QTL mapping of adult plant resistance to stripe rust in the Fundulea 900 × Thatcher RIL population

Xiaocui Yan[1], Huimin Zheng[1], Peipei Zhang[1], Gebrewahid Takele Weldu[1,2], Zaifeng Li[1]*, Daqun Liu[1]*

[1]*Department of Plant Pathology, College of Plant Protection, Hebei Agricultural University, Baoding, Hebei, P.R. China*
[2]*College of Agriculture, Aksum University, Shire-Indaslassie, Tigray, Ethiopia*

*Corresponding authors: lzf7551@aliyun.com; ldq@hebau.edu.cn

Abstract: Wheat stripe rust, caused by *Puccinia striiformis* Westend. f.sp. *tritici* Eriks (*Pst*), is one of the most important diseases of bread wheat worldwide. Breeding resistant wheat cultivars is the most economical, effective and environmentally friendly way for controlling wheat stripe rust in China. The Romanian wheat line Fundulea 900 showed good resistance to wheat stripe rust at the adult stage. The present study aimed to map the quantitative trait loci (QTLs) for stripe rust resistance in 176 $F_{2:6}$ recombinant inbred lines (RIL) derived from the cross of Fundulea 900 × Thatcher. The RIL population was phenotyped for stripe rust (YR) severity at Mianyang in the Sichuan province and Baoding in the Hebei province in the 2016/2017 and 2017/2018 cropping seasons. SSR markers combined with a preferred screened group (PSG) analysis were used to identify the QTLs for stripe rust in the population. Three QTLs for stripe rust resistance were mapped on chromosomes 1AL, 7BL and 7DS, respectively. All three QTLs originated from Fundulea 900 and were detected in all the environments. The QTL on 7DS was provided by the known resistance gene *Yr18/Lr34*. The two QTLs on chromosomes 1AL and 7BL were explained by 9.2 to 21.5% and 5.1 to 10.1% of the phenotypic variance, respectively and might be new QTLs. The QTLs identified in the study and their closely linked markers can be used for marker-assisted selection (MAS) in wheat breeding programmes.

Keywords: APR (adult-plant resistance); molecular mapping; *Puccinia striiformis* Westend. f.sp. *tritici* Eriks (*Pst*); QTLs; SNP array; SSR marker

Common wheat (*Triticum aestivum L.*) is widely cultivated worldwide and is one of the most important staple food crops for humankind (Haile et al. 2013). Stripe rust (YR), caused by *Puccinia striiformis* Westend. f.sp. *tritici* Eriks (*Pst*) (Westendorp 1854), is an important wheat disease worldwide (Chen 2005). Stripe rust is a significant threat in most of the wheat growing regions in the world with the potential to inflict regular regional crop losses of 5 to 25% (Wellings 2011). From the 1950s, stripe rust has occurred in at least 15 crop seasons in China, and has caused yield losses of up to 1 million tonnes in four crop seasons (Wan et al. 2007). The most effective and environmentally sound method to control this disease is widely deploying resistant cultivars with different genes. Resistance to rust is generally classified into two types, race specific and race non-specific (Johnson 1988). Race specific resistance is controlled by a single gene, and often expresses high resistance at the seedling stage. This resistance often loses effectiveness when virulent races arise. The race non-specific resistance also termed as slow

rusting resistance is effective at later plant growth stages conferring resistance by multiple genes and is more durable (Singh et al. 2005).

To date, 83 YR resistance genes have been identified and catalogued in wheat, and 327 YR quantitative trait loci (QTL) have been reported so far (Wang & Chen 2017; Li et al. 2020). Only four genes *Lr34/Yr18/Sr57/Pm38* (Singh et al. 2012), *Lr46/Yr29/Sr58/Pm39* (Singh et al. 2013), *Lr67/Yr46/Sr55/Pm46* (Herrera-Foessel et al. 2011), and *Lr68* (Herrera-Foessel et al. 2012) were reported to be slow rusting resistance genes. *Lr34* confers a moderate level of leaf rust resistance when present alone; however, combinations with additional slow rusting genes generally result in higher resistance levels (Singh et al. 2000). Therefore, mapping more, new slow-rusting genes is important for breeding cultivars with durable resistance.

Molecular markers, including restriction fragment length polymorphisms (RFLP), amplified fragment length polymorphisms (AFLP), simple sequence repeats (SSR), resistance gene analogue polymorphisms (RGAP) and single nucleotide polymorphisms (SNP), have been widely used in the genetic mapping of wheat disease resistance genes. Among them, high-density SNP arrays provide a superior approach for QTL mapping due to their advantages of higher accuracy and, particularly, higher density than other markers (Botstein et al. 1980; Leal et al. 2010; Grativol et al. 2011).

The Romanian wheat line Fundulea 900 introduced to China in 1999 was highly resistant to leaf rust and stripe rust under field conditions. In previous studies, leaf rust resistance genes were identified from the Fundulea 900 × Thatcher $F_{2:3}$ population using SSR markers (Xing et al. 2014; Zhang et al. 2017). In the present study, the RIL population from the cross of Fundulea 900 × Thatcher were used to identify the QTLs to stripe rust using SSR markers and the 55K SNP array.

MATERIAL AND METHODS

Plant materials and *Pst* pathotypes. The resistant parent Fundulea 900, the susceptible parent Thatcher, and 176 $F_{2:6}$ recombinant inbred line (RIL) populations were used to map the Adult-Plant Resistance (APR) QTL to the YR. A urediniospore mixture in equal proportion of two *Pst* pathotypes, CYR32 and CYR33, was used in the field trail. All the wheat accessions were maintained at the Biological Control Center for plant Diseases and Plant Pests of Hebei, Hebei

Agricultural University. The *Pst* pathotypes were kindly provided by the Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing.

Phenotypic characterization of stripe rust resistance in the field. A total of 176 RILs and two parents were grown at Baoding in the Hebei Province in 2016–2017 cropping season (here after referred as 2017BD) and Mianyang in the Sichuan Province in 2016–2017 and 2017–2018 cropping seasons (here after referred as 2017SC and 2018SC, respectively) for evaluating the YR severities. The field trials were conducted in randomised complete blocks with two replicates. The field plots were single 1.5 m rows sown with approximately 50 seeds and 30 cm apart between the rows. The highly susceptible line Mingxian 169 was planted every tenth row to aid the spread of the urediniospores within the trial. Spreader rows of Mingxian 169 were planted perpendicular and adjacent to the test rows. Stripe rust epidemics were initiated by spraying aqueous suspensions of urediniospores in equal proportions (50% of each pathotype) plus a few drops of Tween 20 (0.03%) onto the spreader rows at tillering. The disease severities as percentage of the leaf area covered with uredinia were scored two or three times at about one-week intervals with the first scoring 4 weeks after inoculation in each place according to the modified Cobb scale (Peterson et al. 1948). The final disease severity (FDS) was recorded for each line in each environment when the susceptible check Mingxian 169 were fully rusted. The FDS data were used for the QTL analysis. The phenotypic correlation coefficients between the FDS in the different environments were calculated by the Microsoft Excel analytical tool.

Genotyping. The genomic DNA was extracted by the cetyltrimethylammonium bromide (CTAB) method (Sharp et al. 1988) from non-infected seedling leaves of the 176 RILs and two parents. The DNA concentration was quantified with a Thermo Scientific NanoDrop 2000 (Thermo Scientific, USA) and diluted to a final concentration of 50 ng/μL prior to further analysis. Based on the FDS value, equal amounts of DNA from five typical resistant RILs (FDS < 5%) and five typical susceptible RILs (FDS > 90%), respectively, were selected to form the preferred screened group (PSG) (Hao et al. 2008). At the same time, the Affymetrix Wheat 55K SNP Array (The Chinese Academy of Agricultural Sciences, China) was used to scan the whole genome of the PSG for rapid identifying the QTLs related to the stripe rust resistance.

The DNA samples of the two parents and PSG were screened for polymorphism with SSR markers

Table 1. Final disease severity (FDS) for stripe rust of 176 recombinant inbred lines (RILs) from Fundulea 900 × Thatcher in three environments

Year and locality	Fundulea 900	Thatcher	RIL population		
			mean	minimum	maximum
YR2017BD	1	80	15.7	1	90
YR2017SC	1	80	19.0	1	90
YR2018SC	1	80	19.8	1	100

and SNP markers were used to genotype the entire population. PCR (polymerase chain reaction) for SSR markers was performed following Helguera et al. (2003). The PCRs were performed in volumes of 10 μL containing 5 μL 2 × TapPlusMasterMix (Vazyme, Nanjing, China), 1 μL ddH$_2$O, 2 μL (20 ng/μL) of primer, and 2 μL (20 ng/μL) of the template DNA. The PCR products for the SSR primers were separated in 6 or 8% polyacrylamide denaturing gels that were silver-stained for fragment detection.

QTL detection. The software map manager QTXb17 was used to calculate the linkage distance between the markers, and QTLs ICMapping 4.0 was used to construct the genetic map and analyse the QTLs (http://www.isbreeding.net/software/?type=detail&id=18) (Li et al. 2007). The logarithm of odds (LOD = 2.5) threshold to declare a QTL for each trait was based on 1 000 permutation tests. A walk speed of 1.0 cM was chosen for the QTL detections. The quantitative trait loci effects were estimated as the proportion of the phenotypic variance (R^2) explained (PVE) by the QTL. The flanking sequences of all the molecular markers were subjected to BLAST against the Chinese Spring reference sequence (https://urgi.versailles.inra.fr/blast_iwgsc/blast.php, IWGSC 2018) in order to determine the physical positions. The MapChart 2.3 (https://www.wur.nl/en/show/Mapchart.htm) was used to draw the linkage maps (Voorrips 2002).

RESULTS

Phenotyping of stripe rust responses. The FDS of the susceptible controls Mingxian 169 ranged from 80 to 100% across three environments, indicating stripe rust epidemics for the Fundulea 900 × Thatcher population for the YR developed well in each environment. The mean FDS of Fundulea 900 and Thatcher were 1 and 80%, respectively, across the three environments. The mean YR severities of the RILs ranged from 15.7 to 19.8% across all the environments and the FDS of the RIL population

across all the environments ranged from 1% to 100%, respectively (Table 1). The frequency distribution of FDS in each environment showed a continuous distribution skewed towards resistance (Figure 1), indicating polygenic inheritance. The minority lines showed a higher FDS than Thatcher, indicating transgressive segregation. The FDS in the three environments were significantly correlated with Pearson's correlation coefficients (r) ranging from 0.88 to 0.89 ($P < 0.001$) (Table 2).

Screening of molecular markers. The results based on the scanning with the Affymetrix 55K SNP Array (53 064 markers) showed that the QTLs for the stripe rust were mainly distributed on chromosomes 1A, 4A, 7B and 7D with 0–1 exchange in preferred screened group (PSG) (Figure 2). Therefore, the SSR markers, mainly from these four chromosomes, were used to test for polymorphism in the parents and the PSG. A total of 42 polymorphic SSR markers (Table S1 in the Electronic Supplementary Material (ESM)) were used to genotype the 176 RIL lines and construct the linkage map.

QTL mapping of stripe rust resistance. Three APR QTLs for the YR were detected in the population and designated as *QYr.hebau-1AL*, *QYr.hebau-7BL* and *QYr.hebau-7DS*, respectively (Table 3, Figure 3). All the resistance alleles were contributed by the resistant parent Fundulea 900.

Table 2. Pearson's correlation coefficients (r) for the two-way comparisons of the stripe rust severity data from the different environments

Environment[a]	2017BD	2017SC
2017SC	0.89**	
2018SC	0.89**	0.88**

**Significant at $P = 0.01$; a 2017BD, 2017SC, and 2018SC – the final disease severity (FDS) in the 2016/2017 cropping season in Baoding, Hebei province, and during the 2016/2017 and 2017/2018 cropping seasons in Mianyang, Sichuan province, respectively
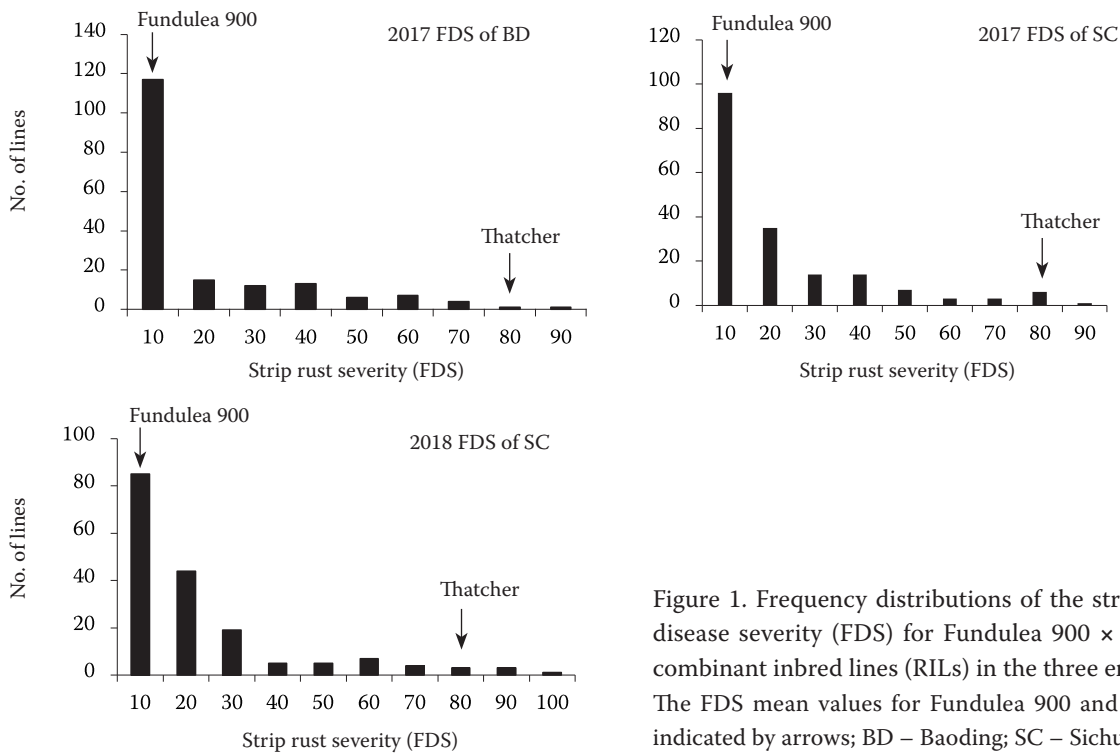
Figure 1. Frequency distributions of the stripe rust final disease severity (FDS) for Fundulea 900 × Thatcher recombinant inbred lines (RILs) in the three environments The FDS mean values for Fundulea 900 and Thatcher are indicated by arrows; BD – Baoding; SC – Sichuan

All three QTLs were stably detected in all three environments (Figure 3). The first stable QTL *QYr.hebau-1AL* flanked by markers *Xgwm164* and *Xwmc611* explained 9.2, 11.2 and 21.5% of the phenotypic variances in 2017BD, 2017SC and 2018SC, respectively. The second QTL *QYr.hebau-7BL* with a flanking marker *Xcfa2040* and *Xgwm344* accounted for 5.1, 10.1 and 6.4% of the phenotypic variance in 2017BD, 2017SC and 2018SC, respectively. *QLr. hebau-7DS* explained 9.3, 9.1 and 13.7% of the phenotypic variances in 2017BD, 2017SC and 2018SC,

respectively (Table 3). The QTL was *Yr18/Lr34* based on the marker detection with *csLV34.* The results showed at least three main QTLs with a stable effect were present in resistant cultivar Fundulea 900, and, in fact, Fundulea 900 also carried the slow rusting gene *Lr46/Yr29* based on a previous report (Zhang et al. 2017), as a result, most of the RILs showed high resistance. The lines with all the identified QTLs could be used as a resistant parent for breeding cultivars with durable resistance to stripe rust.
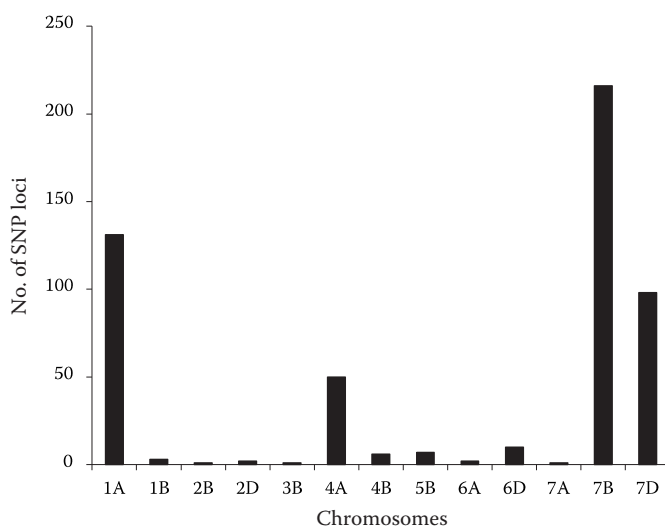

Figure 2. Frequency distributions of the single nucleotide polymorphism (SNP) loci with 0–1 exchange in the preferred screened group (PSG) on the different chromosomes

Table 3. QTLs for stripe rust identified from the Fundulea 900 × Thatcher recombinant inbred line (RIL) populations by IciMapping

QTL[a]	Year and locality	Marker interval	LOD[b]	PVE (%)[c]	Add[d]
	2017BD	gwm164-wmc611	2.8	9.2	−4.1
QYr.hebau-1AL	2017SC	gwm164-wmc611	3.4	11.2	−4.9
	2018SC	gwm164-wmc611	6.1	21.5	−6
	2017BD	cfa2040-gwm344	2.9	5.1	−3.6
QYr.hebau-7BL	2017SC	cfa2040-gwm344	3.9	10.1	−6.5
	2018SC	cfa2040-gwm344	3.3	6.4	−5.8
	2017BD	csLv34-barc92	3.7	9.3	−4.9
QYr.hebau-7DS	2017SC	csLv34-barc92	3.6	9.1	−6.1
	2018SC	csLv34-barc92	5.6	13.7	−7.3

[a]QTLs overlapping within one-log support confidence interval were assigned the same symbol; [b]logarithm of odds (LOD) score; [c]percentage of the phenotypic variance explained by the QTL; [d]additive effect of the resistance allele

DISCUSSION

QYr.hebau-1AL. In the present study, *QYr.hebau-1AL* was mapped between *Xgwm164* and *Xwmc611* on chromosome 1AL near the centromere. So far, four QTLs, *QYr.sun-1A.1* in Janz, *QYr.sun-1A.1* in Renan, *QYr.cim-1A.1* in Pastor and *QYr.caas-1A.1* in Naxos were mapped on chromosome 1AL (Rosewarne et al. 2013). *QYr.caas-1A.1* in *Naxos* was at the end of 1AL. Another three QTLs, *QYr.sun-1A.1_Janz*, *QYr.sun-1A.1_Renan* and *QYr.cim-1A.1_Pastor*, were also close to the centromere, but they had a different genetic background with *QYr.hebau-1AL*. Therefore, the relationship between *QYr.hebau-1AL* and the other reported QTLs requires further investigation.

QYr.hebau-7BL.1. Eight known YR genes or QTLs, *Yr52* in PI 183527 (Ren et al. 2012), *Yr59* in PI 178759 (Zhou et al. 2014), *Yr79* in PI182103 (Feng et al.
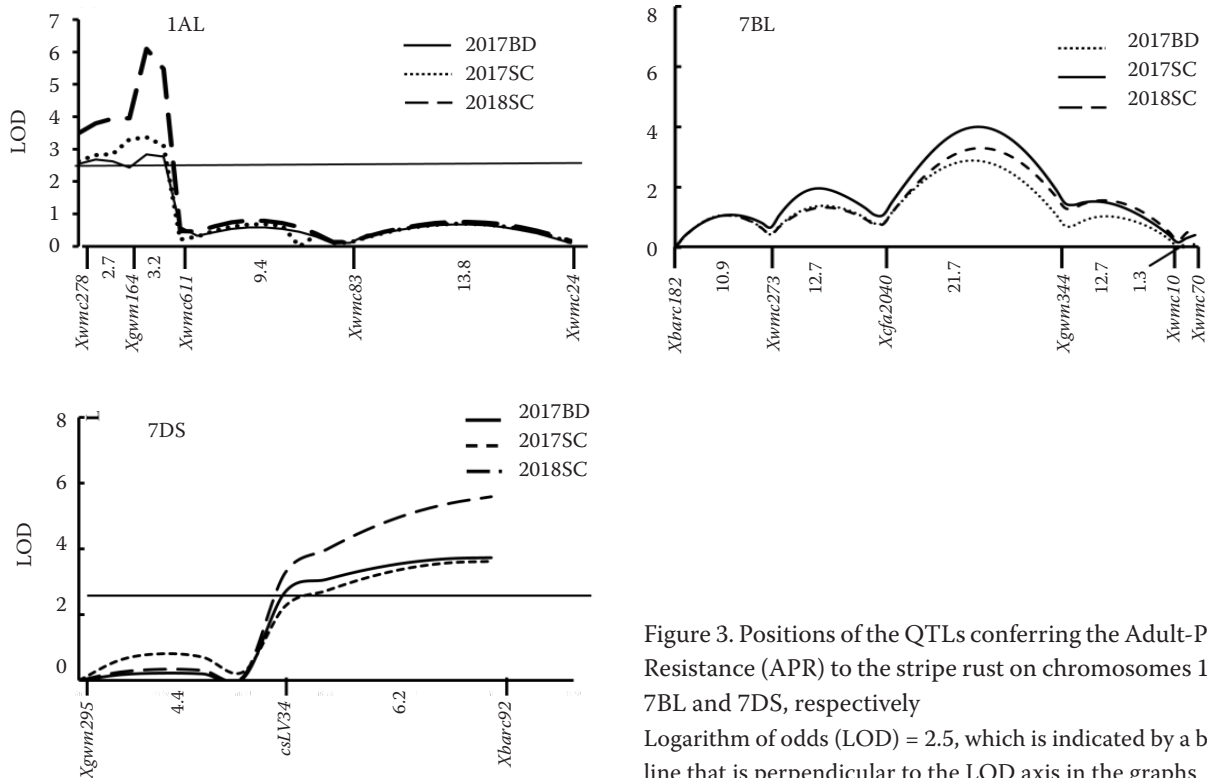

Figure 3. Positions of the QTLs conferring the Adult-Plant Resistance (APR) to the stripe rust on chromosomes 1AL, 7BL and 7DS, respectively

Logarithm of odds (LOD) = 2.5, which is indicated by a black line that is perpendicular to the LOD axis in the graphs

Table 4. Comparison of the physical positions of the QTLs identified in the present study with those reported previously

QTL	Linked markers	Physical position (Mb)[a]	Origin	Reference
QYr.sun-1A	*Xgwm164*	280.6	Janz	Bariana et al. (2010)
QYr.sun-1A	*Xfba118b*	–	Renan	Dedryver et al. (2009)
QRYr1A.1-1AL	*wPt-6005(Xgwm497)*	550.9	Pastor	Rosewarne et al. (2012)
QRYr1A.2-1AL	*Xwmc59(Xbarc213)*	572.2–575.3	Naxos	Ren et al. (2012a)
QYr.hebau-1AL.1	*Xwmc611-Xgwm164*	150.8–280.6	Fundulea 900	present study
Yr52	*Xbarc182–Xwgp5258*	732.4	PI 183527	Ren et al. (2012b)
Yr59	*Xwgp5175–Xbarc32*	723.9	PI 178759	Zhou et al. (2014)
Yr79	*Xwmc335–Xbarc72*	750.6	PI 182103	Feng et al. (2018)
YrZH84	*Xcfa2040–Xbarc32*	723.9–750.6	Zhou 8425B	Li et al. (2006)
Yr39	*Xgwm131*	604.7	Alpowa	Lin and Chen (2007)
QYr.caas-7BL.1	*XwPt-8106*	557	SHA3/CBRD	Ren et al. (2012a)
Lr68	*csGS*	734.2	Parula	Herrera-Foessel et al. (2012)
Yr67	*Xcfa2040–SC-P35M48*	699.9–718.4	C591	Li et al. (2009)
QYr.hebau-7BL	*Xwms577–Xbarc1073*	712.3–723.2	Fuyu 3	Gebrewahid et al. (2020)
QYr.hebau-7BL.1	*Xwmc273-Xcfa2040*	718.4	Fundulea 900	present study
QLr.hebau-7DS	*csLv34*	514.3	Fundulea 900	present study

– Could not be identified in the IWGSC RefSeq v1.0; [a]the physical positions of the markers were available at IWGSC RefSeq v1.0 (https://urgi.versailles.inra.fr/jbrowseiwgsc/gmod_jbrowse/)

2018), *YrZH84* in Zhou 8425B (Li et al. 2006), *Yr39* in Alpowa (Lin & Chen 2007), *Yr67* in C591 (Li et al. 2009), *QYr.caas-7BL.1* in SHA3/CBRD (Ren et al. 2012a), and *QYr.hebau-7BL* in Fuyu 3 (Gebrewahid et al. 2020), were mapped on chromosome 7BL. According to the position of the closely linked markers (Table 4), the eight genes were mapped at 732.4Mb, 723.9Mb, 750.6Mb, 723.9-750.6Mb, 604.7Mb, 699.9-718.4Mb, 557.0Mb, and 712.3-723.2Mb, respectively. In the present study, *QYr.hebau-7BL.1* was closely linked with *Xwmc273* and *Xcfa2040* was at the physical distance 718.4Mb. This QTL was mapped at a similar position with *Yr59*, *YrZH84*, *Yr67*, and *QYr. hebau-7BL*. *Yr59*, *YrZH84*, and *Yr67* were major genes, but *QYr.hebau-7BL.1* was a minor gene in the study; therefore, the three genes might be different from *QYr.hebau-7BL.1*. In addition, *QYr.hebau-7BL* in Fuyu 3 were considered to be *Lr68* (Gebrewahid et al. 2020), but the APR for leaf rust was not identified in Fundulea 900 (Zhang et al. 2017). In the previous study, a seedling gene *LrFun* from Fundulea 900 was also mapped on 7BL at a similar position as *QYr. hebau-7BL.1* (Xing et al. 2014), but the gene had lost its resistance in the field trial (Zhang et al. 2017). Therefore, it is possible that *QYr.hebau-7BL.1* was also different from *QYr.hebau-7BL* and might be a new QTL for YR.

Yr18/Lr34 and Lr46/Yr29. In the current study, *QYr.hebau-7DS* is a known race non-specific APR gene *Yr18*/*Lr34* (Table 4). In the previous study, the gene *Yr18*/*Lr34* also conferred the APR for leaf rust using the same population (Zhang et al. 2017). In fact, the other slow rusting gene *Lr46*/*Yr29* was also present in Fundulea 900 and conferred the APR for leaf rust with comparatively weak effects (Zhang et al. 2017). However, the stripe rust resistance of *Lr46*/*Yr29* was not identified in the present study. *Lr46*/*Yr29* often had a reduced effect when it was in combination with *Lr34*/*Yr18* (Suenaga et al. 2003; Lillemo et al. 2008).

CONCLUSION

In this study, three QTLs for stripe rust resistance were detected in the Fundulea 900 × Thatcher cross. The three QTLs were mapped on the 1AL, 7BL and 7DS chromosomes, respectively. All three QTLs originated from Fundulea 900 and were detected in all the environments. The QTL on 7DS was provided by the known gene *Yr18/Lr34*. The two QTLs on chromosomes 1AL and 7BL might be new QTLs. The QTLs identified in the study and their closely linked markers can be used for marker-assisted selection (MAS) in wheat breeding programmes.

REFERENCES

Bariana H.S., Bansal U.K., Schmidt A., Lehmensiek A., Kaur J., Miah H., Howes N., MCIntyre C.L. (2010): Molecular mapping of adult plant stripe rust resistance in wheat and identification of pyramided QTL genotypes. Euphytica, 176: 251–260.

Botstein D., White R.L., Skolnick M., Davis R.W. (1980): Construction of a genetic linkage map in man using restriction fragment length polymorphisms. American Journal of Human Genetics, 32: 314–331.

Chen X.M. (2005): Epidemiology and control of stripe rust (*Puccinia striiformis* f.sp. *tritici*) on wheat. Canadian Journal of Plant Pathology, 27: 314–337.

Dedryver F., Paillard S., Mallard S., Robert O., Trottet M., Nègre S., Verplancke G., Jahier J. (2009): Characterization of genetic components involved in durable resistance to stripe rust in the bread wheat 'Renan'. Phytopathology, 99: 968–973.

Feng J.Y., Wang M.N., See D.R., Chao S.M., Zheng Y.L., Chen X.M. (2018): Characterization of novel gene *Yr79* and four additional QTL for all-stage and high-temperature adult-plant resistance to stripe rust in spring wheat PI 182103. Phytopathology, 108: 737–747.

Gebrewahid T.W., Zhang P.P., Zhou Y., Yan X. C., Xia X.C., He Z.H., Liu D.Q., Li Z.F. (2020): QTL mapping of adult plant resistance to stripe rust and leaf rust in a Fuyu 3/ Zhengzhou 5389 wheat population. The Crop Journal, 8: 655–665.

Grativol C., da Fonseca Lira-Medeiros C., Hemerly A.S., Ferreira P.C.G. (2011): High efficiency and reliability of inter-simple sequence repeats (ISSR) markers for evaluation of genetic diversity in Brazilian cultivated *Jatropha curcas* L. accessions. Molecular Biology Reports, 38: 4245–4256.

Haile D., Nigussie-Dechassa R., Abdo W., Girma F. (2013): Seeding rate and genotype effects on agronomic performance and grain protein content of durum wheat (*Triticum turgidum* L. var. *durum*) in south-eastern Ethiopia. African Journal of Food, Agriculture Nutrition and Development, 13: 7693–7710.

Hao Y.F., Liu A.F., Wang Y.H., Feng D.S., Gao J.R., Li X.F., Liu S.B., Wang H.G. (2008): *Pm23*: a new allele of *Pm4* located on chromosome 2AL in wheat. Theoretical and Applied Genetics, 117: 1205–1212.

Helguera M., Khan I. A., Kolmer J., Lijavetzky D., Zhongqi I., Dubcovsky J. (2003): PCR assays for the *Lr37-Yr17-Sr38* cluster of rust resistance genes and their use. Crop Science, 43: 1839–1847.

Herrera-Foessel S.A., Lagudah E.S., Huerta-Espino J., Hayden M.J., Bariana H.S., Singh D., Singh R.P. (2011): New slow-rusting leaf rust and stripe rust resistance genes *Lr67* and *Yr46* in wheat are pleiotropic or closely linked. Theoretical and Applied Genetics, 122: 239–249.

Herrera-Foessel S.A., Singh R.P., Huerta-Espino J., Rosewarne G.M., Periyannan S.K., Viccars L., Calvo-Salazar V., Lan X.C., Lagudah E.S. (2012): *Lr68*: a new gene conferring slow rusting resistance to leaf rust in wheat. Theoretical and Applied Genetics, 124: 1475–1486.

Johnson R. (1988): Durable resistance to yellow (stripe) rust in wheat and its implications in plant breeding. In: Simmonds N.W., Rajaram S. (eds.): Breeding Strategies for Resistance to the Rusts of Wheat. CIMMYT, Mexico.

Leal A.A., Mangolin C.A., do Amaral Júnior A.T., Gonçalves L.S.A., Scapim C.A., Mott A.S., Eloi I.B.O., Cordovés V., Da Silva M.F.P. (2010): Efficiency of RAPD versus SSR markers for determining genetic diversity among popcorn lines. Genetics and Molecular Research: GMR, 9: 9–18.

Li H.H., Ye G.Y., Wang J.K. (2007): A modified algorithm for the improvement of composite interval mapping. Genetics, 175: 361–374.

Li J., Dundas I., Dong C., Li G., Trethowan R., Yang Z., Hoxha S., Zhang P. (2020) Identification and characterization of a new stripe rust resistance gene *Yr83* on rye chromosome 6R in wheat. Theoretical and Applied Genetics, 133: 1095–1107.

Li Y., Niu Y.C., Chen X.M. (2009): Mapping a stripe rust resistance gene *YrC591* in wheat variety C591 with SSR and AFLP markers. Theoretical and Applied Genetics, 118: 339–346.

Li Z.F., Zheng T.C., He Z.H., Li G.Q., Xu S.C., Li X.P., Yang G.Y., Singh R.P., Xia X.C. (2006): Molecular tagging of stripe rust resistance gene *YrZH84* in Chinese wheat line Zhou 8425B. Theoretical and Applied Genetics, 112: 1098–1103.

Lillemo M., Asalf B., Singh R.P., Huerta-Espino J., Chen X.M., He Z.H., Bjørnstad Å. (2008): The adult plant rust resistance loci *Lr34/Yr18* and *Lr46/Yr29* are important determinants of partial resistance to powdery mildew in bread wheat line Saar. Theoretical and Applied Genetics, 116: 1155–1166.

Lin F., Chen X.M. (2007): Genetics and molecular mapping of genes for race-specific all-stage resistance and non-race-specific high-temperature adult-plant resistance to stripe rust in spring wheat cultivar Alpowa. Theoretical and Applied Genetics, 114: 1277–1287.

Peterson R.F., Campbell A.B., Hannah A.E. (1948): A diagram-matic scale for rust intensity on leaves and stems of cereals. Canadian Journal of Research, 26: 496–500.

Ren Y., He Z.H., Li J., Lillemo M., Wu L., Bai B., Lu Q.X., Zhu H.Z., Zhou G., Du J.Y., Lu Q.L., Xia X.C. (2012a): QTL mapping of adult-plant resistance to stripe rust

in a population derived from common wheat cultivars Naxos and Shanghai 3/Catbird. Theoretical and Applied Genetics, 125: 1211–1221.

Ren R.S., Wang M.N., Chen X.M., Zhang Z.J. (2012b): Characterization and molecular mapping of *Yr52* for high-temperature adult-plant resistance to stripe rust in spring wheat germplasm PI183527. Theoretical and Applied Genetics, 125: 847–857.

Rosewarne G.M., Singh R.P., Huerta-Espino J., Herrera-Foessel S.A., Forrest K.L., Hayden M.J., Rebetzke G.J. (2012): Analysis of leaf and stripe rust severities reveals pathotype changes and multiple minor QTLs associated with resistance in an Avocet × Pastor wheat population. Theoretical and Applied Genetics, 124: 1283–1294.

Rosewarne G.M., Herrera-Foessel S.A., Singh R.P., Huerta-Espino J., Lan C.X., He Z.H. (2013): Quantitative trait loci of stripe rust resistance in wheat. Theoretical and Applied Genetics, 126: 2427–2449.

Sharp P.J., Kreis M., Shewry P.R., Gale M.D. (1988): Location of b-amylase sequence in wheat and its relatives. Theoretical and Applied Genetics, 75: 286–290.

Singh R.P., Huerta-Espino J., Rajaram S., Barna B., Kiraly Z. (2000): Achieving near-immunity to leaf and stripe rusts in wheat by combining slow rusting resistance genes. Acta Phytopathologica et Entomologica Hungarica, 35: 133–139.

Singh R.P., Huerta-Espino J., William H.M. (2005): Genetics and breeding for durable resistance to leaf and stripe rusts in wheat. Turkish Journal of Agriculture and Forestry, 29: 121–127.

Singh R.P., Herrera-Foessel S.A., Huerta-Espino J., Bariana H., Bansal U., McCallum B., Hiebert C., Bhavani S., Singh S., Lan C.X., Lagudah E.S. (2012): *Lr34/Yr18/Sr57/Pm38/Bdv1/Ltn1* confers slow rusting, adult plant resistance to *Puccinia graminis tritici*. In: Chen W.-Q. (ed.): Proc. 13th Cereal Rusts and Powdery Mildews Conf., Beijing, August 28–September 1, 2012.

Singh R.P., Herrera-Foessel S.A., Huerta-Espino J., Lan C.X., Basnet B.R., Bhavani S., Lagudah E.S. (2013): Pleiotropic gene *Lr46/Yr29/Pm39/Ltn2* confers slow rusting, adult plant resistance to wheat stem rust fungus. In: Proc. Borlaug Global Rust Initiative, 2013, Technical Workshop, New Dehli, Aug 19–22, 2013: 17.1.

Suenaga K., Singh R.P., Huerta-Espino J., William H.M. (2003): Microsatellite markers for genes *Lr34/Yr18* and other quantitative trait loci for leaf rust and stripe rust resistance in bread wheat. Phytopathology, 93: 881–890.

Voorrips R.E. (2002): MapChart: software for the graphical presen-tation of linkage maps and QTLs. Heredity, 93: 77–78.

Wan A.M., Chen X.M., He Z.H. (2007): Wheat stripe rust in China. Crop and Pasture Science, 58: 605–619.

Wang M.N., Chen X.M. (2017): Stripe rust resistance. In: Chen X.M., Kang Z.S. (eds.): Stripe Rust. Dordrecht, Springer.

Wellings C.R. (2011): Global status of stripe rust: A review of historical and current threats. Euphytica, 179: 129–141.

Westendorp G.D. (1854): Quatrième notice surquelques Cryptogames récemment découvertes en Belgique Bull. Académeie Royale de Belgique Bulletin de la Classe des Sciences, 21: 229–246.

Xing L.F., Wang C.F., Xia X.C., He Z.H., Chen W.Q., Liu T.G., Li Z.F., Liu D.Q. (2014): Molecular mapping of leaf rust resistance gene *LrFun* in Romanian wheat line Fundulea 900. Molecular Breeding, 33: 931–937.

Zhang P.P., Qi A. , Zhou Y., Xia X.C., He Z.H., Li Z.F., Liu D.Q. (2017) Quantitative trait loci mapping of adult-plant resistance to leaf rust in a Fundulea 900×'Thatcher'wheat cross. Plant Breeding, 136: 1–7.

Zhou X.L., Wang M.N., Chen X.M., Lu Y., Kang Z.S., Jing J.X. (2014): Identification of *Yr59* conferring high-temperature adult-plant resistance to stripe rust in wheat germplasm PI 178759. Theoretical and Applied Genetics, 127: 935–945.