. 2015
AgNPs	<i>Spatoglossum asperum</i> and <i>Hedophyllum sessile</i> extracts	antibacterial activity	<i>Xanthomonas axonopodis</i> pv. <i>citri</i> and <i>X. oryzae</i> pv. <i>oryzae</i>	Jothirethinam et al. 2015
Spherical to polyhedral shaped AgNPs	–	larvicidal effect	<i>Aedes aegypti</i>	Vimala et al. 2015
Spherical AgNPs	–	reduction in disease severity	<i>Bipolaris sorokiniana</i>	Mishra et al. 2014
AgNPs	–	larvicidal and pupicidal activity	<i>Aedes aegypti</i>	Sundaravadivelan and Padmanabhan 2014
AgNPs	–	antifungal activity	<i>Fusarium oxysporum</i>	Gopinath and Velusamy 2013
AgNPs	–	antibacterial and antifungal activity	<i>Pseudomonas flavescens</i> , <i>Erwinia amylovora</i> , <i>Fusarium solani</i> , <i>F. oxysporum</i> , <i>Rhizoctonia solani</i> , <i>Helminthosporium</i> spp., <i>Alternaria alternate</i> and <i>Sclerotinia sclerotiorum</i>	Hafez and Kabeil 2013
DNA-directed AgNPs grown on graphene oxide	–	reduction in severity of bacterial spot disease	<i>Xanthomonas perforans</i>	Ocsoy et al. 2013
AgNPs	–	antibacterial activity	18 plant pathogens	Kim et al. 2012
Silver and gold NPs	–	high mortality	<i>Aedes aegypti</i>	Soni and Prakash 2012
Colloidal silver NPs	–	delayed growth of fungi	<i>Colletotrichum gloeosporioides</i>	Aguilar-Méndez et al. 2011
AgNPs aqueous liquids	–	antifungal activity	<i>Sclerotium cepivorum</i>	Jung et al. 2010
AgNPs	–	reduced spore germination	<i>Fusarium culmorum</i>	Kasprowicz et al. 2010
AgNPs	–	antimicrobial activity	<i>Vibrio cholera</i> and <i>Escherichia coli</i>	Krishnaraj et al. 2010
AgNPs	–	high antimicrobial activity	<i>Bipolaris sorokiniana</i> and <i>Magnaporthe grisea</i>	Jo et al. 2009
AgNPs	–	antifungal activity	<i>Raffaelea</i> spp.	Kim et al. 2009
AgNPs	–	growth inhibition	<i>Rhizoctonia solani</i> , <i>Sclerotinia sclerotiorum</i> and <i>S. minor</i>	Min et al. 2009
AgNPs colloidal solution	–	antifungal properties	<i>Podospaera pannosa</i>	Kim et al. 2008
<b>Copper nanoparticles (CuNPs)</b>				
Cu <sub>2</sub> ONPs	–	antifungal activity	<i>Saccharomyces cerevisiae</i>	Giannousi et al. 2014

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Table S6 to be continued

Nanoformulation/carrier	Active ingredient(s)	Attributes	Target organism(s)	References
CuNPs	–	novel antifungal activity	<i>Curvularia lunata</i> , <i>Phoma destructive</i> and <i>Alternaria alternata</i>	Kanhed et al. 2014
CuNPs	–	antibacterial properties	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Mondal and Mani 2012
CuNPs	–	antimicrobial activity	<i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , <i>Micrococcus luteus</i> , <i>Klebsiella pneumoniae</i> , <i>Pseudomonas aeruginosa</i> , <i>Aspergillus flavus</i> , <i>A. niger</i> , <i>Candida albicans</i>	Ramyadevi et al. 2012
CuNPs	–	herbicidal properties	<i>Elodea densa</i>	Nekrasova et al. 2011
CuNPs	–	antibacterial properties	<i>Xanthomonas campestris</i> pv. <i>phaseoli</i> and <i>Xanthomonas oryzae</i> pv. <i>oryzae</i>	Mondal et al. 2010; Mondal and Mani 2009
Spherical CuNPs	–	antimicrobial activity	<i>Escherichia coli</i> , <i>Bacillus subtilis</i> and <i>Staphylococcus aureus</i>	Ruparelia et al. 2008
<b>Titanium dioxide nanoparticles (TiO<sub>2</sub>NPs)</b>				
Ag doped hollow TiO <sub>2</sub> nanoparticles	–	enhanced fungicidal activity	<i>Fusarium solani</i> and <i>Venturia inaequalis</i>	Boxi et al. 2016
TiO <sub>2</sub> NPs and TiO <sub>2</sub> NPs doped with zinc and silver	–	antibacterial activity	<i>Xanthomonas perforans</i>	Paret et al. 2013a,b
TiO <sub>2</sub> NPs	–	antibacterial activity	<i>Xanthomonas hortorum</i> pv. <i>pelargonii</i> and <i>Xanthomonas axonopodis</i> pv. <i>poinsetticola</i>	Norman and Chen 2011
<b>Other metal and non-metal nanoparticles</b>				
Zinc oxide nanoparticles (ZnONPs)	thiram	synergistic antifungal activity	<i>Phytophthora capsici</i>	Xue et al. 2014
Nano-sulphur	–	increased efficacy against powdery mildew	<i>Erysiphe cichoracearum</i>	Gogoi et al. 2013
ZnONPs	–	prevention of conidiophores and conidial development and death of fungal hyphae	<i>Botrytis cinerea</i> and <i>Penicillium expansum</i>	He et al. 2011
Nano-sized calcium carbonate	validamycin	enhanced loading efficiency, stability and sustained release	<i>Rhizoctonia solani</i>	Qian et al. 2011

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## REFERENCES

- Abouelkassem S., El-Borady O.M., Mohamed M.B. (2016): Remarkable enhancement of cyhalothrin upon loading into silver nanoparticles as larvicidal. *International Journal of Contemporary Applied Sciences*, 3: 252–264.
- Abreu F.O., Oliveira E.F., Paula H.C., de Paula R.C. (2012): Chitosan/cashew gum nanogels for essential oil encapsulation. *Carbohydrate Polymers*, 89: 1277–1282.
- Adak T., Kumar J., Shakil N.A., Walia S. (2012): Development of controlled release formulations of imidacloprid employing novel nano-ranged amphiphilic polymers. *Journal of Environmental Science and Health, Part B. Pesticides*, 47: 217–225.
- Aguilar-Méndez M.A., Martín-Martínez E.S., Ortega-Arroyo L., Cobián-Portillo G., Sánchez-Espíndola E. (2011): Synthesis and characterization of silver nanoparticles: Effect on phytopathogen *Colletotrichum gloeosporioides*. *Journal of Nanoparticle Research*, 13: 2525–2532.
- Anjali C.H., Khan S.S., Margulis-Goshen K., Magdassi S., Mukherjee A., Chandrasekaran N. (2010): Formulation of water-dispersible nanopermethrin for larvicidal applications. *Ecotoxicology and Environmental Safety*, 73: 1932–1936.
- Anjali C.H., Sharma Y., Mukherjee A., Chandrasekaran N. (2012): Neem oil (*Azadirachta indica*) nanoemulsion as potent larvicidal agent against *Culex quinquefasciatus*. *Pest Management Science*, 68: 158–163.
- Aouada F.A., de Moura M.R., Capparelli Mattoso L.H. (2011): Biodegradable hydrogel as delivery vehicle for the controlled release of pesticide. In: Stoytcheva M. (ed.): *Pesticides: Formulations, Effects, Fate*. Rijeka, InTech Publication: 81–102.
- Bang S.H., Yu Y.M., Hwang I.C., Park H.J. (2009): Formation of size-controlled nano carrier systems by self-assembly. *Journal of Microencapsulation*, 26: 722–733.
- Barik T.K., Sahu B., Swain V. (2008): Nanosilica—from medicine to pest control. *Parasitology Research*, 103: 253. doi: 10.1007/s00436-008-0975-7
- Bhagat D., Samanta S.K., Bhattacharya S. (2013): Efficient management of fruit pests by pheromone nanogels. *Scientific Reports*, 3: 1294. doi: 10.1038/srep01294
- Bhan S., Mohan L., Srivastava C.N. (2014): Relative larvicidal potentiality of nano-encapsulated Temephos and Imidacloprid against *Culex quinquefasciatus*. *Journal of Asia-Pacific Entomology*, 17: 787–791.
- Bin Hussein M.Z., Yahaya A.H., Zainal Z., Kian L.H. (2005): Nanocomposite-based controlled release formulation of an herbicide, 2,4-dichlorophenoxyacetate encapsulated in zinc–aluminium-layered double hydroxide. *Science and Technology of Advanced Materials*, 6: 956–962.
- Boxi S.S., Mukherjee K., Paria S. (2016): Ag doped hollow TiO<sub>2</sub> nanoparticles as an effective green fungicide against *Fusarium solani* and *Venturia inaequalis* phytopathogens. *Nanotechnology*, 27: 085103. doi: 10.1088/0957-4484/27/8/085103
- Brunel F., El Gueddari N.E., Moerschbacher B.M. (2013): Complexation of copper (II) with chitosan nanogels. Toward control of microbial growth. *Carbohydrate Polymer*, 92: 1348–1356.
- Campos E.V.R., de Oliveira J.L., da Silva C.M.G., Pascoli M., Pasquoto T., Lima R., Abhilash P.C., Fraceto L.F. (2015): Polymeric and solid lipid nanoparticles for sustained release of carbendazim and tebuconazole in agricultural applications. *Scientific Reports*, 5: 13809. doi: 10.1038/srep13809
- Cao L., Zhou Z., Niu S., Cao C., Li X., Shan Y., Huang Q. (2018): Positive-charge functionalized mesoporous silica nanoparticles as nanocarriers for controlled 2,4-dichlorophenoxy acetic acid sodium salt release. *Journal of Agricultural and Food Chemistry*, 66: 6594–6603.
- Cardoso da Silva V., Vermelho A.B., Ricci Junior E., Rodrigues I.A., Mazotto A.M., Supuran C.T. (2018): Antileishmanial activity of sulphonamide nanoemulsions targeting the  $\beta$ -carbonic anhydrase from *Leishmania species*. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 33: 850–857.
- Celis R., Hermosin M.C., Carrizosa M.J., Cornejo J. (2002): Inorganic and organic clays as carriers for controlled release of the herbicide hexazinone. *Journal of Agricultural and Food Chemistry*, 50: 2324–2330.
- Chevillard A., Angellier-Coussy H., Guillard V., Gontard N., Gastaldi E. (2012): Investigating the biodegradation pattern of an ecofriendly pesticide delivery system based on wheat gluten and organically modified montmorillonites. *Polymer Degradation and Stability*, 97: 2060–2068.
- Chi Y., Zhang G., Xiang Y., Cai D., Wu Z. (2017): Fabrication of a temperature-controlled release herbicide using a nanocomposite. *ACS Sustainable Chemistry and Engineering*, 5: 4969–4975.
- Choudhary G., Kumar J., Walia S., Parsad R., Parmar B.S. (2006): Development of controlled release formulations of carbofuran and evaluation of their efficacy against *Meloidogyne incognita*. *Journal of Agricultural and Food Chemistry*, 54: 4727–4733.
- Choudhury S.R., Pradhan S., Goswami A. (2012): Preparation and characterization of acephate nano-encapsulated complex. *Nanoscience Methods*, 1: 9–15.
- Chuesiang P., Siripatrawan U., Sanguandeeikul R., McLandsborough L., McClements D.J. (2018): Optimization of cinnamon oil nanoemulsions using Phase Inversion Temperature method: Impact of oil phase composition and surfactant concentration. *Journal of Colloid and Interface Science*, 514: 208–216.
- Clemente Z., Grillo R., Jonsson M., Santos N.Z., Feitosa L.O., Lima R., Fraceto L.F. (2014): Ecotoxicological evaluation

<https://doi.org/10.17221/102/2020-PPS>

- of poly(epsilon-caprolactone) nanocapsules containing triazine herbicides. *Journal of Nanoscience and Nanotechnology*, 14: 4911–4917.
- Cui B., Lv Y., Gao F., Wang C., Zeng Z., Wang Y., Sun C., Zhao X., Shen Y., Liu G., Cui H. (2019): Improving abamectin bioavailability via nanosuspension constructed by wet milling technique. *Pest Management Science*, 75: 2756–2764.
- da Costa J.T., Forim M.R., Costa E.S., De Souza J.R., Mondego J.M., Junior A.L.B. (2014): Effects of different formulations of neem oil-based products on control *Zabrotes subfasciatus* (Boheman, 1833) (Coleoptera: Bruchidae) on beans. *Journal of Stored Products Research*, 56: 49–53.
- de Oliveira J.L., Campos E.V.R., da Silva C.M.G., Pasquoto T., Lima R., Fraceto L.F. (2015): Solid lipid nanoparticles co-loaded with simazine and atrazine: Preparation, characterization, and evaluation of herbicidal activity. *Journal of Agricultural and Food Chemistry*, 63: 422–432.
- Debnath N., Das S., Seth D., Chandra R., Bhattacharya S.C., Goswami A. (2011): Entomotoxic effect of silica nanoparticles against *Sitophilus oryzae* (L.). *Journal of Pest Science*, 84: 99–105.
- Díaz-Blancas V., Medina D.I., Padilla-Ortega E., Bortolini-Zavala R., Olvera-Romero M., Luna-Bárceñas G. (2016): Nanoemulsion formulations of fungicide tebuconazole for agricultural applications. *Molecules*, 21: 1271. doi: 10.3390/molecules21101271
- Ding G.L., Li D.G., Liu Y., Guo M.C., Duan Y.H., Li J.Q., Cao Y.S. (2014): Preparation and characterization of kasuga-silica-conjugated nanospheres for sustained antimicrobial activity. *Journal of Nanoparticle Research*, 16: 2671. doi: 10.1007/s11051-014-2671-0
- Ebeid A.R., Abdel A.A., Gesraha M.A. (2013): Impact of diatomaceous earth (silica nano particles) on alfalfa grasshopper, *Heteracris littoralis* (Rambur) (Orthoptera: Acrididae) under laboratory conditions. *Egyptian Journal of Biological Pest Control*, 23: 325–330.
- El-Bendary H.M., El-Helaly A.A. (2013): First record nanotechnology in agricultural: Silica nano-particles a potential new insecticide for pest control. *Applied Science Reports*, 4: 241–246.
- Elbeshehy E.K.F., Elazzazy A.M., Aggelis G. (2015): Silver nanoparticles synthesis mediated by new isolates of *Bacillus spp.*, nanoparticle characterization and their activity against Bean Yellow Mosaic Virus and human pathogens. *Frontiers in Microbiology*, 6: 453. doi: 10.3389/fmicb.2015.00453
- Elek N., Hoffman R., Raviv U., Resh R., Ishaaya I., Magdassi S. (2010): Novaluron nanoparticles: Formation and potential use in controlling agricultural insect pests. *Colloids and Surfaces A: Physicochemical and Engineering*, 372: 66–72.
- Esmaeili A., Saremnia B. (2012): Preparation of extract-loaded nanocapsules from *Onopordon leptolepis* DC. *Industrial Crops and Products*, 37: 259–263.
- Fan C., Guo M., Liang Y., Dong H., Ding G., Zhang W., Tang G., Yang J., Kong D., Cao Y. (2017): Pectin-conjugated silica microcapsules as dual-responsive carriers for increasing the stability and antimicrobial efficacy of kasugamycin. *Carbohydrate Polymers*, 172: 322–331.
- Feng B.H., Peng L.F. (2012): Synthesis and characterization of carboxymethyl chitosan carrying ricinoleic functions as an emulsifier for azadirachtin. *Carbohydrate Polymers*, 88: 576–582.
- Feng B.H., Zhang Z.Y. (2011): Carboxymethyl chitosan grafted ricinoleic acid group for nanopesticide carriers. *Advanced Materials Research*, 236–238: 1783–1788.
- Fernandes C.P., de Almeida F.B., Silveira A.N., Gonzalez M.S., Mello C.B., Feder D., Apolinário R., Santos M.G., Carvalho J.C.T., Tietbohl L.A.C., Rocha L., Falcão D.Q. (2014): Development of an insecticidal nanoemulsion with *Manilkara subsericea* (Sapotaceae) extract. *Journal of Nanotechnology*, 12: 22. doi: 10.1186/1477-3155-12-22
- Forim M.R., Costa E.S., Da Silva M.F.D.G.F., Fernandes J.B., Mondego J.M., Boiça Junior A.L. (2013): Development of a new method to prepare nano-/microparticles loaded with extracts of *Azadirachta indica*, their characterization and use in controlling *Plutella xylostella*. *Journal of Agricultural and Food Chemistry*, 61: 9131–9139.
- Garde-Cerdán T., Mancini V., Carrasco-Quiroz M., Servili A., Gutiérrez-Gamboa G., Foglia R., Pérez-Álvarez E.P., Romanazzi G. (2017): Chitosan and laminarin as alternatives to copper for *Plasmopara viticola* control: Effect on grape amino acid. *Journal of Agricultural and Food Chemistry*, 65: 7379–7386.
- Garrido-Herrera F.J., Gonzalez-Pradas E., Fernández-Pérez M. (2006): Controlled release of isoprotruron, imidacloprid, and cyromazine from alginate-bentonite-activated carbon formulations. *Journal of Agricultural and Food Chemistry*, 54: 10053–10060.
- Ghazali S.A.I.S.M., Hussein M.Z., Sarijo S.H. (2013): 3,4-Dichlorophenoxyacetate interleaved into anionic clay for controlled release formulation of a new environmentally friendly agrochemical. *Nanoscale Research Letters*, 8: 362. doi: 10.1186/1556-276X-8-362
- Giannousi K., Sarafidis G., Mourdikoudis S., Pantazaki A., Dendrinou-Samara C. (2014): Selective synthesis of Cu<sub>2</sub>O and Cu/Cu<sub>2</sub>O NPs: Antifungal activity to yeast *Saccharomyces cerevisiae* and DNA interaction. *Inorganic Chemistry*, 53: 9657–9666.
- Gogoi R., Singh P.K., Kumar R., Nair K.K., Alam I., Srivastava C., Yadav S., Gopal M., Choudhury S.R., Goswami A. (2013): Suitability of nano-sulphur for biorational management

<https://doi.org/10.17221/102/2020-PPS>

- of powdery mildew of okra (*Abelmoschus esculentus* Moench) caused by *Erysiphe cichoracearum*. *Journal of Plant Pathology and Microbiology*, 4: 171. doi: 10.4172/2157-7471.1000171
- Gopinath V., Velusamy P. (2013): Extracellular biosynthesis of silver nanoparticles using *Bacillus* sp. GP-23 and evaluation of their antifungal activity towards *Fusarium oxysporum*. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 106: 170–174.
- Goswami A., Roy I., Sengupta S., Debnath N. (2010): Novel applications of solid and liquid formulations of nanoparticles against insect pests and pathogens. *Thin Solid Films*, 519: 1252–1257.
- Grillo R., dos Santos N.Z.P., Maruyama C.R., Rosa A.H., de Lima R., Fraceto L.F. (2012): Poly( $\epsilon$ -caprolactone) nanocapsules as carrier systems for herbicides: Physicochemical characterization and genotoxicity evaluation. *Journal of Hazardous Materials*, 231–232: 1–9.
- Grillo R., Pereira A.E., Nishisaka C.S., De Lima R., Oehlke K., Greiner R., Fraceto L.F. (2014): Chitosan/tripolyphosphate nanoparticles loaded with paraquat herbicide: An environmentally safer alternative for weed control. *Journal of Hazardous Materials*, 278: 163–171.
- Guan H., Chi D., Yu J., Li X. (2008): A novel photodegradable insecticide: Preparation, characterization and properties evaluation of nano-imidacloprid. *Pesticide Biochemistry and Physiology*, 92: 83–91.
- Guilger M., Pasquoto-Stigliani T., Bilesky-Jose N., Grillo R., Abhilash P.C., Fraceto L.F., De Lima R. (2017): Biogenic silver nanoparticles based on *Trichoderma harzianum*: Synthesis, characterization, toxicity evaluation and biological activity. *Scientific Reports*, 7: 44421. doi: 10.1038/srep44421
- Guo Y., Yang Q., Yan W., Li B., Qian K., Li T., Xiao W., He L. (2014): Controlled release of acetochlor from poly (butyl methacrylate-diacetone acrylamide) based formulation prepared by nanoemulsion polymerisation method and evaluation of the efficacy. *International Journal of Environmental Analytical Chemistry*, 94: 1001–1012.
- Guo M., Zhang W., Ding G., Guo D., Zhu J., Wang B., Punyapitak D., Cao Y. (2015): Preparation and characterization of enzyme-responsive emamectin benzoate microcapsules based on a copolymer matrix of silica–epichlorohydrin–carboxymethylcellulose. *RSC Advances*, 5: 93170–93179.
- Hafez E.E., Kabeil S.S. (2013): Antimicrobial activity of nano-silver particles produced by micro algae. *Journal of Pure and Applied Microbiology*, 7: 35–42.
- Hashem A.S., Awadalla S.S., Zayed G.M., Maggi F., Benelli G. (2018): *Pimpinella anisum* essential oil nanoemulsions against *Tribolium castaneum*-insecticidal activity and mode of action. *Environmental Science and Pollution Research*, 25: 18802–18812.
- Hazrati H., Saharkhiz M.J., Niakousari M., Moein M. (2017): Natural herbicide activity of *Satureja hortensis* L. essential oil nanoemulsion on the seed germination and morpho-physiological features of two important weed species. *Ecotoxicology and Environmental Safety*, 142: 423–430.
- He L., Liu Y., Mustapha A., Lin M. (2011): Antifungal activity of zinc oxide nanoparticles against *Botrytis cinerea* and *Penicillium expansum*. *Microbiological Research*, 166: 207–215.
- Hellmann C., Greiner A., Wendorff J.H. (2011): Design of pheromone releasing nanofibers for plant protection. *Polymers for Advanced Technologies*, 22: 407–413.
- Hermosin M.C., Celis R., Facenda G., Carrizosa M.J., Ortega-Calvo J.J., Cornejo J. (2006): Bioavailability of the herbicide 2,4-D formulated with organoclays. *Soil Biology and Biochemistry*, 38: 2117–2124.
- Hossain F., Follett P., Salmieri S., Vu K.D., Harich M., Lacroix M. (2019): Synergistic effects of nanocomposite films containing essential oil nanoemulsions in combination with ionizing radiation for control of rice weevil *Sitophilus oryzae* in stored grains. *Journal of Food Science*, 84: 1439–1446.
- Hwang I., Kim T., Bang S., Kim K., Kwon H., Seo M., Youn Y., Park H., Yasunaga-Aoki C., Yu Y. (2011): Insecticidal effect of controlled release formulations of etofenprox based on nano-bio technique. *Journal of the Faculty of Agriculture, Kyushu University*, 56: 33–40.
- Jia X., Sheng W.B., Li W., Tong Y.B., Liu Z.Y., Zhou F. (2014): Adhesive polydopamine coated avermectin microcapsules for prolonging foliar pesticide retention. *ACS Applied Materials and Interfaces*, 6: 19552–19558.
- Jiang L.C., Basri M., Omar D., Rahman M.B.A., Salleh A.B., Rahman R.N.Z.R.A., Selamat A. (2012): Green nano-emulsion intervention for water-soluble glyphosate isopropylamine (IPA) formulations in controlling *Eleusine indica* (*E. indica*). *Pesticide Biochemistry and Physiology*, 102: 19–29.
- Jo Y.K., Kim B.H., Jung G. (2009): Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. *Plant Disease*, 93: 1037–1043.
- Jothirethinam A., Prathiba S., Shanthi N., Arunkumar K. (2015): Green synthesized silver nanoparticles prepared from the antimicrobial crude extracts of two brown seaweeds against plant pathogens. *American Journal of Nanotechnology*, 6: 31. doi: 10.3844/ajntsp.2015.31.39
- Jung J.H., Kim S.W., Min J.S., Kim Y.J., Lamsal K., Kim K.S., Lee Y.S. (2010): The effect of nano-silver liquid against the white rot of the green onion caused by *Sclerotium cepivorum*. *Mycobiology*, 38: 39–45.
- Kalaitzaki A., Papanikolaou N.E., Karamaouna F., Dourtoglou V., Xenakis A., Papadimitriou V. (2015): Biocompatible colloidal dispersions as potential formulations of natural pyrethrins: A structural and efficacy study. *Langmuir*, 31: 5722–5730.

<https://doi.org/10.17221/102/2020-PPS>

- Kang M.A., Seo M.J., Hwang I.C., Jang C., Park H.J., Yu Y.M., Youn Y.N. (2012): Insecticidal activity and feeding behavior of the green peach aphid, *Myzus persicae*, after treatment with nano types of pyrifluquinazon. *Journal of Asia-Pacific Entomology*, 15: 533–541.
- Kanhed P., Birla S., Gaikwad S., Gade A., Seabra A.B., Rubilar O., Duran N., Rai M. (2014): *In vitro* antifungal efficacy of copper nanoparticles against selected crop pathogenic fungi. *Materials Letters*, 115: 13–17.
- Kasprowicz M.J., Kozioł M., Gorczyca A. (2010): The effect of silver nanoparticles on phytopathogenic spores of *Fusarium culmorum*. *Canadian Journal of Microbiology*, 56: 247–253.
- Kaushik P., Shakil N.A., Kumar J., Singh M.K., Singh M.K., Yadav S.K. (2013): Development of controlled release formulations of thiram employing amphiphilic polymers and their bioefficacy evaluation in seed quality enhancement studies. *Journal of Environmental Science and Health, Part B*, 48: 677–685.
- Kaziem A.E., Gao Y., He S., Li J. (2017): Synthesis and insecticidal activity of enzyme-triggered functionalized hollow mesoporous silica for controlled release. *Journal of Agricultural and Food Chemistry*, 65: 7854–7864.
- Kettel M.J., Schaefer K., Groll J., Moeller M. (2014): Nanogels with high active  $\beta$ -cyclodextrin content as physical coating system with sustained release properties. *ACS Applied Materials and Interfaces*, 6: 2300–2311.
- Kim H.S., Kang H.S., Chu G.J., Byun H.S. (2008): Antifungal effectiveness of nanosilver colloid against rose powdery mildew in greenhouses. In: Rhee C.K. (ed.): *Solid State Phenomena*. Kapellweg, Trans Tech Publications Ltd.: 15–18.
- Kim S.W., Kim K.S., Lamsal K., Kim Y.J., Kim S.B., Jung M.Y., Sim S.J., Kim H.S., Chang S.J., Kim J.K., Lee Y.S. (2009): An *in vitro* study of the antifungal effect of silver nanoparticles on oak wilt pathogen *Raffaelea* sp. *Journal of Microbiology and Biotechnology*, 19: 760–764.
- Kim S.W., Jung J.H., Lamsal K., Kim Y.S., Min J.S., Lee Y.S. (2012): Antifungal effects of silver nanoparticles (AgNPs) against various plant pathogenic fungi. *Mycobiology*, 40: 53–58.
- Krishnaraj C., Jagan E.G., Rajasekar S., Selvakumar P., Kalichelvan P.T., Mohan N.J.C.S.B.B. (2010): Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens. *Colloids and Surfaces B: Biointerfaces*, 76: 50–56.
- Kumar J., Shakil N.A., Singh M.K., Pankaj, Singh M.K., Pandey A., Pandey R.P. (2010): Development of controlled release formulations of azadirachtin-A employing poly (ethylene glycol) based amphiphilic copolymers. *Journal of Environmental Science and Health Part B*, 45: 310–314.
- Kumar R.S., Shiny P.J., Anjali C.H., Jerobin J., Goshen K.M., Magdassi S., Mukherjee A., Chandrasekaran N. (2013): Distinctive effects of nano-sized permethrin in the environment. *Environmental Science and Pollution Research*, 20: 2593–2602.
- Kumar S., Bhanjana G., Sharma A., Sidhu M.C., Dilbaghi N. (2014): Synthesis, characterization and on field evaluation of pesticide loaded sodium alginate nanoparticles. *Carbohydrate Polymers*, 101: 1061–1067.
- Kumar S., Chauhan N., Gopal M., Kumar R., Dilbaghi N. (2015): Development and evaluation of alginate-chitosan nanocapsules for controlled release of acetamiprid. *International Journal of Biological Macromolecules*, 81: 631–637.
- Lai F., Wissing S.A., Müller R.H., Fadda A.M. (2006): *Artemisia arborescens* L essential oil-loaded solid lipid nanoparticles for potential agricultural application: preparation and characterization. *AAPS PharmSciTech*, 7: E10. doi: 10.1208/pt070102
- Lanzotti V., Barile E., Antignani V., Bonanomi G., Scala F. (2012a): Antifungal saponins from bulbs of garlic, *Allium sativum* L. var. Voghiera. *Phytochemistry*, 78: 126–134.
- Lanzotti V., Romano A., Lanzuise S., Bonanomi G., Scala F. (2012b): Antifungal saponins from bulbs of white onion, *Allium cepa* L. *Phytochemistry*, 74: 133–139.
- Lao S.B., Zhang Z.X., Xu H.H., Jiang G.B. (2010): Novel amphiphilic chitosan derivatives: Synthesis, characterization and micellar solubilization of rotenone. *Carbohydrate Polymers*, 82: 1136–1142.
- Li Z.Z., Chen J.F., Liu F., Liu A.Q., Wang Q., Sun H.Y., Wen L.X. (2007): Study of UV-shielding properties of novel porous hollow silica nanoparticle carriers for avermectin. *Pest Management Science*, 63: 241–246.
- Li M., Huang Q., Wu Y. (2011): A novel chitosan-poly(lactide) copolymer and its submicron particles as imidacloprid carriers. *Pest Management Science*, 67: 831–836.
- Liang Y., Guo M., Fan C., Dong H., Ding G., Zhang W., Tang G., Yang J., Kong D., Cao Y. (2017): Development of novel urease-responsive pendimethalin microcapsules using silica-IPTS-PEI as controlled release carrier materials. *ACS Sustainable Chemistry and Engineering*, 5: 4802–4810.
- Lim G.O., Jang S.A., Song K.B. (2010): Physical and antimicrobial properties of *Gelidium corneum*/nano-clay composite film containing grapefruit seed extract or thymol. *Journal of Food Engineering*, 98: 415–420.
- Lin L., Cui H., Zhou H., Zhang X., Bortolini C., Chen M., Dong M. (2015): Nanoliposomes containing *Eucalyptus citriodora* as antibiotic with specific antimicrobial activity. *Chemical Communications*, 51: 2653–2655.
- Liu F., Wen L.X., Li Z.Z., Yu W., Sun H.Y., Chen J.F. (2006): Porous hollow silica nanoparticles as controlled delivery system for water-soluble pesticide. *Materials Research Bulletin*, 41: 2268–2275.

<https://doi.org/10.17221/102/2020-PPS>

- Liu Y., Tong Z., Prud'homme R.K. (2008): Stabilized polymeric nanoparticles for controlled and efficient release of bifenthrin. *Pest Management Science*, 64: 808–812.
- Loha K.M., Shakil N.A., Kumar J., Singh M.K., Adak T., Jain S. (2011): Release kinetics of  $\beta$ -Cyfluthrin from its encapsulated formulations in water. *Journal of Environmental Science and Health Part B*, 46: 201–206.
- Loha K.M., Shakil N.A., Kumar J., Singh M.K., Srivastava C. (2012): Bio-efficacy evaluation of nanoformulations of  $\beta$ -cyfluthrin against *Callosobruchus maculatus* (Coleoptera: Bruchidae). *Journal of Environmental Science and Health, Part B*, 47: 687–691.
- Lu W.C., Huang D.W., Wang C.C., Yeh C.H., Tsai J.C., Huang Y.T., Li P.H. (2018): Preparation, characterization, and antimicrobial activity of nanoemulsions incorporating citral essential oil. *Journal of Food and Drug Analysis*, 26: 82–89.
- Maruyama C.R., Guilger M., Pascoli M., Bileshy-José N., Abhilash P.C., Fraceto L.F., De Lima R. (2016): Nanoparticles based on chitosan as carriers for the combined herbicides imazapic and imazapyr. *Scientific Reports*, 6: 19768. doi: 10.1038/srep19768
- Mattos B.D., Rojas O.J., Magalhães W.L. (2017): Biogenic silica nanoparticles loaded with neem bark extract as green, slow-release biocide. *Journal of Cleaner Production*, 142: 4206–4213.
- Memarizadeh N., Ghadamyari M., Adeli M., Talebi K. (2014): Preparation, characterization and efficiency of nanoencapsulated imidacloprid under laboratory conditions. *Ecotoxicology and Environmental Safety*, 107: 77–83.
- Mewis I., Ulrichs C. (2001): Action of amorphous diatomaceous earth against different stages of the stored product pests *Tribolium confusum*, *Tenebrio molitor*, *Sitophilus granarius* and *Plodia interpunctella*. *Journal of Stored Products Research*, 37: 153–164.
- Meyer W.L., Gurman P., Stelinski L.L., Elman N.M. (2015): Functional nano-dispensers (FNDs) for delivery of insecticides against phytopathogen vectors. *Green Chemistry*, 17: 4173–4177.
- Min J.S., Kim K.S., Kim S.W., Jung J.H., Lamsal K., Kim S.B., Jung M.Y., Lee Y.S. (2009): Effects of colloidal silver nanoparticles on sclerotium-forming phytopathogenic fungi. *Plant Pathology Journal*, 25: 376–380.
- Mishra S., Singh B.R., Singh A., Keswani C., Naqvi A.H., Singh H.B. (2014): Biofabricated silver nanoparticles act as a strong fungicide against *Bipolaris sorokiniana* causing spot blotch disease in wheat. *PLoS One*, 9: e97881. doi: 10.1371/journal.pone.0097881
- Mondal K.K., Mani C. (2009): Suppression of common bacterial blight in mungbean by phylloplane resident *Pseudomonas fluorescens* strain MBPF-01 alone and in combination with nanocopper. *Indian Phytopathology*, 62: 445–448.
- Mondal K.K., Mani C. (2012): Investigation of the antibacterial properties of nanocopper against *Xanthomonas axonopodis* pv. *punicae*, the incitant of pomegranate bacterial blight. *Annals of Microbiology*, 62: 889–893.
- Mondal K.K., Bhar L.M., Mani C. (2010): Combined efficacy of *Pseudomonas fluorescens* strain MBPF-01 and nanocopper against bacterial leaf blight in rice. *Indian Phytopathology*, 63: 266–268.
- Naeini A.T., Adeli M., Vossoughi M. (2010): Poly(citric acid)-block-poly(ethylene glycol) copolymers – New biocompatible hybrid materials for nanomedicine. *Nanomedicine: Nanotechnology, Biology and Medicine*, 6: 556–562.
- Nekrasova G.F., Ushakova O.S., Ermakov A.E., Uimin M.A., Byzov I.V. (2011): Effects of copper (II) ions and copper oxide nanoparticles on *Elodea densa* Planch. *Russian Journal of Ecology*, 42: 458. doi: 10.1134/S1067413611060117
- Nguyen H.M., Hwang I.C., Park J.W., Park H.J. (2012): Enhanced payload and photo-protection for pesticides using nanostructured lipid carriers with corn oil as liquid lipid. *Journal of Microencapsulation*, 29: 596–604.
- Norman D.J., Chen J. (2011): Effect of foliar application of titanium dioxide on bacterial blight of geranium and *Xanthomonas* leaf spot of poinsettia. *HortScience*, 46: 426–428.
- Ocsoy I., Paret M.L., Ocsoy M.A., Kunwar S., Chen T., You M., Tan W. (2013): Nanotechnology in plant disease management: DNA-directed silver nanoparticles on graphene oxide as an antibacterial against *Xanthomonas perforans*. *ACS Nano*, 7: 8972–8980.
- Pankaj, Shakil N.A., Kumar J., Singh M.K., Singh K. (2012): Bioefficacy evaluation of controlled release formulations based on amphiphilic nano-polymer of carbofuran against *Meloidogyne incognita* infecting tomato. *Journal of Environmental Science and Health, Part B*, 47: 520–528.
- Pant M., Dubey S., Patanjali P.K., Naik S.N., Sharma S. (2014): Insecticidal activity of eucalyptus oil nanoemulsion with karanja and jatropha aqueous filtrates. *International Biodeterioration and Biodegradation*, 91: 119–127.
- Paret M.L., Vallad G.E., Averett D.R., Jones J.B., Olson S.M. (2013a): Photocatalysis: Effect of light-activated nanoscale formulations of TiO<sub>2</sub> on *Xanthomonas perforans* and control of bacterial spot of tomato. *Phytopathology*, 103: 228–236.
- Paret M.L., Palmateer A.J., Knox G.W. (2013b): Evaluation of a light-activated nanoparticle formulation of titanium dioxide with zinc for management of bacterial leaf spot on rosa 'Noare'. *HortScience*, 48: 189–192.
- Park M., Lee C.I., Seo Y.J., Woo S.R., Shin D., Choi J. (2010): Hybridization of the natural antibiotic, cinnamic acid, with layered double hydroxides (LDH) as green pesticide. *Environmental Science and Pollution Research*, 17: 203–209.

<https://doi.org/10.17221/102/2020-PPS>

- Pascual-Villalobos M.J., Guirao P., Díaz-Baños F.G., Cantó-Tejero M., Villora G. (2019): Oil in water nanoemulsion formulations of botanical active substances. In: Koul O. (ed.): Nano-Biopesticides Today and Future Perspectives. Cambridge, Academic Press: 223–247.
- Paula H.C., Sombra F.M., de Freitas Cavalcante R., Abreu F.O., de Paula R.C. (2011): Preparation and characterization of chitosan/cashew gum beads loaded with *Lippia sidoides* essential oil. *Materials Science and Engineering: C*, 31: 173–178.
- Pereira A.E., Grillo R., Mello N.F., Rosa A.H., Fraceto L.F. (2014): Application of poly(epsilon-caprolactone) nanoparticles containing atrazine herbicide as an alternative technique to control weeds and reduce damage to the environment. *Journal of Hazardous Materials*, 268: 207–215.
- Pérez-Córdoba L.J., Norton I.T., Batchelor H.K., Gkatzionis K., Spyropoulos F., Sobral P.J. (2018): Physico-chemical, antimicrobial and antioxidant properties of gelatin-chitosan based films loaded with nanoemulsions encapsulating active compounds. *Food Hydrocolloids*, 79: 544–559.
- Popat A., Liu J., Hu Q., Kennedy M., Peters B., Lu G.Q.M., Qiao S.Z. (2012): Adsorption and release of biocides with mesoporous silica nanoparticles. *Nanoscale*, 4: 970–975.
- Pradhan S., Roy I., Lodh G., Patra P., Choudhury S.R., Samanta A., Goswami A. (2013): Entomotoxicity and biosafety assessment of PEGylated acephate nanoparticles: A biologically safe alternative to neurotoxic pesticides. *Journal of Environmental Science and Health, Part B*, 48: 559–569.
- Prado A.G., Moura A.O., Nunes A.R. (2011): Nanosized silica modified with carboxylic acid as support for controlled release of herbicides. *Journal of Agricultural and Food Chemistry*, 59: 8847–8852.
- Qian K., Shi T., Tang T., Zhang S., Liu X., Cao Y. (2011): Preparation and characterization of nano-sized calcium carbonate as controlled release pesticide carrier for validamycin against *Rhizoctonia solani*. *Microchimica Acta*, 173: 51–57.
- Qin X., Xiang X., Sun X., Ni H., Li L. (2016): Preparation of nanoscale *Bacillus thuringiensis* chitinases using silica nanoparticles for nematicide delivery. *International Journal of Biological Macromolecules*, 82: 13–21.
- Qin H., Zhang H., Li L., Zhou X., Li J., Kan C. (2017): Preparation and properties of lambda-cyhalothrin/polyurethane drug-loaded nanoemulsions. *RSC Advances*, 7: 52684–52693.
- Rahman A., Seth D., Mukhopadhyaya S.K., Brahmachary R.L., Ulrichs C., Goswami A. (2009): Surface functionalized amorphous nanosilica and microsilica with nanopores as promising tools in biomedicine. *Naturwissenschaften*, 96: 31–38.
- Ramyadevi J., Jeyasubramanian K., Marikani A., Rajakumar G., Rahuman A.A. (2012): Synthesis and antimicrobial activity of copper nanoparticles. *Materials Letters*, 71: 114–116.
- Rattanapet P., Aht-Ong D. (2019): Preparation of coating materials with antifungal property from nanocellulose and clove essential oil nanoemulsions. In: PACCON 2019 – Pure and Applied Chemistry International Conference, Feb 7–8, 2019, Bangkok, Thailand: PO5–PO10.
- Robinson D.K.R., Salejova-Zadrazilova G. (2010): Nanotechnologies for nutrient and biocide delivery in agricultural production. Working Paper Version: 285–297. Available at <http://www.observatorynano.eu/project/filesystem/files/Controlled%20delivery.pdf>.
- Rodrigues L.A.S., Figueiras A., Veiga F., de Freitas R.M., Nunes L.C., da Silva F.E., da Silva L.C. (2013): The systems containing clays and clay minerals from modified drug release: A review. *Colloids and Surfaces B: Biointerfaces*, 103: 642–651.
- Ruparelia J.P., Chatterjee A.K., Duttagupta S.P., Mukherji S. (2008): Strain specificity in antimicrobial activity of silver and copper nanoparticles. *Acta Biomaterialia*, 4: 707–716.
- Saharan V., Mehrotra A., Khatik R., Rawal P., Sharma S.S., Pal A. (2013): Synthesis of chitosan based nanoparticles and their *in vitro* evaluation against phytopathogenic fungi. *International Journal of Biological Macromolecules*, 62: 677–683.
- Saini P., Gopal M., Kumar R., Srivastava C. (2014): Development of pyridalyl nanocapsule suspension for efficient management of tomato fruit and shoot borer (*Helicoverpa armigera*). *Journal of Environmental Science and Health, Part B*, 49: 344–351.
- Salma U., Chen N., Richter D.L., Filson P.B., Dawson-Andoh B., Matuana L., Heiden P. (2010): Amphiphilic core/shell nanoparticles to reduce biocide leaching from treated wood, 1 – Leaching and biological efficacy. *Macromolecular Materials and Engineering*, 295: 442–450.
- Sarkar D.J., Singh A. (2017): Base triggered release of insecticide from bentonite reinforced citric acid crosslinked carboxymethyl cellulose hydrogel composites. *Carbohydrate Polymers*, 156: 303–311.
- Sarkar D.J., Kumar J., Shakil N.A., Walia S. (2012): Release kinetics of controlled release formulations of thiamethoxam employing nano-ranged amphiphilic PEG and diacid based block polymers in soil. *Journal of Environmental Science and Health, Part A*, 47: 1701–1712.
- Sarlak N., Taherifar A., Salehi F. (2014): Synthesis of nanopesticides by encapsulating pesticide nanoparticles using functionalized carbon nanotubes and application of new nanocomposite for plant disease treatment. *Journal of Agricultural and Food Chemistry*, 62: 4833–4838.
- Shakil N.A., Singh M.K., Pandey A., Kumar J., Pankaj, Parmar V.S., Singh M.K., Pandey R.P., Watterson A.C. (2010): Development of poly (ethylene glycol) based amphiphilic copolymers for controlled release delivery of carbofuran.



<https://doi.org/10.17221/102/2020-PPS>

- Journal of Macromolecular Science, Part A: Pure and Applied Chemistry, 47: 241–247.
- Song M.R., Cui S.M., Gao F., Liu Y.R., Fan C.L., Lei T.Q., Liu D.C. (2012): Dispersible silica nanoparticles as carrier for enhanced bioactivity of chlorfenapyr. *Journal of Pesticide Science*, 37: 258–260.
- Soni N., Prakash S. (2012): Efficacy of fungus mediated silver and gold nanoparticles against *Aedes aegypti* larvae. *Parasitology Research*, 110: 175–184.
- Strayer A., Ocoy I., Tan W., Jones J.B., Paret M.L. (2016): Low concentrations of a silver-based nanocomposite to manage bacterial spot of tomato in the greenhouse. *Plant Disease*, 100: 1460–1465.
- Sun D., Hussain H.I., Yi Z., Siegele R., Cresswell T., Kong L., Cahill D.M. (2014): Uptake and cellular distribution, in four plant species, of fluorescently labeled mesoporous silica nanoparticles. *Plant Cell Reports*, 33: 1389–1402.
- Sundaravadivelan C., Padmanabhan M.N. (2014): Effect of mycosynthesized silver nanoparticles from filtrate of *Trichoderma harzianum* against larvae and pupa of dengue vector *Aedes aegypti* L. *Environmental Science and Pollution Research*, 21: 4624–4633.
- Suriyaprabha R., Karunakaran G., Kavitha K., Yuvakkumar R., Rajendran V., Kannan N. (2013): Application of silica nanoparticles in maize to enhance fungal resistance. *IET Nanobiotechnology*, 8: 133–137.
- Tong Y., Wu Y., Zhao C., Xu Y., Lu J., Xiang S., Zong F., Wu X. (2017): Polymeric nanoparticles as a metolachlor carrier: Water-based formulation for hydrophobic pesticides and absorption by plants. *Journal of Agricultural and Food Chemistry*, 65: 7371–7378.
- Ulrichs C., Mewis I., Goswami A. (2005): Crop diversification aiming nutritional security in West Bengal: Biotechnology of stinging capsules in nature's water-blooms. *Annual Tech Issue of State Agri Technologists Service Association*: 1–18.
- Ulrichs C., Krause F., Rocksch T., Goswami A., Mewis I. (2006): Electrostatic application of inert silica dust based insecticides onto plant surfaces. *Communications in Agricultural and Applied Biological Sciences*, 71: 171–178.
- Usha Rani P., Madhusudhanamurthy J., Sreedhar B. (2014): Dynamic adsorption of  $\alpha$ -pinene and linalool on silica nanoparticles for enhanced antifeedant activity against agricultural pests. *Journal of Pest Science*, 87: 191–200.
- Vimala R.T.V., Sathishkumar G., Sivaramakrishnan S. (2015): Optimization of reaction conditions to fabricate nano-silver using *Couroupita guianensis* Aubl. (leaf & fruit) and its enhanced larvicidal effect. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 135: 110–115.
- Wang L., Li X., Zhang G., Dong J., Eastoe J. (2007): Oil-in-water nanoemulsions for pesticide formulations. *Journal of Colloid and Interface Science*, 314: 230–235.
- Wang Y., Wang A., Wang C., Cui B., Sun C., Zhao X., Zeng Z., Shen Y., Gao F., Liu G., Cui H. (2017): Synthesis and characterization of emamectin-benzoate slow-release microspheres with different surfactants. *Scientific Reports*, 7: 12761. doi: 10.1038/s41598-017-12724-6
- Wang Y., Zhang S., Benoit D.S. (2018): Degradable poly(ethylene glycol) (PEG)-based hydrogels for spatiotemporal control of siRNA/nanoparticle delivery. *Journal of Controlled Release*, 287: 58–66.
- Wang C., Cui B., Guo L., Wang A., Zhao X., Wang Y., Liu G. (2019a): Fabrication and evaluation of lambda-cyhalothrin nanosuspension by one-step melt emulsification technique. *Nanomaterials*, 9: 145. doi: 10.3390/nano9020145
- Wang C., Cui B., Zhao X., Wang Y., Zeng Z., Sun C., Yang D., Liu G., Cui H. (2019b): Optimization and characterization of lambda-cyhalothrin solid nanodispersion by self-dispersing method. *Pest Management Science*, 75: 380–389.
- Wanyika H. (2013): Sustained release of fungicide metalaxyl by mesoporous silica nanospheres. In: Diallo M.S., Fromer N.A., Jhon M.S. (eds): *Nanotechnology for Sustainable Development*. Cham, Springer: 321–329.
- Wibowo D., Zhao C.X., Peters B.C., Middelberg A.P. (2014): Sustained release of fipronil insecticide *in vitro* and *in vivo* from biocompatible silica nanocapsules. *Journal of Agricultural and Food Chemistry*, 62: 12504–12511.
- Xiang C., Taylor A.G., Hinestroza J.P., Frey M.W. (2013): Controlled release of nonionic compounds from poly(lactic acid)/cellulose nanocrystal nanocomposite fibers. *Journal of Applied Polymer Science*, 127: 79–86.
- Xu H., Yan F., Monson E.E., Kopelman R. (2003): Room-temperature preparation and characterization of poly(ethylene glycol)-coated silica nanoparticles for biomedical applications. *Journal of Biomedical Materials Research*, 66: 870–879.
- Xue J., Luo Z., Li P., Ding Y., Cui Y., Wu Q. (2014): A residue-free green synergistic antifungal nanotechnology for pesticide thiram by ZnO nanoparticles. *Scientific Reports*, 4: 5408. doi: 10.1038/srep05408
- Yang F.L., Li X.G., Zhu F., Lei C.L. (2009): Structural characterization of nanoparticles loaded with garlic essential oil and their insecticidal activity against *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Journal of Agricultural and Food Chemistry*, 57: 10156–10162.
- Ye Z., Guo J., Wu D., Tan M., Xiong X., Yin Y., He G. (2015): Photo-responsive shell cross-linked micelles based on carboxymethyl chitosan and their application in controlled release of pesticide. *Carbohydrate Polymers*, 132: 520–528.
- Yin Y.H., Guo Q.M., Yun H.A.N., Wang L.J., Wan S.Q. (2012): Preparation, characterization and nematocidal activity of lansiumamide B nano-capsules. *Journal of Integrative Agriculture*, 11: 1151–1158.

<https://doi.org/10.17221/102/2020-PPS>

- Yousef N., Niloufar M., Elena P. (2019): Antipathogenic effects of emulsion and nanoemulsion of cinnamon essential oil against *Rhizopus* rot and grey mold on strawberry fruits. *Foods and Raw Materials*, 7: 210–216.
- Zainuddin N.J., Ashari S.E., Salim N., Asib N., Omar D., Lian G.E.C. (2019): Optimization and characterization of palm oil-based nanoemulsion loaded with *Parthenium hysterophorus* crude extract for natural herbicide formulation. *Journal of Oleo Science*, 68: 747–757.
- Zeng H., Li X., Zhang G., Dong J. (2008): Preparation and characterization of beta cypermethrin nanosuspensions by diluting O/W microemulsions. *Journal of Dispersion Science and Technology*, 29: 358–361.
- Zhang J., Li M., Fan T., Xu Q., Wu Y., Chen C., Huang Q. (2013): Construction of novel amphiphilic chitosan copolymer nanoparticles for chlorpyrifos delivery. *Journal of Polymer Research*, 20: 107. doi: 10.1007/s10965-013-0107-7
- Zhang W., He S., Liu Y., Geng Q., Ding G., Guo M., Deng Y., Zhu J., Li J., Cao Y. (2014): Preparation and characterization of novel functionalized prochloraz microcapsules using silica–alginate–elements as controlled release carrier materials. *ACS Applied Materials and Interfaces*, 6: 11783–11790.
- Ziaee M., Moharramipour S., Mohsenifar A. (2014): MA-chitosan nanogel loaded with *Cuminum cyminum* essential oil for efficient management of two stored product beetle pests. *Journal of Pest Science*, 87: 691–699.