

SHORT COMMUNICATION

Freezing Tolerance and Proline Content of *in vitro* Selected Hydroxyproline Resistant Winter Oilseed Rape

ANNA JANSKÁ^{1,2}, SYLVA ZELENKOVÁ¹, MIROSLAV KLÍMA², MIROSLAVA VYVADILOVÁ²
and ILJA T. PRÁŠIL²

¹Department of Plant Physiology, Faculty of Science, Charles University in Prague, Czech Republic; ²Crop Research Institute, Prague-Ruzyně, Czech Republic

Abstract: Twelve doubled haploid (DH) winter oilseed rape plants with altered levels of proline and/or freezing tolerance were obtained by *in vitro* selection for resistance to *trans*-4-hydroxy-L-proline (Hyp) in five segregating microspore populations. No significant response to selection either in proline content or in freezing tolerance, compared with the non-selected control populations, was observed. When data from all examined materials were combined, a weak correlation between proline content and freezing tolerance was observed.

Keywords: abiotic stress; *Brassica napus*; microspore culture; proline

Low temperature is a major abiotic stress factor affecting rapeseed production in the Czech Republic. To survive this stress, plants need to acquire an increased freezing tolerance. During the acclimation of the plants, extensive changes are induced in gene expression and metabolic pathways (IBA 2002; WANG *et al.* 2003).

The chilling and freezing tolerance of many plants is supported by the accumulation of osmotically active agents, called compatible solutes. Among them, one of the most widespread is the amino acid proline. Thanks to its amphiphilic nature, it is able to retain water within the cells under conditions of osmotic stress. And also to stabilize the membrane and cytoplasmic proteins, as well as the membranes themselves. The metabolism of proline *per se* is also of importance, because of the regulation of both the synthesis and the

degradation of this amino acid it comes into the cycle process between proline and its intermediates under various conditions. Proline can be a means of reducing the cytoplasm acidity, as well as being a compound that is able to scavenge reactive oxygen species. After recovering from stress, proline becomes the source of carbon, nitrogen, energy, and reducing equivalents, which makes the recovery of normal plant metabolism easier (HARE *et al.* 1998; WANG *et al.* 2003).

Selection for improved freezing tolerance with Hyp was successfully used for callus cultures of potato (VAN SWAAIJ *et al.* 1987; ANJUM & VILLIERS 1998) and for both the cell lines and the embryogenic calluses of cereals (DÖRFFLING *et al.* 1993, 1994, 1997a, b; TANTAU *et al.* 2004). Recently, MCCLINCHEY and KOTT (2008) successfully used combined UV mutagenesis and

in vitro selection in microspore cultures with several proline analogues for the improvement of freezing tolerance in Canadian spring oilseed rape. Free Hyp is a phytotoxic analogue of proline for plants. The negative effect of Hyp applied to the microspore or callus cultures can be reduced by an increased level of proline inside the cells. Thus, it is suggested that regenerants which survived on a Hyp medium, might have an elevated level of proline and thus probably also improved freezing tolerance.

The aim of our study was to obtain DH winter oilseed rape plants with increased levels of proline in leaves, and consequently with higher freezing tolerance by *in vitro* selection in microspore cultures with Hyp as the selecting agent. A protocol for *in vitro* selection in microspore cultures was established and used for the selection of Hyp resistant (HypR) regenerants in the collection of Czech breeding rapeseed materials. The determination of the proline content in the leaves of regenerated plants was carried out after two-month acclimation at 4°C, following the freezing tests. The other goal was to find the relationship between the level of proline in the leaves and the degree of freezing tolerance achieved.

Five F₁ hybrids (breeding materials of winter oilseed rape, highly embryogenically responsive in microspore culture) originated from two breeding organizations: Oseva Pro, Ltd. – Research Institute of Oilseed Crops at Opava (OP22, OP24, OP25), and Selgen, a.s. – Breeding Station at Chlumec nad Cidlinou (CH993, CH2008). Two open-pollinated breeding materials (lines) of winter rapeseed HP3 and HP8, with contrasting freezing tolerances from the Research Institute of Oilseed Crops at Opava, were used in the experiments (Table 1). Donor plants for microspore cultures were vernalized for two months (4°C ± 2°C), and then grown under a controlled environment in a growth chamber (light intensity 84 μmol/m²/s, 22/20°C day/night, and photoperiod 16/8 h).

Flower buds 2.5–3.5 mm in size were selected and collected from donor plants. The isolation and culture of microspores were carried out according to KLÍMA *et al.* (2004); however, immediately after isolation, Hyp was added to the NLN liquid medium (LICHTER 1985). The concentration of microspore suspension was adjusted to 100 000 per 1 ml of the culture medium. The concentrations of Hyp in the medium were: 0 (control), 1, 4, 10, 16, and 24mM. Two Petri dishes, each with 10 ml of the

Table 1. Characterisation of initial breeding materials used for *in vitro* selection

Donor plant	Origin (A × B)
OP22	OP-1901/02 × OP-1101/02
OP24	OP-BN-07 × OP-1101/02
OP25	OP-1611/02 × OP-1012/02
CH993	I 60/4-85 × I 77/8-193
CH2008	A 4-37 × I 61/2
HP3	frost sensitive line OP-4032
HP8	frost tolerant line OP-BN-07

A – freezing tolerant parent; B – freezing sensitive parent

microspore suspension, were prepared for each Hyp concentration and genotype. All experiments were replicated three times. For further analysis, regenerants from media with 10 and 16mM Hyp were selected, where only a few globular embryos developed to the torpedo stage. Less concentrated media appeared to be ineffective for selection (more than one third of embryos developed) and the concentration 24mM was toxic (no globular embryos developed to the torpedo stage). For the mean number of developed torpedo embryos per Petri dish after 30 days of cultivation in NLN medium see Table 2. For chromosome doubling, the plantlets from microspore-derived embryos were treated with 50 mg/l colchicine solution for 24 h, according to VYVADÍLOVÁ *et al.* (1993) before transfer to soil (horticulture substrate). Plantlets were grown in a greenhouse, and then cold acclimated at 4°C ± 2°C, 8 h light/16 h dark. After 60 days of cold acclimation (control plants and respective HypR regenerants together), leaf samples were collected for the determination of proline content and freezing tolerance.

Proline content was determined in freeze-dried leaf material, according to BATES *et al.* (1973). Calibrations were made with L-proline (Sigma-Aldrich Chemie, GmbH, Steinheim, Germany) as a standard. The concentration of proline was calculated according to the formula [(ml toluene) × (μg proline/ml)]/(g sample × 2/3) = μg proline/g dry weight. Two independent proline measurements of each variant were done. For evaluation, the *t*-test was used. The degree of freezing tolerance was determined after a laboratory freezing test via the electrolyte leakage method (according

Table 2. Mean number of developed torpedo embryos per Petri dish after 30 days of cultivation in NLN medium

Genotype	Mean number of torpedo embryos per Petri dish					
	control	1mM Hyp	4mM Hyp	10mM Hyp	16mM Hyp	24mM Hyp
OP22	226.3	205.5	169.7	7.7	1.8	0.0
OP24	255.3	220.0	138.8	15.3	2.8	0.0
OP25	106.0	75.3	57.5	3.2	1.2	0.0
CH993	80.8	59.8	48.5	8.0	2.2	0.0
CH2008	107.2	87.0	68.2	3.2	1.0	0.0
HP3	243.8	225.0	112.5	8.7	4.5	0.0
HP8	114.0	97.3	56.7	4.2	0.3	0.0

to PRÁŠIL & ZÁMEČNÍK 1998) on leaf discs after gradual freezing from -3°C to -30°C . The software package developed by JANÁČEK and PRÁŠIL (1991) was used to calculate LT_{50} values (the temperature producing freezing injury in 50%).

Proline accumulates in plants as a response to abiotic stresses such as low temperature, drought, and high salinity (HARE *et al.* 1998). From this aspect proline is known to protect plant tissues in the osmotic stress pathway. As the participation of proline in the cold stress tolerance of rapeseed is not fully understood, the goal of our study was to find out regenerants with elevated proline content (selected *in vitro* with the analogue hydroxyproline) having a higher degree of freezing tolerance.

Nine hydroxyproline-resistant (HypR) DH regenerants derived from five F_1 hybrids and three HypR DH regenerants derived from two open-pollinated breeding materials of winter oilseed rape were obtained via *in vitro* selection in the presence of Hyp. Efficient concentrations of Hyp for selection of HypR microspores were 10 and 16mM, which is in concordance with concentrations used in spring canola by McCLINCHEY and KOTT (2008). Regenerated plants were treated first with colchicine, cultivated in *ex vitro* conditions, and then evaluated for their freezing tolerance and proline content in the leaves.

Two HypR DH regenerants (CH993/3, CH2008/1) showed significantly higher amounts of proline in their leaves than did the respective control plants (i.e. regenerants without Hyp treatment) after cold acclimation (Table 3).

Many studies have shown a positive correlation between the resistance of tissue cultures to Hyp and the content of proline in their tissues, particularly in cereals (TANTAU & DÖRFFLING 1991;

DÖRFFLING *et al.* 1993, 1994, 1997 a, b; TANTAU *et al.* 2004) and in potatoes (VAN SWAAIJ *et al.* 1987; ANJUM & VILLIERS 1998). In contrast, there are also studies in which this correlation was not confirmed (e.g. in HypR cell lines of lucerne the constitutively increased proline level was not found (PETRUSA & WINICOV 1997)). In spite of this, the plants showed an increased resistance to high salinity. The difference between the control and HypR variants was the more rapid accumulation of proline in the roots of the HypR plants after exposure to high salinity. For this reason, it should be interesting to study proline accumulation dynamics in the case of selected HypR plants – not only during acclimation, but particularly after exposure to temperature shock. MAGGIO *et al.* (1997) described another mechanism in carrot cell lines, where no significant correlation between the resistance to Hyp and the proline content was observed. This could be due to some changes in the membrane transport system that could block the uptake of Hyp. The osmotolerance of the carrot cells changed with the developmental stage; therefore, plants accumulating higher amounts of proline might only be more tolerant to osmotic stress during a specific stage of development. Another possible reason is the enzymatic conversion of some Hyp to proline (VARNER 1980). In conclusion, while proline is known to protect plant tissues in osmotic stress, the pathways of proline participation in the cold stress tolerance of oilseed rape are not fully understood. Considering all these possibilities, it is clear that more comprehensive studies of proline dynamics during cold acclimation and freezing temperatures in winter oilseed rape plants are necessary.

Higher freezing tolerances, observed in four HypR plants (Table 3 – OP22/2, OP24/1, CH993/4

Table 3. Proline content and freezing resistance (LT₅₀ values) in the leaves of control and hydroxyproline resistant (HypR) oilseed rape regenerants

Regenerant		Proline (mg/g DW)		LT ₅₀ (°C)	
Control	HypR	control	HypR	control	HypR
OP22	OP22/1	69.0	57.0	-17.7	-15.8*
	OP22/2		72.9		-18.2
OP24	OP24/1	57.9	53.2	-15.0	-15.3*
OP25	OP25/1	40.1	27.2*	-15.7	-12.9*
CH993	CH993/1	36.2	30.6	-15.0	-14.3*
	CH993/2		34.5		-13.4*
	CH993/3		48.2*		-13.6*
	CH993/4		48.1		-15.8
CH2008	CH2008/1	44.6	93.7*	-15.6	-14.6*
HP3	HP3/1	35.3	52.4	-14.5	-12.9*
	HP3/2		38.2		-13.1
HP8	HP8/1	39.2	43.9	-13.7	-14.0

Significant differences between control and Hyp-resistant regenerants are marked with (*) for $P \leq 0.05$

and HP8/1), were statistically significant in only one DH regenerant (OP24/1). The freezing tolerance in the other HypR regenerants was lower than in the initial donor (mother) plants. Microspore cultures of *Brassica napus* L. are a valuable contribution to breeders, not only for the shortening of the breeding processes but also for the prospect of uncovering mutations or recessive phenotypes. Variability in freezing tolerances in the progeny of DH plants obtained from a single mother plant (OP22/1, OP22/2 and CH993/1, 2, 3, 4) was observed (Table 3).

Many previous studies that dealt with the problems of cold stress showed a positive correlation between the accumulated proline in plant tissues or cell cultures and the plant's freezing tolerance, particularly in cereals (CHAREST & PHAN 1990; DÖRFFLING *et al.* 1993, 1994, 1997a, b; CHEN & LI 2002; TANTAU *et al.* 2004). However, in yet other studies this relationship was not so explicit (e.g. VAN SWAAIJ *et al.* 1985, FAHL *et al.* 1994). Our study showed that there existed a significant correlation between the freezing tolerance and the proline content in leaves of winter oilseed rape, although selection in this case was not successful (Figure 1). However, three DH regenerants (Table 3 – OP22/2, CH993/4 and HP8/1) had both the higher levels of proline and the increased

freezing tolerance. Since we did not use any mutagenic agents, the selection of DH lines with elevated proline exploited the natural variability of F₁ hybrid progeny. Therefore, control plants with higher accumulation of proline or improved freezing tolerance were, in fact, randomly selected resistant variants of this progeny.

Two of the *in vitro* selected regenerants with increased proline content (CH993/3 and CH2008/1) had significantly lower freezing tolerance (Table 3). Therefore, it can be concluded that the *in vitro* selection method used in the chosen breeding materials did not lead to increased proline contents, combined with higher freezing tolerance. Nevertheless, it cannot be concluded that the role of proline in plant protection against low temperatures is trivial. It is possible that more important than an elevated proline level is the dynamics of its increase and/or the turnover of proline *per se*; and that by influencing the redox potential of the cell, it contributes to a decrease in oxidative injury (HARE *et al.* 1998). Although these preliminary results do not show so clear conclusions, they afford a methodical tool that could be tested by breeders in larger amounts of samples.

Acknowledgements. This work was supported by the Ministry of Agriculture of the Czech Republic,

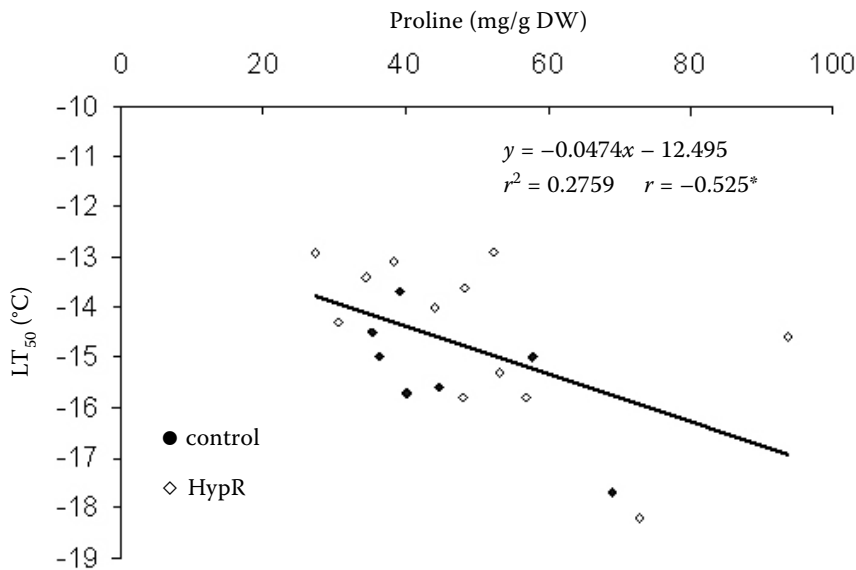


Figure 1. Correlations between proline content and freezing resistance (LT_{50} values) in the leaves of control and hydroxyproline resistant (HypR) oilseed rape regenerants

*correlation coefficient significant at $P \leq 0.05$, $n = 19$

Projects No. 0002700604 and QH 82285. We thank V. STŘESKOVÁ and P. BARTOŠOVÁ for their technical assistance.

References

- ANJUM M.A., VILLIERS T.A. (1998): Selection of hydroxyproline-resistant cell lines from *Solanum tuberosum* L. callus. I. Stability and frost tolerance. *Journal of Genetics and Breeding*, **53**: 113–117.
- BATES L.S., WALDREN R.P., TEARE I.D. (1973): Rapid determination of free proline for water-stress studies. *Plant and Soil*, **39**: 205–207.
- CHAREST CH., PHAN CH.T. (1990): Cold acclimation of wheat (*Triticum aestivum*): properties of enzymes involved in proline metabolism. *Physiologia Plantarum*, **80**: 159–168.
- CHEN W.P., LI P.H. (2002): Membrane stabilization by abscisic acid under cold aids proline in alleviating chilling injury in maize (*Zea mays* L.) cultured cells. *Plant Cell and Environment*, **25**: 955–962.
- DÖRFFLING K., DÖRFFLING H., LESSELICH G. (1993): *In vitro*-selection and regeneration of hydroxyproline-resistant lines of winter wheat with increased proline content and increased frost tolerance. *Journal of Plant Physiology*, **142**: 222–225.
- DÖRFFLING K., DÖRFFLING H., LESSELICH G., MELZ G., JÜRGENS H.U. (1994): *In vitro* selection and genetic analysis of increased frost tolerance in winter wheat. In: DÖRFFLING K., BRETTSCHEIDER B., TANTAU H., PITHAN K. (eds): *Crop Adaptation to Cool Climates*. Workshop COST 814. Hamburg, 259–264.
- DÖRFFLING K., DÖRFFLING H., LESSELICH G., LUCK E., MELZ G. (1997a): Improvement of frost tolerance in winter wheat by *in vitro* selection of proline-overproducing mutants. *Acta Agronomica Hungarica*, **45**: 295–299.
- DÖRFFLING K., DÖRFFLING H., LESSELICH G., LUCK E., ZIMMERMANN C., MELZ G., JÜRGENS H.U. (1997b): Heritable improvement of frost tolerance in winter wheat by *in vitro*-selection of hydroxyproline-resistant proline overproducing mutants. *Euphytica*, **93**: 1–10.
- FAHL E., KUNKEL S., DÖRFFLING K. (1994): Low-temperature induced changes in frost tolerance and proline content in three winter rape varieties under field and growth chamber conditions. In: DÖRFFLING K., BRETTSCHEIDER B., TANTAU H., PITHAN K. (eds): *Crop Adaptation to Cool Climates*. Workshop COST 814. Hamburg, 99–106.
- HARE P.D., CRESS W.A., VAN STADEN J. (1998): Dissecting the roles of osmolyte accumulation during stress. *Plant Cell and Environment*, **21**: 535–553.
- IBA K. (2002): Acclimative response to temperature stress in higher plants: approaches of gene engineering for temperature tolerance. *Annual Review of Plant Biology*, **53**: 225–245.
- JANÁČEK J., PRÁŠIL I. (1991): Quantification of plant frost injury by nonlinear fitting of an S-shaped function. *Cryo-Letters*, **12**: 47–52.
- KLÍMA M., VYVADILOVÁ M., KUČERA V. (2004): Production and utilization of doubled haploids in *Brassica oleracea* vegetables. *Horticultural Science (Prague)*, **31**: 119–123.
- LICHTER R. (1985): From microspores to rape plants. A tentative way to low glucosinolate strains. In: SORENSEN H. (ed.): *Advance in the Production and Utilisation of Cruciferous Crops*. Nijhoff M., Junk W. Publishers, Martinus Dordecht, Boston, Lancaster, 268–277.

- MAGGIO A., BRESSAN R.A., HASEGAWA P.M., LOCY R.D. (1997): Moderately increased constitutive proline does not alter osmotic stress tolerance. *Physiologia Plantarum*, **101**: 240–246.
- MCCLINCHEY S.L., KOTT L.S. (2008): Production of mutants with high cold tolerance in spring canola (*Brassica napus*). *Euphytica*, **162**: 51–67.
- PETRUSA L.M., WINICOV I. (1997): Proline status in salt-tolerant and salt-sensitive alfalfa cell lines and plants in response to NaCl. *Plant Physiology and Biochemistry*, **35**: 303–310.
- PRÁŠIL I., ZÁMEČNÍK J. (1998): The use of a conductivity measurement method for assessing freezing injury. I. Influence of leakage time, segment number, size and shape in a sample on evaluation of the degree of injury. *Environmental and Experimental Botany*, **40**: 1–10.
- TANTAU H., DÖRFFLING K. (1991): *In vitro*-selection of hydroxyproline-resistant cell lines of wheat (*Triticum aestivum*): accumulation of proline, decrease in osmotic potential, and increase in frost tolerance. *Physiologia Plantarum*, **82**: 243–248.
- TANTAU H., BALKO CH., BRETTSCHEIDER B., MELZ G., DÖRFFLING K. (2004): Improved frost tolerance and winter survival in winter barley (*Hordeum vulgare* L.) by *in vitro* selection of proline overaccumulating lines. *Euphytica*, **139**: 19–32.
- VAN SWAAIJ A.C., JACOBSEN E., FEENSTRA W.J. (1985): Effect of cold hardening, wilting and exogenously applied proline on leaf proline content and frost tolerance of several genotypes of *Solanum*. *Physiologia Plantarum*, **64**: 230–236.
- VAN SWAAIJ A.C., NIJDAM H., JACOBSEN E., FEENSTRA W.J. (1987): Increased frost tolerance and amino acid content in leaves, tubers and leaf callus of regenerated hydroxyproline resistant potato clones. *Euphytica*, **36**: 369–380.
- VARNER J.E. (1980): The direct conversion of hydroxyproline to proline. *Biochemical and Biophysical Research Communications*, **96**: 692–696.
- VYVADILOVÁ M., ZELENKOVÁ S., TOMÁŠKOVÁ D., KOŠNER J. (1993): Diploidization and cytological control of *Brassica napus* L. haploids. *Rostlinná Výroba*, **39**: 129–137.
- WANG W., VINOCUR B., ALTMAN A. (2003): Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta*, **218**: 1–14.

Received for publication July 13, 2009

Accepted after corrections January 3, 2010

Corresponding author:

Ing. MIROSLAVA VYVADILOVÁ, CSc., Výzkumný ústav rostlinné výroby, v.v.i., Odbor genetiky, šlechtění a kvality produkce, Drnovská 507, 161 06 Praha 6-Ruzyně, Česká republika
tel.: + 420 233 022 325; e-mail: vyvadiлова@vurv.cz
